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(54) METHODS FOR TREATING PATIENTS WITH COMPLEMENT-ASSOCIATED DISORDERS WITH HIGH CONCENTRATION FORMULATIONS OF ANTI-C5 ANTIBODIES

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CPC C07K 16/18 (2013.01); A61K 39/3955 (2013.01); A61K 39/39591 (2013.01); A61K 2039/505 (2013.01); A61K 2039/54 (2013.01); C07K 2317/24 (2013.01); C07K 2317/94 (2013.01)

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See application file for complete search history.

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(57) ABSTRACT

The present disclosure relates to, inter alia, stable aqueous solutions comprising a high concentration of an antibody that binds to human complement component C5 and methods for preparing the solutions. The disclosure also provides methods for treating or preventing complement-associated disorders (for example, age-related macular degeneration or rheumatoid arthritis) using the solutions. Also featured are therapeutic kits containing one or more of the solutions and a means for administering the solutions to a patient in need such a treatment.

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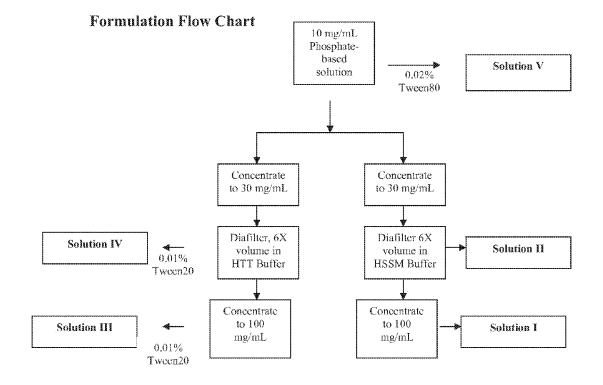
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METHODS FOR TREATING PATIENTS WITH COMPLEMENT-ASSOCIATED DISORDERS WITH HIGH CONCENTRATION FORMULATIONS OF ANTI-C5 ANTIBODIES

RELATED APPLICATIONS

This application is a divisional of U.S. application Ser. No. 13/413,268, filed Mar. 6, 2012, which claims the benefit of priority to U.S. Provisional Application Ser. No. 61/450,334, filed Mar. 8, 2011. The specifications of each of the foregoing applications are incorporated herein by reference in their entirety.

SEQUENCE LISTING

The instant application contains a Sequence Listing which has been submitted via EFS-Web and is hereby incorporated by reference in its entirety. Said ASCII copy, created on Apr. 12, 2016, is named AXJ_160CP2DV_SL.txt, and is 56,153 bytes in size.

C5 convertase. See, e.g., Schrei Acad Sci USA 75: 3948-3952 an Natl Acad Sci USA 77: 559-562. The CP C3 convertase is feet and the contained at the

TECHNICAL FIELD

The field of the invention is medicine, immunology, ²⁵ molecular biology, and protein chemistry.

BACKGROUND

The complement system acts in conjunction with other immunological systems of the body to defend against intrusion of cellular and viral pathogens. There are at least 25 complement proteins, which are found as a complex collection of plasma proteins and membrane cofactors. The plasma proteins make up about 10% of the globulins in vertebrate 35 serum. Complement components achieve their immune defensive functions by interacting in a series of intricate but precise enzymatic cleavage and membrane binding events. The resulting complement cascade leads to the production of products with opsonic, immunoregulatory, and lytic functions. A concise summary of the biologic activities associated with complement activation is provided, for example, in The Merck Manual, 16^{th} Edition.

The complement cascade can progress via the classical pathway (CP), the lectin pathway, or the alternative pathway 45 (AP). The lectin pathway is typically initiated with binding of mannose-binding lectin (MBL) to high mannose substrates. The AP can be antibody independent, and can be initiated by certain molecules on pathogen surfaces. The CP is typically initiated by antibody recognition of, and binding to, an antigenic site on a target cell. These pathways converge at the C3 convertase—the point where complement component C3 is cleaved by an active protease to yield C3a and C3b.

The AP C3 convertase is initiated by the spontaneous hydrolysis of complement component C3, which is abundant 55 in the plasma in the blood. This process, also known as "tickover," occurs through the spontaneous cleavage of a thioester bond in C3 to form C3i or C3(H₂O). Tickover is facilitated by the presence of surfaces that support the binding of activated C3 and/or have neutral or positive charge characteristics (e.g., bacterial cell surfaces). This formation of C3(H₂O) allows for the binding of plasma protein Factor B, which in turn allows Factor D to cleave Factor B into Ba and Bb. The Bb fragment remains bound to C3 to form a complex containing C3(H₂O)Bb—the "fluid-phase" or "initiation" C3 65 convertase. Although only produced in small amounts, the fluid-phase C3 convertase can cleave multiple C3 proteins

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into C3a and C3b and results in the generation of C3b and its subsequent covalent binding to a surface (e.g., a bacterial surface). Factor B bound to the surface-bound C3b is cleaved by Factor D to thus form the surface-bound AP C3 convertase complex containing C3b,Bb. (See, e.g., Müller-Eberhard (1988) *Ann Rev Biochem* 57:321-347.)

The AP C5 convertase—(C3b)₂,Bb—is formed upon addition of a second C3b monomer to the AP C3 convertase. (See, e.g., Medicus et al. (1976) *J Exp Med* 144:1076-1093 and Fearon et al. (1975) *J Exp Med* 142:856-863.) The role of the second C3b molecule is to bind C5 and present it for cleavage by Bb. (See, e.g., Isenman et al. (1980) *J Immunol* 124:326-331.) The AP C3 and C5 convertases are stabilized by the addition of the trimeric protein properdin as described in, e.g., Medicus et al. (1976), supra. However, properdin binding is not required to form a functioning alternative pathway C3 or C5 convertase. See, e.g., Schreiber et al. (1978) *Proc Natl Acad Sci USA* 75: 3948-3952 and Sissons et al. (1980) *Proc Natl Acad Sci USA* 77: 559-562.

The CP C3 convertase is formed upon interaction of complement component C1, which is a complex of C1q, C1r, and C1s, with an antibody that is bound to a target antigen (e.g., a microbial antigen). The binding of the C1q portion of C1 to the antibody-antigen complex causes a conformational change in C1 that activates C1r. Active C1r then cleaves the C1-associated C1s to thereby generate an active serine protease. Active C1s cleaves complement component C4 into C4b and C4a. Like C3b, the newly generated C4b fragment contains a highly reactive thiol that readily forms amide or ester bonds with suitable molecules on a target surface (e.g., a microbial cell surface). C1s also cleaves complement component C2 into C2b and C2a. The complex formed by C4b and C2a is the CP C3 convertase, which is capable of processing C3 into C3a and C3b. The CP C5 convertase—C4b, C2a,C3b—is formed upon addition of a C3b monomer to the CP C3 convertase. See, e.g., Miller-Eberhard (1988), supra and Cooper et al. (1970) J Exp Med 132:775-793.

In addition to its role in C3 and C5 convertases, C3b also functions as an opsonin through its interaction with complement receptors present on the surfaces of antigen-presenting cells such as macrophages and dendritic cells. The opsonic function of C3b is generally considered to be one of the most important anti-infective functions of the complement system. Patients with genetic lesions that block C3b function are prone to infection by a broad variety of pathogenic organisms, while patients with lesions later in the complement cascade sequence, i.e., patients with lesions that block C5 functions, are found to be more prone only to *Neisseria* infection, and then only somewhat more prone.

The AP and CP C5 convertases cleave C5, which is a 190 kDa beta globulin found in normal human serum at approximately 75 μg/ml (0.4 M). C5 is glycosylated, with about 1.5-3 percent of its mass attributed to carbohydrate. Mature C5 is a heterodimer of a 999 amino acid 115 kDa alpha chain that is disulfide linked to a 655 amino acid 75 kDa beta chain. C5 is synthesized as a single chain precursor protein product of a single copy gene (Haviland et al. (1991) *J Immunol* 146:362-368). The cDNA sequence of the transcript of this gene predicts a secreted pro-C5 precursor of 1658 amino acids along with an 18 amino acid leader sequence (see, e.g., U.S. Pat. No. 6 355.245).

The pro-C5 precursor is cleaved after amino acids 655 and 659, to yield the beta chain as an amino terminal fragment (amino acid residues+1 to 655 of the above sequence) and the alpha chain as a carboxyl terminal fragment (amino acid

residues 660 to 1658 of the above sequence), with four amino acids (amino acid residues 656-659 of the above sequence) deleted between the two.

C5a is cleaved from the alpha chain of C5 by either alternative or classical C5 convertase as an amino terminal frag- 5 ment comprising the first 74 amino acids of the alpha chain (i.e., amino acid residues 660-733 of the above sequence). Approximately 20 percent of the 11 kDa mass of C5a is attributed to carbohydrate. The cleavage site for convertase action is at, or immediately adjacent to, amino acid residue 733 of the above sequence. A compound that would bind at, or adjacent, to this cleavage site would have the potential to block access of the C5 convertase enzymes to the cleavage site and thereby act as a complement inhibitor. A compound that binds to C5 at a site distal to the cleavage site could also 15 have the potential to block C5 cleavage, for example, by way of steric hindrance-mediated inhibition of the interaction between C5 and the C5 convertase. A compound, in a mechanism of action consistent with that of the tick saliva complement inhibitor OmCI, may also prevent C5 cleavage by 20 reducing flexibility of the C345C domain of the alpha chain of C5, which reduces access of the C5 convertase to the cleavage site of C5. See, e.g., Fredslund et al. (2008) Nat Immunol 9(7):753-760.

C5 can also be activated by means other than C5 convertase 25 activity. Limited trypsin digestion (see, e.g., Minta and Man (1997) *J Immunol* 119:1597-1602 and Wetsel and Kolb (1982) *J Immunol* 128:2209-2216) and acid treatment (Yamamoto and Gewurz (1978) *J Immunol* 120:2008 and Damerau et al. (1989) *Molec Immunol* 26:1133-1142) can also cleave 30 C5 and produce active C5b.

Cleavage of C5 releases C5a, a potent anaphylatoxin and chemotactic factor, and leads to the formation of the lytic terminal complement complex, C5b-9. C5a and C5b-9 also have pleiotropic cell activating properties, by amplifying the 35 release of downstream inflammatory factors, such as hydrolytic enzymes, reactive oxygen species, arachidonic acid metabolites and various cytokines.

The first step in the formation of the terminal complement complex involves the combination of C5b with C6, C7, and 40 C8 to form the C5b-8 complex at the surface of the target cell. Upon the binding of the C5b-8 complex with several C9 molecules, the membrane attack complex (MAC, C5b-9, terminal complement complex-TCC) is formed. When sufficient numbers of MACs insert into target cell membranes the openings they create (MAC pores) mediate rapid osmotic lysis of the target cells. Lower, non-lytic concentrations of MACs can produce other effects. In particular, membrane insertion of small numbers of the C5b-9 complexes into endothelial cells and platelets can cause deleterious cell activation. In some cases activation may precede cell lysis.

As mentioned above, C3a and C5a are anaphylatoxins. These activated complement components can trigger mast cell degranulation, which releases histamine from basophils and mast cells, and other mediators of inflammation, resulting in smooth muscle contraction, increased vascular permeability, leukocyte activation, and other inflammatory phenomena including cellular proliferation resulting in hypercellularity. C5a also functions as a chemotactic peptide that serves to attract pro-inflammatory granulocytes to the site of complement activation.

C5a receptors are found on the surfaces of bronchial and alveolar epithelial cells and bronchial smooth muscle cells. C5a receptors have also been found on eosinophils, mast cells, monocytes, neutrophils, and activated lymphocytes.

While a properly functioning complement system provides a robust defense against infecting microbes, inappropriate 4

regulation or activation of complement has been implicated in the pathogenesis of a variety of disorders including, e.g., rheumatoid arthritis (RA); lupus nephritis; asthma; ischemiareperfusion injury; atypical hemolytic uremic syndrome (aHUS); dense deposit disease (DDD); paroxysmal nocturnal hemoglobinuria (PNH); macular degeneration (e.g., age-related macular degeneration (AMD)); hemolysis, elevated liver enzymes, and low platelets (HELLP) syndrome; thrombotic thrombocytopenic purpura (TTP); spontaneous fetal loss; Pauci-immune vasculitis; epidermolysis bullosa; recurrent fetal loss; multiple sclerosis (MS); traumatic brain injury; and injury resulting from myocardial infarction, cardiopulmonary bypass and hemodialysis. (See, e.g., Holers et al. (2008) Immunological Reviews 223:300-316.) Inhibition of complement (e.g., inhibition of: terminal complement formation, C5 cleavage, or complement activation) has been demonstrated to be effective in treating several complementassociated disorders both in animal models and in humans. See, e.g., Rother et al. (2007) Nature Biotechnology 25(11): 1256-1264; Wang et al. (1996) Proc Natl Acad Sci USA 93:8563-8568; Wang et al. (1995) Proc Natl Acad Sci USA 92:8955-8959; Rinder et al. (1995) J Clin Invest 96:1564-1572; Kroshus et al. (1995) Transplantation 60:1194-1202; Homeister et al. (1993) J Immunol 150:1055-1064; Weisman et al. (1990) Science 249:146-151; Amsterdam et al. (1995) Am J Physiol 268:H448-H457; and Rabinovici et al. (1992) J Immunol 149:1744 1750.

SUMMARY

This disclosure relates to stable, highly-concentrated liquid formulations of antibodies as well as methods for making and using the formulations. The disclosure provides, among other things, formulation conditions suitable for maintaining over considerable time the physical and functional stability of an anti-C5 antibody (e.g., eculizumab) in high concentration solutions. For example, the disclosure provides formulation conditions capable of maintaining an anti-C5 antibody in predominantly monomeric form for up to 2 years at 2° C. to 8° C., even when the antibody is maintained in solutions at concentrations of approximately 100 mg/mL. In addition, as described herein and exemplified in the working examples, such formulations also minimize aggregation, fragmentation, or degradation of an anti-C5 antibody within the highlyconcentrated solutions. For example, the disclosure provides formulation conditions capable of maintaining for two years an anti-C5 antibody in a highly-concentrated form with no detectable antibody fragmentation or degradation products (as determined using size exclusion chromatography-high performance liquid chromatography (SEC-HPLC)) and no more than 2% aggregate. Also provided herein are conditions suitable for formulating solutions of an anti-C5 antibody such as eculizumab at greater than 200 mg/mL.

The benefits of stable, highly-concentrated aqueous solutions of an anti-C5 antibody are numerous. First, for therapeutic applications which require the antibody to be administered to a patient in a small volume, therapeutic efficacy often turns on the amount of antibody that can be administered in that small volume. In the absence of the ability to formulate an anti-C5 antibody to high concentrations, use of, for example, subcutaneous, intravitreal, and/or intraarticular delivery routes would often be precluded. Relatedly, highly-concentrated antibody formulations allow for more patient choice regarding the route of administration. For therapeutic applications that require frequent and/or chronic administration, self-delivery or -administration is made possible by high concentration formulations and can be more appealing to

patients than intravenous infusion. For example, high concentration formulations of an anti-C5 antibody can allow a patient to self-administer the antibody by, e.g., subcutaneous injection. Therefore, the ability to formulate the antibody at high concentrations can increase compliance of administration by providing an easy home administration alternative to patients with complement-associated disorders.

Furthermore, methods for producing the aqueous solutions described herein do not require a lyophilization step, nor do the featured high concentration aqueous solutions need to be reconstituted from lyophilized material. The instantly featured high concentration antibody solutions provide several advantages over reconstituted lyophilized antibody formulations. First, medical practitioners must locally reconstitute lyophilized antibody solutions aseptically, which increases the opportunity for microbial contamination of the solution prior to administration. In addition, reconstitution requires considerable care to be certain that all of the solids contained in the reconstitution vessel are properly dissolved in solution. The high concentration agueous solutions provided herein 20 herein are sterile solutions. thus provide the medical practitioner, caregiver, and/or patient with a fast, easy, safe, and efficient means for delivering a therapeutic antibody to a patient in need thereof.

Other benefits of high concentration formulations include, e.g., manufacturing cost savings from decreasing bulk storage space and/or the number of product fills. In addition, the ability to produce a product having a longer shelf-life will ultimately require fewer production runs, which ultimately reduces cost for the manufacturer and consumer of the highly-concentrated therapeutic antibody.

In one aspect, the disclosure features an aqueous solution comprising an anti-C5 antibody at a concentration of 40 mg/mL to 200 mg/mL. In another aspect, the disclosure features an aqueous solution comprising an anti-C5 antibody at a concentration of greater than 200 mg/mL. Additional exemplary concentrations, including fixed concentrations as well as exemplary ranges of concentrations, are provided herein.

In some embodiments, any of the solutions described herein comprise at least one buffering agent at a concentration of 10 mM to 300 mM, inclusive. In some embodiments, any 40 of the solutions described herein comprise at least one buffering agent at a concentration of 10 mM to 200 mM, inclusive. In some embodiments, the at least one buffering agent is present in the solution at a concentration of at least, or equal to, 20 mM. In some embodiments, the at least one buffering 45 agent is present in the solution at a concentration of at least, or equal to, 50 mM. In some embodiments, the at least one buffering agent is an amino acid. The amino acid can be, e.g., one selected from the group consisting of histidine (e.g., L-histidine), serine (e.g., L-serine), and glycine (e.g., L-gly-50 cine). In some embodiments, any of the solutions described herein comprise two or more buffering agents. The two or more buffering agents can be, e.g., histidine and serine. In some embodiments, the two or more buffering agents are histidine and glycine.

In some embodiments, any of the solutions described herein comprise at least one carbohydrate excipient at a concentration of 0.1 to 5%. In some embodiments, the at least one carbohydrate excipient is present in the solution at a concentration of at least, or equal to, 1.5%. In some embodiments, 60 the at least one carbohydrate excipient is present in the solution at a concentration of at least, or equal to, 3%. The at least one carbohydrate excipient can be, e.g., one selected from the group consisting of sorbitol and mannitol. In some embodiments, any of the solutions described herein comprise two or 65 more carbohydrate excipients. At least two of the excipients can be, e.g., sorbitol and mannitol.

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In some embodiments, any of the solutions described herein comprise a formulation that comprises, or consists of, the following composition: (i) at least 20 mM histidine; at least 50 mM glycine; at least 3% (w/v) sorbitol; and at least 1.5% (w/v) mannitol; (ii) 20 mM histidine; 50 mM glycine; 3% (w/v) sorbitol; and 1.5% (w/v) mannitol; (iii) at least 20 mM histidine; at least 50 mM serine; at least 3% (w/v) sorbitol; and at least 1.5% (w/v) mannitol; or (iv) at least 20 mM histidine; at least 50 mM serine; at least 2.5% (w/v) sorbitol; and at least 1.5% (w/v) mannitol. Additional exemplary formulations are set forth herein.

In some embodiments, any of the solutions described herein comprise a surfactant. The surfactant can be, e.g., polysorbate 20 or polysorbate 80. The concentration of the surfactant in the solution can be, e.g., between 0.001% to 0.02%, inclusive.

In some embodiments, any of the solutions described herein can have a pH between 6.5 and 7.5.

In some embodiments, any of the solutions described herein are sterile solutions.

In some embodiments of any of the solutions described herein, the anti-C5 antibody is eculizumab.

In some embodiments of any of the solutions described herein, the anti-C5 antibody remains at least 95 (e.g., at least 96, 97, 98, or 99) % monomeric during storage at 2° C. to 8° C. for at least six months as determined by SEC-HPLC. In some embodiments of any of the solutions described herein, the anti-C5 antibody remains at least 95 (e.g., at least 96, 97, 98, or 99) % monomeric during storage at 2° C. to 8° C. for at least one year as determined by SEC-HPLC. In some embodiments of any of the solutions described herein, the anti-C5 antibody remains at least 95 (e.g., at least 96, 97, 98, or 99) %monomeric during storage at 2° C. to 8° C. for at least six months as determined by SEC-HPLC. In some embodiments of any of the solutions described herein, the anti-C5 antibody remains at least 95 (e.g., at least 96, 97, 98, or 99) % monomeric during storage at 2° C. to 8° C. for at least one year as determined by SEC-HPLC. In some embodiments of any of the solutions described herein, the anti-C5 antibody remains at least 95 (e.g., at least 96, 97, 98, or 99) % monomeric during storage at 2° C. to 8° C. for at least 18 months as determined by SEC-HPLC. In some embodiments of any of the solutions described herein, the anti-C5 antibody remains at least 95 (e.g., at least 96, 97, 98, or 99) % monomeric during storage at 2° C. to 8° C. for at least two years as determined by SEC-HPLC.

In some embodiments of any of the solutions described herein, less than 2% of the anti-C5 antibody in the solution is aggregated as determined by SEC-HPLC. In some embodiments of any of the solutions described herein, less than 1% of the anti-C5 antibody in the solution is aggregated as determined by SEC-HPLC.

In some embodiments of any of the solutions described herein, less than 1% of the anti-C5 antibody in the solution is fragmented as determined by SEC-HPLC. In some embodiments of any of the solutions described herein, less than 0.5% of the anti-C5 antibody in the solution is fragmented as determined by SEC-HPLC. In some embodiments of any of the solutions described herein, during storage at 2° C. to 8° C. for at least six months the anti-C5 antibody retains at least 80 (e.g., at least 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, or 99) % of its C5-binding activity, as compared to a reference anti-C5 antibody corresponding to the anti-C5 antibody prior to storage. In some embodiments of any of the solutions described herein, during storage at 2° C. to 8° C. for at least one year the anti-C5 antibody retains at least 80 (e.g., at least 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91,

92, 93, 94, 95, 96, 97, 98, or 99) % of its C5-binding activity, as compared to a reference anti-C5 antibody corresponding to the anti-C5 antibody prior to storage. In some embodiments of any of the solutions described herein, during storage at 2° C. to 8° C. for at least 18 months the anti-C5 antibody retains 5 at least 80 (e.g., at least 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, or 99) % of its C5-binding activity, as compared to a reference anti-C5 antibody corresponding to the anti-C5 antibody prior to storage. In some embodiments of any of the solutions described herein, during 10 storage at 2° C. to 8° C. for at least two years the anti-C5 antibody retains at least 80 (e.g., at least 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, or 99) % of its C5-binding activity, as compared to a reference anti-C5 antibody corresponding to the anti-C5 antibody prior to storage. 15 In some embodiments of any of the solutions described herein, during storage at 2° C. to 8° C. for at least six months the anti-C5 antibody retains at least 80 (e.g., at least 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, or 99) % of its ability to inhibit hemolysis, as compared to a 20 reference anti-C5 antibody corresponding to the anti-C5 antibody prior to storage. In some embodiments of any of the solutions described herein, during storage at 2° C. to 8° C. for at least one year the anti-C5 antibody retains at least 80 (e.g., at least 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 25 96, 97, 98, or 99) % of its ability to inhibit hemolysis, as compared to a reference anti-C5 antibody corresponding to the anti-C5 antibody prior to storage. In some embodiments of any of the solutions described herein, during storage at 2° C. to 8° C. for at least 18 months the anti-C5 antibody retains 30 at least 80 (e.g., at least 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, or 99) % of its ability to inhibit hemolysis, as compared to a reference anti-C5 antibody corresponding to the anti-C5 antibody prior to storage. In some embodiments of any of the solutions described herein, during 35 storage at 2° C. to 8° C. for at least two years the anti-C5 antibody retains at least 80 (e.g., at least 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, or 99) % of its ability to inhibit hemolysis, as compared to a reference anti-C5 antibody corresponding to the anti-C5 antibody prior to 40 storage.

In another aspect, the disclosure features an aqueous solution comprising an anti-C5 antibody at a concentration of 100 ± 20 (e.g., 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 101, 102, 103, 104, 105, 106, 45, 107, 108, 109, 110, 111, 112, 113, 114, 115, 116, 117, 118, 119, or 120) mg/mL; 20 ± 5 (e.g., 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25) mM L-histidine; 50 ± 15 (e.g., 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, or 65) mM L-serine; 3 ± 1 (e.g., 50, 2, 2.1, 2.2, 2.3, 2.4, 2.5, 2.6, 2.7, 2.8, 2.9, 3, 3.1, 3.2, 3.3, 3.4, 3.5, 3.6, 3.7, 3.8, 3.9, or 4) % sorbitol; and 1.5 ± 0.5 (e.g., 1, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, or 2) % mannitol, wherein the solution has a pH of 7.1 ± 0.5 (e.g., 6.6, 6.7, 6.8, 6.9, 7, 7.1, 7.2, 7.3, 7.4, 7.5, or 7.6).

In yet another aspect, the disclosure features a method for producing a concentrated antibody solution comprising greater than (or equal to) 100 mg/mL of an anti-C5 antibody. The method comprises: providing a first aqueous solution comprising an anti-C5 antibody, the first aqueous solution 60 having a first formulation and comprising no more than 50 (e.g., no more than 49, 48, 47, 46, 45, 44, 43, 42, 41, 40, 39, 38, 37, 36, 35, 34, 33, 32, or 31) mg/mL of the anti-C5 antibody; subjecting the first aqueous solution to diafiltration to thereby produce a second aqueous solution, wherein the 65 second aqueous solution has a second formulation as a result of the diafiltration; and concentrating the second aqueous

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solution to produce a concentrated antibody solution comprising greater than (or equal to) 100 mg/mL of the anti-C5 antibody. In some embodiments, the first aqueous solution comprises greater than 30 mg/mL, but no more than 50 mg/mL, of the anti-C5 antibody. In some embodiments, the first aqueous solution comprises greater than 35 mg/mL, but no more than 50 mg/mL, of the anti-C5 antibody. In some embodiments, the first aqueous solution comprises greater than 35 mg/mL, but no more than 45 mg/mL, of the anti-C5 antibody. In some embodiments, the anti-C5 antibody is not lyophilized prior to or following the diafiltration or concentrating.

In some embodiments of any of the methods, the first formulation is a phosphate buffer-based formulation. The first formulation can comprise, e.g.: at least 20 mM sodium phosphate and at least 80 mM sodium chloride.

In some embodiments of any of the above methods, the second formulation comprises: at least 20 mM histidine; at least 50 mM serine; at least 2.5% (w/v) sorbitol; and at least 1.5% (w/v) mannitol.

In some embodiments of any of the above methods, the concentrating comprises tangential flow filtration and/or use of a stir cell.

In some embodiments of any of the above methods, more than one round of diafiltration is performed. In some embodiments, at least two rounds of diafiltration are performed. In some embodiments, at least four rounds of diafiltration are performed. In some embodiments of any of the above methods, the diafiltration comprises continuous addition of a buffer having the second formulation.

In some embodiments of any of the above methods, the concentrated antibody solution comprises greater than (or equal to) 105 (e.g., greater than, or equal to, 106, 107, 108, 109, 110, 115, 120, 125, 130, 135, 140, 145, 150, 155, 160, 165, 170, 175, 180, 185, 190, 195, 200, 205, or 208) mg/mL of the anti-C5 antibody.

In some embodiments of any of the above methods, at least 80 (e.g., at least 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, or 99) % of the anti-C5 antibody present in the first aqueous solution is recovered in the high concentration aqueous solution. In some embodiments of any of the above methods, at least 90% of the anti-C5 antibody present in the first aqueous solution is recovered in the high concentration aqueous solution.

In some embodiments of any of the above methods, the anti-C5 antibody is eculizumab.

In yet another aspect, the disclosure features an aqueous solution comprising an anti-C5 antibody at a concentration of greater than 100 mg/mL produced by any of the above methods.

In another aspect, the disclosure features a kit comprising: (i) any of the solutions described herein; and (ii) a means for delivering the solution to a patient in need thereof.

In some embodiments of any of the kits described herein, the means is suitable for subcutaneous delivery of the solution to the patient. In some embodiments of any of the kits described herein, the means is suitable for delivery of the solution to the eye. In some embodiments of any of the kits described herein, the means is suitable for intraarticular delivery of the solution to the patient.

In some embodiments of any of the kits described herein, the means is a syringe or a double-barreled syringe. In some embodiments of any of the kits described herein, the means is:
(a) a transscleral patch comprising the solution; or (b) a contact lens comprising the solution or partially coated in the solution.

In some embodiments of any of the kits described herein, the means is suitable for intrapulmonary delivery of the solution to the patient. For example, the means can be an inhaler or a nebulizer.

In some embodiments of any of the kits described herein, 5 the solution is formulated for aerosol administration or nebulized administration to the patient.

In some embodiments, any of the kits described herein further comprise at least one additional active agent for use in treating a complement-associated disorder in a subject. Such 10 agents are recited herein.

In yet another aspect, the disclosure features a kit comprising one or more containers, wherein each container comprises an aqueous solution described herein and wherein each container comprises at least one pharmaceutical unit dosage form 15 of the anti-C5 antibody. In some embodiments, each container comprises between 0.05 mg to 10 mg of the anti-C5 antibody. In some embodiments, the kit comprises between about 1 mg and 100 mg of the anti-C5 antibody.

In some embodiments of any of the kits described herein, 20 each container has a volume of 0.01 mL to 1 mL, inclusive. In some embodiments of any of the kits described herein, at least one container comprises an aqueous solution suitable for intravitreal injection to a patient, intraarticular injection to a patient, intramuscular injection to a patient, subcutaneous 25 injection to a patient, and/or intrapulmonary administration to a patient. For example, a kit described herein can comprise at least one container comprising an aqueous solution suitable for use with a nebulizer or inhaler.

In another aspect, the disclosure features a pre-filled 30 syringe comprising any of the aqueous solutions described herein. In some embodiments, the solution is formulated for intraocular, intravitreal, and/or intraarticular administration.

In some embodiments, the solution is formulated for intramuscular or subcutaneous administration.

In some embodiments, any of the pre-filled syringes described herein comprise at least one pharmaceutical unit dosage form of the anti-C5 antibody in the solution. Each pharmaceutical unit dosage form can have, e.g., a volume of between 0.02 mL to 0.1 mL, inclusive. In some embodiments, 40 the pharmaceutical unit dosage form has a volume of no more than 0.05 mL.

In some embodiments, any of the pre-filled syringes described herein comprise between 0.05 mg to 10 mg of the anti-C5 antibody. In some embodiments, the syringe comprises between about 1 mg and 100 mg of the anti-C5 antibody.

In yet another aspect, the disclosure features a method for treating a patient afflicted with a complement-associated disorder. The method comprises administering to a patient 50 afflicted with a complement-associated disorder a therapeutically effective amount of any of the aqueous solutions described herein to thereby treat the complement-associated disorder. In some embodiments, the methods can be performed using any of the kits or pre-filled syringes described 55 herein. In some embodiments, the method can further comprise, prior to administering the aqueous solution to the patient, determining that the patient is afflicted with the complement-associated disorder.

In some embodiments, the complement-associated disorder is a complement-associated disorder of the eye. For example, the complement-associated disorder of the eye can be age-related macular degeneration (AMD), a diabetes-associated ocular disorder, or central retinal vein occlusion. In some embodiments, the complement-associated disorder of 65 the eye is wet AMD. In some embodiments, the disorder is dry AMD. In such embodiments, the aqueous solution can be

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administered to the patient by way of intravitreal injection. In such embodiments, the aqueous solution can be administered to the patient by way of a transscleral patch or as an eye drop (for example, the solution can be formulated for use as an eye drop).

In some embodiments, the complement-associated disorder is rheumatoid arthritis. In such embodiments, e.g., the aqueous solution can be administered to the patient by way of intraarticular injection. In some embodiments, the aqueous solution can be administered by way of intravenous or subcutaneous injection.

In some embodiments, the complement-associated disorder is a pulmonary disorder. The pulmonary disorder can be selected from the group consisting of, e.g., asthma, chronic obstructive pulmonary disease, acute respiratory distress syndrome, pulmonary fibrosis, α -1 anti-trypsin deficiency, emphysema, bronchiectasis, bronchiolitis obliterans, sarcoidosis, a collagen vascular disorder, and bronchitis. In such embodiments, the aqueous solution can be delivered to the patient by way of intrapulmonary administration, e.g., through the use of a nebulizer or an inhaler.

In some embodiments, the complement-associated disorder is selected from the group consisting of ischemia-reperfusion injury, atypical hemolytic uremic syndrome, thrombotic thrombocytopenic purpura, paroxysmal nocturnal hemoglobinuria, dense deposit disease, age-related macular degeneration, spontaneous fetal loss, Pauci-immune vasculitis, epidermolysis bullosa, recurrent fetal loss, multiple sclerosis, traumatic brain injury, myasthenia gravis, cold agglutinin disease, dermatomyositis, Degos' disease, Graves' disease, Hashimoto's thyroiditis, type I diabetes, psoriasis, pemphigus, autoimmune hemolytic anemia, idiopathic thrombocytopenic purpura, Goodpasture syndrome, multifocal motor neuropathy, neuromyelitis optica, antiphospholipid syndrome, and catastrophic antiphospholipid syndrome.

In some embodiments, any of the therapeutic methods described herein further comprise administering to the patient one or more additional therapeutic agents for (a) treating a complement-associated disorder or (b) ameliorating one or more symptoms associated with the complement-associated disorder.

In yet another embodiment, the disclosure features an aqueous solution comprising an anti-C5 antibody at a concentration of at least 40 mg/mL. The anti-C5 antibody in the solution remains at least 97% monomeric during storage at 2° C. to 8° C. for at least six months as determined by SEC-HPLC. In some embodiments, the concentration of the anti-C5 antibody in the solution is at least 50 (e.g., at least 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, or 200) mg/mL. In some embodiments, the concentration of the anti-C5 antibody in the solution is greater than 200 mg/mL. The antibody can be, e.g., eculizumab.

In another aspect, the disclosure features an aqueous solution comprising an anti-C5 antibody at a concentration of greater than 40 mg/mL with the proviso that the solution is not formulated as follows: 20 mM histidine, 50 mM glycine, 3% (w/v) sorbitol, 1.5% (w/v) mannitol, 0.001% to 0.02% Tween 80, and a pH of 6 to 8 (e.g., with a physiologic osmolality). In some embodiments, the concentration of the anti-C5 antibody in the solution is at least 50 (e.g., at least 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, or 200) mg/mL. In some embodiments, the concentration of the anti-C5 antibody in the solution is greater than 200 mg/mL. The antibody can be, e.g., eculizumab.

In some embodiments of the aqueous solutions described above, the anti-C5 antibody remains at least 98% monomeric

during storage at 2° C. to 8° C. for at least one year (e.g., at least 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, or 24 months) as determined by SEC-HPLC.

In some embodiments of the aqueous solutions described above, less than 2 (e.g., less than 1.9, 1.8, 1.7, 1.6, 1.5, 1.4, 5 1.3, 1.2, 1.1, 1.0, 0.9, 0.8, 0.7, 0.6, 0.5, 0.4, 0.3, or 0.2) % of the anti-C5 antibody in the solution is aggregated as determined by SEC-HPLC.

In some embodiments of the aqueous solutions described above, less than 0.5 (e.g., less than 0.4, 0.3, 0.2, or 0.1) % of the anti-C5 antibody in the solution is fragmented as determined by SEC-HPLC.

In some embodiments of the aqueous solutions described above, during storage at 2° C. to 8° C. for at least six (e.g., at least seven, eight, nine, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 15 20, 21, 22, 23, or 24) months the anti-C5 antibody retains at least 90 (e.g., at least 91, 92, 93, 94, 95, 96, 97, 98, or 99) % of its C5-binding activity, as compared to a reference anti-C5 antibody corresponding to the anti-C5 antibody prior to storage. Suitable methods for evaluating the binding activity of a sample of a stored solution containing a specified concentration of anti-C5 antibody are known in the art and described herein.

In some embodiments of the aqueous solutions described above, during storage at 2° C. to 8° C. for at least six (e.g., at 25 least seven, eight, nine, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, or 24) months the anti-C5 antibody retains at least 90 (e.g., at least 91, 92, 93, 94, 95, 96, 97, 98, or 99) % of its ability to inhibit hemolysis, as compared to a reference anti-C5 antibody corresponding to the anti-C5 antibody prior 30 to storage. Suitable methods for evaluating the ability of a sample of a stored solution containing a specified concentration of anti-C5 antibody to inhibit hemolysis are known in the art and described herein.

In some embodiments of the aqueous solutions described above, the solutions can be sterile. In some embodiments of any of the aqueous solutions described above, the solutions can comprise: at least 20 mM histidine; at least 50 mM serine; at least 3% (w/v) sorbitol; and at least 1.5% (w/v) mannitol. In some embodiments of the aqueous solutions described above, 40 the solutions can comprise: at least 2.0 mM histidine; at least 50 mM serine; at least 2.5% (w/v) sorbitol; and at least 1.5% (w/v) mannitol. In some embodiments of any of the aqueous solutions described above, the solutions can contain: 20 mM histidine; 50 mM serine; 3% (w/v) sorbitol; and 1.5% (w/v) 45 mannitol.

In some embodiments of the aqueous solutions described above, the solution can comprise a surfactant such as, for example, polysorbate 20 or polysorbate 80. The concentration of the surfactant in the solution can be, e.g., between 50 0.001% to 0.02%, inclusive.

In some embodiments of the aqueous solutions described above, the pH of the solution can be, for example, between 6.5 and 7.5, inclusive.

As used throughout the present disclosure, the term "antibody" refers to a whole antibody (e.g., IgM, IgG, IgA, IgD, or IgE) molecule as well as antigen-binding fragments thereof, which fragments can be generated by any one of a variety of methods that are known in the art and described herein. The term "antibody" includes a polyclonal antibody, a monoclonal antibody, a chimerized or chimeric antibody, a humanized antibody, a deimmunized human antibody, and a fully human antibody. The antibody can be made in or derived from any of a variety of species, e.g., mammals such as humans, non-human primates (e.g., monkeys, baboons, or chimpanzees), horses, cattle, pigs, sheep, goats, dogs, cats, rabbits, guinea pigs, gerbils, hamsters, rats, and mice. The antibody

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can be a purified or a recombinant antibody. "Antibody fragments," "antigen-binding fragments," or similar terms refer to a fragment of an antibody that retains the ability to bind to an antigen (e.g., human complement component C5 or a biologically active fragment thereof such as C5a or C5b), e.g., a single chain antibody, a single chain Fv fragment (scFv), an Fd fragment, an Fab fragment, an Fab' fragment, or an F(ab') fragment. An scFv fragment is a single polypeptide chain that includes both the heavy and light chain variable regions of the antibody from which the scFv is derived. In addition, diabodies (Poljak (1994) Structure 2(12):1121-1123; Hudson et al. (1999) J Immunol Methods 23(1-2):177-189, the disclosures of both of which are incorporated herein by reference in their entirety), minibodies, single domain or nanobodies (Huang et al. (2010) Expert Rev Mol Diagn 10(6):777-785; Smolarek et al. (2010) Cell Mol Life Sci 67(19):3371-3387), and intrabodies (Huston et al. (2001) Hum Antibodies 10(3-4): 127-142; Wheeler et al. (2003) Mol Ther 8(3):355-366; Stocks (2004) Drug Discov Today 9(22):960-966, the disclosures of each of which are incorporated herein by reference in their entirety) that bind to human C5 protein are embraced by the definition of "antigen-binding fragment" and can be incorporated into the compositions and used in the methods described herein.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this disclosure pertains. In case of conflict, the present document, including definitions, will control. Preferred methods and materials are described below, although methods and materials similar or equivalent to those described herein can also be used in the practice or testing of the presently disclosed methods and compositions. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety.

Other features and advantages of the present disclosure, e.g., methods for treating a complement-associated disorder, will be apparent from the following description, the examples, and from the claims.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 is a flow chart depicting the formulation scheme for preparing five different solutions of eculizumab. A detailed description of the particular composition of solutions I to V, as well as the nature of the HSSM and HTT buffers is set forth in Example 2 (infra).

DETAILED DESCRIPTION

The disclosure features stable, aqueous solutions containing a high concentration of an antibody that binds to human complement component C5. The solutions can be used in a variety of therapeutic applications such as methods for treating or preventing complement-associated disorders. While in no way intended to be limiting, exemplary solutions, formulations, therapeutic kits, and methods for making and using any of the foregoing are elaborated on below and are exemplified in the working Examples.

Highly-Concentrated Antibody Solutions

The disclosure provides aqueous solutions comprising a high concentration of an antibody that binds to human complement component C5 [hereinafter an "anti-C5 antibody" or an "anti-human C5 antibody"] such as eculizumab. Such solutions are sometimes referred to herein as "high concentration antibody solutions." As used herein, a "high concentration" of an anti-C5 antibody in an aqueous solution is a concentration of the antibody that is at least, equal to, or

greater than, 40 (e.g., at least, equal to, or greater than, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, 150, 155, 160, 165, 170, 175, 180, 185, 190, 195, 200, 205, 210, 215, 220, 225, 230, 235, 240, 245, 250, 255, 260, 265, 270, 275, 280, 285, or 290) mg/mL. In 5 some embodiments, the anti-C5 antibody is present in the solution at a concentration of more than 200 (e.g., more than 200, 205, 210, 215, 220, 225, 230, 235, 240, 245, 250, 255, 260, 265, 270, 275, 280, 285, or 290) mg/mL. In some embodiments, the antibody is present in the solution at a 10 concentration of, e.g., 40 mg/mL to 200 mg/mL, 50 mg/mL to 200 mg/mL, 60 mg/mL to 200 mg/mL, 70 mg/mL to 200 mg/mL, 80 mg/mL to 200 mg/mL, 90 mg/mL to 200 mg/mL, 100 mg/mL to 200 mg/mL, 110 mg/mL to 200 mg/mL, 120 mg/mL to 200 mg/mL, 130 mg/mL to 200 mg/mL, 140 15 mg/mL to 200 mg/mL, 150 mg/mL to 200 mg/mL, 40 mg/mL to 100 mg/mL, 50 mg/mL to 100 mg/mL, 60 mg/mL to 100 mg/mL, 70 mg/mL to 100 mg/mL, 80 mg/mL to 100 mg/mL, 90 mg/mL to 100 mg/mL, 40 mg/mL to 150 mg/mL, 50 mg/mL to 150 mg/mL, 60 mg/mL to 150 mg/mL, 70 mg/mL 20 to 150 mg/mL, 80 mg/mL to 150 mg/mL, 90 mg/mL to 150 mg/mL, 100 mg/mL to 150 mg/mL, 110 mg/mL to 150 mg/mL, 120 mg/mL to 150 mg/mL, 40 mg/mL to 50 mg/mL, 40 mg/mL to 250 mg/mL, 50 mg/mL to 250 mg/mL, 60 mg/mL to 250 mg/mL, 70 mg/mL to 250 mg/mL, 80 mg/mL 25 to 250 mg/mL, 90 mg/mL to 250 mg/mL, 100 mg/mL to 250 mg/mL, 110 mg/mL to 250 mg/mL, 120 mg/mL to 250 mg/mL, 130 mg/mL to 250 mg/mL, 140 mg/mL to 250 mg/mL, 150 mg/mL to 250 mg/mL, 160 mg/mL to 250 mg/mL, 170 mg/mL to 250 mg/mL, 180 mg/mL to 250 30 mg/mL, 190 mg/mL to 250 mg/mL, 200 mg/mL to 250 mg/mL, greater than 200 mg/mL (e.g., at least 201 mg/mL) to 250 mg/mL, or greater than 200 mg/mL (e.g., 201 mg/mL or greater) to 300 mg/mL.

In some embodiments, the anti-C5 antibody binds to an 35 epitope in the human pro-C5 precursor protein. For example, the anti-C5 antibody can bind to an epitope in the human complement component C5 protein comprising, or consisting of, the amino acid sequence depicted in SEQ ID NO:1 (NCBI Accession No. AAA51925 and Haviland et al., supra).

An "epitope" refers to the site on a protein (e.g., a human complement component C5 protein) that is bound by an antibody. "Overlapping epitopes" include at least one (e.g., two, three, four, five, or six) common amino acid residue(s).

In some embodiments, the anti-C5 antibody binds to an 45 epitope in the human pro-C5 precursor protein lacking the leader sequence. For example, the anti-C5 antibody can bind to an epitope in the human complement component C5 protein comprising, or consisting of, the amino acid sequence depicted in SEQ ID NO:2, which is a human C5 protein 50 lacking the amino terminal leader sequence.

In some embodiments, the anti-C5 antibody can bind to an epitope in the alpha chain of the human complement component C5 protein. For example, the anti-C5 antibody can bind to an epitope within, or overlapping with, a protein having the 55 amino acid sequence depicted in SEQ ID NO:3, which is the human complement component C5 alpha chain protein. Antibodies that bind to the alpha chain of C5 are described in, for example, Ames et al. (1994) *J Immunol* 152:4572-4581.

In some embodiments, the anti-C5 antibody can bind to an 60 epitope in the beta chain of the human complement component C5 protein. For example, the anti-C5 antibody can bind to an epitope within, or overlapping with, a protein having the amino acid sequence depicted in SEQ ID NO:4, which is the human complement component C5 beta chain protein. Antibodies that bind to the C5 beta chain are described in, e.g., Moongkarndi et al. (1982) *Immunobiol* 162:397;

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Moongkarndi et al. (1983) *Immunobiol* 165:323; and Mollnes et al. (1988) *Scand J Immunol* 28:307-312.

In some embodiments, the anti-C5 antibody can bind to an epitope within, or overlapping with, an antigenic peptide fragment of a human complement component C5 protein. For example, the anti-C5 antibody can bind to an epitope within, or overlapping with, an antigen peptide fragment of a human complement component C5 protein, the fragment containing, or consisting of, the following amino acid sequence: VIDHQGTKSSKCVRQKVEGSS (SEQ ID NO:5) or KSSKC (SEQ ID NO:6).

In some embodiments, the anti-C5 antibody can bind to an epitope within, or overlapping with, a fragment of a human complement component C5 protein, the fragment containing, or consisting of, any one of the following amino acid sequences (which are exemplary antigenic fragments of SEQ ID NO: 1): NFSLETWFGKEILVKTLRVVPEGVKR-ESYSGVTLDPRGIYGTISRRKEFPYRIPLD LVPK-TEIKRILSVKGLLVGEILSAVLSQEGI-

NILTHLPKGSAEAELMSVVPVFYVFH
YLETGNHWNIFHSD (SEQ ID NO:7);
SESPVIDHQGTKSSKCVRQKVEGSSSHLVTFTVLPLEIGLHNINFSLETWFGKEILV KTLRVVPEGVKRESYSGVTLDPRGIYGTISR-

RKEFPYRIPLDLVPKTEIKRILSVKG
LLVGEILSAVLSQEGINILTHLPKGSAEAELMSVVPVFYVFHYLETGNHWNIFHSD
PLIEKQKLKKKLKEGMLSIMSYRNADYSYS (SEQ ID NO:8); SHKDMQLGRLHMKTLLPVSKPEIRSYFPES
(SEQ ID NO:9); SHKDMQLGRLHMKTLLPVSKPEIRSYFPESWLWEVHLVPRRKQLQFALPDSLTT WEIQGIGISNTGICVADTVKAKVFKDVFLEMNIPYSVVRGEQIQLKGTVYNYRTS
GMQFCVKMSAVEGICT-

35 SESPVIDHQGTKSSKCVRQKVEG-SSSHLVTFTVLPLEIG LHNINFSLETWFGKEILVKTL-RVVPEGVKRESYSGVTLDPRGIYGTISRRKEFPYRI PLDLVPKTEIKRILSVKGLLVGEILSAV-LSQEGINILTHLPKGSAEAELMSVVPVFY VFHYLET-40 GNHWNIFHSDPLIEKQKLKKKLKEGML-SIMSYRNADYSYS (SEQ ID NO: 10); and DHQGTKSSKCVRQKVEG (SEQ ID NO: 11).

Additional exemplary antigenic fragments of human complement component C5 are disclosed in, e.g., U.S. Pat. No. 6,355,245, the disclosure of which is incorporated herein by reference.

In some embodiments, the anti-C5 antibody specifically binds to a human complement component C5 protein (e.g., the human C5 protein having the amino acid sequence depicted in SEQ ID NO: 1). The terms "specific binding" or "specifically binds" refer to two molecules forming a complex (e.g., a complex between an antibody and a complement component C5 protein) that is relatively stable under physiologic conditions. Typically, binding is considered specific when the association constant (K_a) is higher than 10^6 M^{-1} . Thus, an antibody can specifically bind to a C5 protein with a K_a of at least (or greater than) 10^6 (e.g., at least or greater than 10^{7} , 10^{8} , 10^{9} , 10^{10} , 10^{11} 10^{12} , 10^{13} , 10^{14} , or 10^{15} or higher) M^{-1} . Examples of antibodies that specifically bind to a human complement component C5 protein are described in, e.g., U.S. Pat. No. 6,355,245, the disclosure of which is incorporated herein by reference in its entirety.

Methods for determining whether an antibody binds to a protein antigen and/or the affinity for an antibody to a protein antigen are known in the art. For example, the binding of an antibody to a protein antigen can be detected and/or quantified using a variety of techniques such as, but not limited to,

Western blot, dot blot, surface plasmon resonance (SPR) method (e.g., BIAcore™ system; Pharmacia Biosensor AB, Uppsala, Sweden and Piscataway, N.J.), or enzyme-linked immunosorbent assays (ELISA). See, e.g., Harlow and Lane (1988) "Antibodies: A Laboratory Manual" Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.; Benny K. C. Lo (2004) "Antibody Engineering: Methods and Protocols," Humana Press (ISBN: 1588290921); Borrebaek (1992) "Antibody Engineering, A Practical Guide," W.H. Freeman and Co., NY; Borrebaek (1995) "Antibody Engineering," 2nd 10 Edition, Oxford University Press, NY, Oxford; Johne et al. (1993) *J Immunol Meth* 160:191-198; Jonsson et al. (1993) *Ann Biol Clin* 51:19-26; and Jonsson et al. (1991) *Biotechniques* 11:620-627. See also, U.S. Pat. No. 6,355,245.

In some embodiments, the anti-C5 antibody can cross-block binding of another antibody that binds to an epitope within, or overlapping with, a human complement component C5 protein. In some embodiments, the anti-C5 antibody can crossblock binding of an antibody that binds to an epitope within, or overlapping with, a peptide fragment of a human complement component C5 protein. The peptide fragment can be a fragment of a human complement component C5 protein having the amino acid sequence depicted in any one of SEQ ID NOS:1-11. For example, the peptide fragment can contain, or consist of, the following amino acid sequence: 25 VIDHQGTKSSKCVRQKVEGSS (SEQ ID NO:5).

As used herein, the term "crossblocking antibody" refers to an antibody that lowers the amount of binding of anti-C5 antibody to an epitope on a complement component C5 protein relative to the amount of binding of the anti-C5 antibody 30 to the epitope in the absence of the antibody. Suitable methods for determining whether a first antibody crossblocks binding of a second antibody to an epitope are known in the art. For example, crossblocking antibodies can be identified by comparing the binding of the 5G1.1 anti-C5 monoclonal 35 antibody (produced by the hybridoma cell line ATCC designation HB-11625; see U.S. Pat. No. 6,355,245) in the presence and absence of a test antibody. Decreased binding of the 5G1.1 antibody in the presence of the test antibody as compared to binding of the 5G1.1 antibody in the absence of the 40 test antibody indicates the test antibody is a crossblocking antibody.

Methods for identifying the epitope to which a particular antibody (e.g., an anti-C5 antibody) binds are also known in the art. For example, the binding epitope of an anti-C5 anti-45 body can be identified by measuring the binding of the antibody to several (e.g., three, four, five, six, seven, eight, nine, 10, 15, 20, or 30 or more) overlapping peptide fragments of a complement component C5 protein (e.g., several overlapping fragments of a protein having the amino acid sequence 50 depicted in any one of SEQ ID NOs: 1-11). Each of the different overlapping peptides is then bound to a unique address on a solid support, e.g., separate wells of a multi-well assay plate. Next, the anti-C5 antibody is interrogated by contacting it to each of the peptides in the assay plate for an 55 amount of time and under conditions that allow for the antibody to bind to its epitope. Unbound anti-C5 antibody is removed by washing each of the wells. Next, a detectablylabeled secondary antibody that binds to the anti-C5 antibody, if present in a well of the plate, is contacted to each of the 60 wells, and unbound secondary antibody is removed by washing steps. The presence or amount of the detectable signal produced by the detectably-labeled secondary antibody in a well is an indication that the anti-C5 antibody binds to the particular peptide fragment associated with the well. See, 65 e.g., Harlow and Lane (supra), Benny K. C. Lo (supra), and U.S. Patent Application Publication No. 20060153836, the

disclosure of which is incorporated by reference in its entirety. A particular epitope to which an antibody binds can also be identified using BIAcore™ chromatographic techniques (see, e.g., Pharmacia BIAtechnology Handbook, "Epitope Mapping," Section 6.3.2, (May 1994); and Johne et al. (1993) *J Immunol Methods* 160:191-8).

The anti-C5 antibodies described herein can have activity in blocking the generation or activity of the C5a and/or C5b active fragments of a complement component C5 protein (e.g., a human C5 protein). Through this blocking effect, the anti-C5 antibodies inhibit, e.g., the proinflammatory effects of C5a and the generation of the C5b-9 membrane attack complex (MAC) at the surface of a cell. Anti-C5 antibodies that have the ability to block the generation of C5a are described in, e.g., Moongkarndi et al. (1982) *Immunobiol* 162:397 and Moongkarndi et al. (1983) *Immunobiol* 165:323.

Inhibition of complement component C5 can also reduce the cell-lysing ability of complement in a subject's body fluids. Such reductions of the cell-lysing ability of complement present can be measured by methods well known in the art such as, for example, by a conventional hemolytic assay such as the hemolysis assay described by Kabat and Mayer (eds), "Experimental Immunochemistry, 2nd Edition," 135-240, Springfield, Ill., CC Thomas (1961), pages 135-139, or a conventional variation of that assay such as the chicken erythrocyte hemolysis method as described in, e.g., Hillmen et al. (2004) N Engl J Med 350(6):552.

In some embodiments, an anti-C5 antibody, or antigenbinding fragment thereof, can reduce the ability of a C5 protein to bind to human complement component C3b (e.g., C3b present in an AP or CP C5 convertase complex) by greater than 50 (e.g., greater than 55, 60, 65, 70, 75, 80, 85, 90, or 95 or more) %. In some embodiments, upon binding to a C5 protein, the anti-C5 antibody or antigen-binding fragment thereof can reduce the ability of the C5 protein to bind to complement component C4b (e.g., C4b present in a CP C5 convertase) by greater than 50 (e.g., greater than 55, 60, 65, 70, 75, 80, 85, 90, or 95 or more) %. Methods for determining whether an antibody can block the generation or activity of the C5a and/or C5b active fragments of a complement component C5 protein, or binding to complement component C4b or C3b, are known in the art and described in, e.g., U.S. Pat. No. 6,355,245 and Wurzner et al. (1991) Complement Inflamm 8:328-340. (See also below.)

In some embodiments, an anti-C5 antibody binds to an amino-terminal region of the alpha chain of a complement component C5 protein, but does not bind to free C5a. Epitopes for an anti-C5 antibody within the amino-terminal region of the alpha chain include, e.g., epitopes within the human sequence VIDHQGTKSSKCVRQKVEGSS (SEQ ID NO:5).

In some embodiments, the composition comprises, and/or the antibody is, eculizumab (Soliris®; Alexion Pharmaceuticals, Inc., Cheshire, Conn.) or a biologically-active fragment thereof. (See, e.g., Kaplan (2002) *Curr Opin Investig Drugs* 3(7): 1017-23; Hill (2005) *Clin Adv Hematol Oncol* 3(11): 849-50; and Rother et al. (2007) *Nature Biotechnology* 25(11): 1256-1488.) The amino acid sequence of the light chain of eculizumab is as follows: DIQMTQSPSSLSAS-VGDRVTITCGASENIYGALNWYQQK-

PGKAPKLLIYGATNLA DGVPSRFSGSGSGTD-FTLTISSLQPEDFATYYCQNVLNTPLTFGQGTKVEIK RTVA APSVFIFPPSDEQLKSGTASVVCLLNN-FYPREAKVQWKVDNALQSGNSQESVTE QDSKD-STYSLSSTLTLSKADYEKHKVY-

ACEVTHQGLSSPVTKSFNRGEC (SEQ ID NO: 12). The amino acid sequence of the heavy chain of eculizumab is as

QVQLVQSGAEVKKPGASVKVSCKASfollows: GYIFSNYWIQWVRQAPGQGLEWMGEILP STEYTENFKDRVTMTRDTST-STVYMELSSLRSEDTAVYYCARYFFGSSPNW YFDVWGQGTLVTVSSASTKGPSVF-WNS-PLAPCSRSTSESTAALGCLVKDYFPEPVTVS GALTSGVHTFPAVLQSSGLYSLSSV-VTVPSSNFGTQTYTCNVDHKPSNTKV DKTVERKCCVECPPCPAPPVAGPSVFLF-PPKPKDTLMISRTPEVTCVVVDVSQED PEVQFNW- 10 YVDGVEVHNAKTKPREEQFNSTYRVVSV-LTVLHQDWLNGKEYKCK VSNKGLPSSIEK-TISKAKGQPREPQVYTLPPSQEEMTKN-QVSLTCLVKGFYPSDIA VEWESNGQPENNYKTTPPV- 15 LDSDGSFFLYSRLTVDKSRWQEGNVFSCSVMHEAL HNHYTQKSLSLSLGK (SEQ ID NO:13).

As described herein and exemplified in the working examples, the featured aqueous solutions provide the anti-C5 antibody formulated therein with marked stability—both 20 physical/chemical stability as well as functional stability. For example, the formulations described herein are capable of maintaining the structural integrity of an anti-C5 antibody present at high concentrations in a solution. That is, an anti-C5 antibody in a featured aqueous buffer can remain pre- 25 dominantly monomeric after storage for at least one month (e.g., at least two months, three months, four months, five months, six months, seven months, eight months, nine months, 10 months, 11 months, 12 months, 13 months, 14 months, 15 months, 16 months, 17 months, 18 months, 19 30 months, 20 months, 21 months, 22 months, 23 months, 24 months, or more) at approximately 2° C. to 8° C. (e.g., storage at, e.g., 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10° C.). As exemplified in the working examples described herein, the inventors provide formulations suitable for maintaining an anti-C5 antibody at 35 approximately 30 mg/mL or approximately 100 mg/mL in predominantly monomeric form for up to two years at approximately 2° C. to 8° C. As used herein, an anti-C5 antibody formulated at a high concentration in a featured aqueous solution is "predominantly monomeric," or in "pre-40 dominantly monomeric form," if the antibody present in the solution is at least 95 (e.g., at least 95.1, 95.2, 95.3, 95.4, 95.5, 95.6, 95.7, 95.8, 95.9, 96, 96.1, 96.2, 96.3, 96.4, 96.5, 96.6, 96.7, 96.8, 96.9, 97, 97.1, 97.2, 97.3, 97.4, 97.5, 97.6, 97.7, 97.8, 97.9, 98, 98.1, 98.2, 98.3, 98.4, 98.5, 98.6, 98.7, 98.8, 45 98.9, 99, 99.1, 99.2, 99.3, 99.4, 99.5, 99.6, 99.7, 99.8, or 99.9 or greater) % monomeric, e.g., as determined using size exclusion chromatography high performance liquid chromatography (SEC-HPLC). That is: less than 5 (e.g., less than 4.9. 4.8, 4.7, 4.6, 4.5, 4.4, 4.3, 4.2, 4.1, 4.0, 3.9, 3.8, 3.7, 3.6, 3.5, 503.4, 3.3, 3.2, 3.1, 3.0, 2.9, 2.8, 2.7, 2.6, 2.5, 2.4, 2.3, 2.2, 2.1, 2, 1.9, 1.8, 1.7, 1.6, 1.5, 1.4, 1.3, 1.2, 1.1, 1, 0.9, 0.8, 0.7, 0.6, 0.5, 0.4, 0.3, 0.2, or 0.1) % of the antibody in the solution is oligomeric, aggregated, and/or fragmented. As used herein, antibody fragmentation refers to improperly assembled con- 55 stituents or degradation products of a whole antibody having a lower molecular weight than the whole antibody. Such fragmentation forms include, but are not limited to, a free monomeric heavy chain polypeptide, a dimeric heavy chain polypeptide (e.g., disulfide-linked heavy chain polypeptide), 60 a dimeric heavy chain polypeptide bound to one light chain polypeptide, a monomeric heavy chain polypeptide bound to one light chain polypeptide, or further degradation product(s) or fragment(s) of a light chain or heavy chain polypeptide. In some embodiments, less than 2 (e.g., less than 1.9, 1.8, 1.7, 65 1.6, 1.5, 1.4, 1.3, 1.2, 1.1, 1, 0.9, 0.8, 0.7, 0.6, 0.5, 0.4, 0.3, 0.2, or 0.1) % of the antibody is aggregated after storage for at

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least one month (e.g., at least two months, three months, four months, five months, six months, seven months, eight months, nine months, 10 months, 11 months, 12 months, 13 months, 14 months, 15 months, 16 months, 17 months, 18 months, 19 months, 20 months, 21 months, 22 months, 23 months, 24 months, or more) at 2° C. to 8° C. In some embodiments, less than 1 (e.g., less than 0.9, 0.8, 0.7, 0.6, 0.5, 0.4, 0.3, 0.2, or 0.1) % of the antibody is fragmented after storage for at least one month (e.g., at least two months, three months, four months, five months, six months, seven months, eight months, nine months, 10 months, 11 months, 12 months, 13 months, 14 months, 15 months, 16 months, 17 months, 18 months, 19 months, 20 months, 21 months, 22 months, 23 months, 24 months, or more) at 2° C. to 8° C. Methods for determining the amount of monomeric antibody, as well as the amount of oligomeric, aggregated, or fragmented forms of the anti-C5 antibody present in solution are described herein and exemplified in the working examples. For example, a skilled artisan can determine the percentage of whole, fragmented, unfolded intermediates, and/or aggregated antibody species present in a given solution using, e.g., size exclusion chromatography high-performance liquid chromatography (SEC-HPLC), static light scattering (SLS), Fourier transform infrared spectroscopy (FTIR), circular dichroism (CD), urea-induced protein unfolding techniques, intrinsic tryptophan fluorescence, non-reducing sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE), and differential scanning calorimetry (DSC). In the working examples described herein, the inventors exemplify the use of, among others, SEC-HPLC and SDS-PAGE to determine the physical state of the anti-C5 antibodies in solu-

some embodiments, the formulation conditions described herein are capable of maintaining the anti-C5 antibody in at least 95 (e.g., at least 95.1, 95.2, 95.3, 95.4, 95.5, 95.6, 95.7, 95.8, 95.9, 96, 96.1, 96.2, 96.3, 96.4, 96.5, 96.6, 96.7, 96.8, 96.9, 97, 97.1, 97.2, 97.3, 97.4, 97.5, 97.6, 97.7, 97.8, 97.9, 98, 98.1, 98.2, 98.3, 98.4, 98.5, 98.6, 98.7, 98.8, 98.9, 99, 99.1, 99.2, 99.3, 99.4, 99.5, 99.6, 99.7, 99.8, or 99.9 or greater) % monomeric form when stored for at least one month (e.g., at least two months, three months, four months, five months, six months, seven months, eight months, nine months, 10 months, 11 months, 12 months, 13 months, 14 months, 15 months, 16 months, 17 months, 18 months, 19 months, 20 months, 21 months, 22 months, 23 months, 24 months, or more) at approximately -20° C. (e.g., -20±5° C.). The percentage of monomeric form of the antibody in solution can be determined using SEC-HPLC. That is: less than 5 (e.g., less than 4.9. 4.8, 4.7, 4.6, 4.5, 4.4, 4.3, 4.2, 4.1, 4.0, 3.9, 3.8, 3.7, 3.6, 3.5, 3.4, 3.3, 3.2, 3.1, 3.0, 2.9, 2.8, 2.7, 2.6, 2.5, 2.4, 2.3, 2.2, 2.1, 2, 1.9, 1.8, 1.7, 1.6, 1.5, 1.4, 1.3, 1.2, 1.1, 1, 0.9, 0.8, 0.7, 0.6, 0.5, 0.4, 0.3, 0.2, or 0.1) % of the antibody in the solution can become oligomeric, aggregated, and/or fragmented, when the aqueous solution is stored for at least one month at -20° C. As described above, in some embodiments, less than 2 (e.g., less than 1.9, 1.8, 1.7, 1.6, 1.5, 1.4, 1.3, 1.2, 1.1, 1, 0.9, 0.8, 0.7, 0.6, 0.5, 0.4, 0.3, 0.2, or 0.1)% of the antibody is aggregated after storage for at least one month (e.g., at least two months, three months, four months, five months, six months, seven months, eight months, nine months, 10 months, 11 months, 12 months, 13 months, 14 months, 15 months, 16 months, 17 months, 18 months, 19 months, 20 months, 21 months, 22 months, 23 months, 24 months, or more) at approximately -20° C. In some embodiments, less than 1 (e.g., less than 0.9, 0.8, 0.7, 0.6, 0.5, 0.4, 0.3, 0.2, or 0.1) % of the antibody is fragmented after storage for at least one month (e.g., at least two months, three months,

four months, five months, six months, seven months, eight months, nine months, 10 months, 11 months, 12 months, 13 months, 14 months, 15 months, 16 months, 17 months, 18 months, 19 months, 20 months, 21 months, 22 months, 23 months, 24 months, or more) at -20° C.

As described herein and exemplified in the working examples, the anti-C5 antibody containing solutions featured herein can retain at least 90 (e.g., 91, 92, 93, 94, 95, 96, 97, 98, 99, or even 100) % of their biological/functional activity (e.g., ability to bind to human C5) after storage for at least one month (e.g., at least two months, three months, four months, five months, six months, seven months, eight months, nine months, 10 months, 11 months, 12 months, 13 months, 14 months, 15 months, 16 months, 17 months, 18 months, 19 months, 20 months, 21 months, 22 months, 23 months, 24 months, or more) at 2° C. to 8° C. Antibody present in a featured solution can retain, in some embodiments, at least 90 (e.g., 91, 92, 93, 94, 95, 96, 97, 98, 99, or even 100) % of its activity to inhibit hemolysis after storage for at least one 20 month (e.g., at least two months, three months, four months, five months, six months, seven months, eight months, nine months, 10 months, 11 months, 12 months, 13 months, 14 months, 15 months, 16 months, 17 months, 18 months, 19 months, 20 months, 21 months, 22 months, 23 months, 24 25 months, or more) at 2° C. to 8° C. Suitable hemolytic assay methods for determining whether an antibody in a featured solution retains its activity are described herein and known in the art, e.g., in vitro hemolytic assays using avian or porcine erythrocytes. Suitable methods for evaluating the ability of an 30 antibody preparation to bind to human complement component C5 are known in the art and described herein.

In some embodiments, any of the aqueous solutions described herein can contain one or more common excipients and/or additives such as buffering agents, sugars or saccha- 35 rides, salts, and surfactants. Additionally or alternatively, the solutions can further contain one or more solubilizers, diluents, binders, stabilizers, salts, lipophilic solvents, amino acids, chelators, or preservatives.

The solutions described herein can also include a buffering 40 or pH-adjusting agent. In some embodiments, any of the aqueous solutions described herein can have, or can be adjusted to have, a neutral pH. As used herein, "neutral pH" is a pH that is between, and inclusive of, 7 and 8. Accordingly, as used herein neutral pH is inclusive of particular pH values 45 such as 7, 7.1, 7.2, 7.3, 7.4, 7.5, 7.6, 7.7, 7.8, 7.9, and 8.0. In some embodiments, neutral pH is at least pH 7 (e.g., at least pH 7.1, 7.2, 7.3, 7.4, 7.5, 7.6, 7.7, 7.7 or 7.9), but less than pH 8 (e.g., less than pH 7.9, 7.8, 7.7, 7.6, 7.5, 7.4, 7.3, 7.2, or 7.1). That is, in some embodiments neutral pH can be, e.g., at least 50 pH 7, but less than pH 7.5. In some embodiments, neutral pH can be between pH 7 and pH 7.5. In some embodiments, neutral pH can be between pH 7 and pH 7.2. In some embodiments, neutral pH can be, e.g., pH 7. One of skill in the art will a healthy patient) has a neutral pH as defined herein, e.g., the pH of human blood is approximately pH 7.35 to pH 7.45. See, e.g., Boron and Boulpaep (2003) "Medical physiology: a cellular and molecular approach," W.B. Saunders, New York (ISBN:0721632564). In some embodiments, the pH of a 60 highly-concentrated antibody solution described herein is between approximately 6.4 and 7.5, inclusive (e.g., approximately 6.2, 6.3, 6.4, 6.5, 6.6, 6.7, 6.8, 6.9, 7, 7.1, 7.2, 7.3, 7.4, 7.5, 7.6, or 7.7).

Buffering agents useful in the aqueous solutions described 65 herein include, e.g., salts of citric acid, ascorbic acid, gluconic acid, carbonic acid, tartaric acid, succinic acid, acetic

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acid, or phthalic acid. In some embodiments, the buffer is a Tris-based or phosphate buffer.

In some embodiments, the aqueous solutions described herein can include one or more amino acids, which can, among other things, provide buffering capacity. Suitable amino acids for use in the solutions featured herein include, e.g., histidine, glycine, and serine. In some embodiments, the featured solutions do not include a free amino acid as a buffering agent. In some embodiments, the featured solutions include but one free amino acid (e.g., histidine) as a buffering agent. In some embodiments, the featured solutions can include two or more (e.g., two, three, four, five, six, or seven or more) different amino acids as buffering agents, e.g., serine and histidine.

The buffering agents are generally used at concentrations between approximately 1 mM and 200 mM, depending, in part, on the buffering capacity required. In some embodiments, an aqueous solution described herein can include a buffering agent at a concentration of less than, or approximately, 300 (e.g., less than, or approximately, 290, 280, 270, 260, 250, 240, 230, 220, 210, 200, 190, 180, 170, 160, 150, 140, 130, 120, 110, 100, 90, 80, 70, 60, 50, 40, 30, 25, 20, 15, or 10) mM. In some embodiments, an aqueous solution described herein contains a buffering agent at a concentration of at least 10 (e.g., at least 15, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, 200, 210, 220, 230, 240, 250, 260, 270, 280, 290, or 300 or more) mM. In some embodiments, an aqueous solution described herein can include a buffering agent at a concentration of between about 10 mM to 50 mM, 15 mM to 50 mM, 20 mM to 50 mM, 25 mM to 50 mM, 30 mM to 50 mM, 40 mM to 50 mM, 10 mM to 100 mM, 15 mM to 100 mM, 20 mM to 100 mM, 25 mM to 100 mM, 30 mM to 100 mM, 40 mM to 100 mM, 10 mM to 150 mM, 15 mM to 150 mM, 20 mM to 150 mM, 25 mM to 150 mM, 30 mM to 150 mM, 40 mM to 150 mM, 50 mM to 100 mM, 60 mM to 100 mM, 70 mM to 100 mM, 80 mM to 100 mM, 50 mM to 150 mM, 60 mM to 150 mM, 70 mM to 150 mM, 80 mM to 150 mM, 90 mM to 150 mM, 100 mM to 150 mM, 10 mM to 200 mM, 15 mM to 200 mM, 20 mM to 200 mM, 25 mM to 200 mM, 30 mM to 200 mM, 40 mM to 200 mM, 50 mM to 200 mM, 60 mM to 200 mM, 70 mM to 200 mM, 80 mM to 200 mM, 90 mM to 200 mM, 100 mM to 200 mM, 150 mM to 200 mM, 10 mM to 250 mM, 15 mM to 250 mM, 20 mM to 250 mM, 25 mM to 250 mM, 30 mM to 250 mM, 40 mM to 250 mM, 50 mM to 250 mM, 60 mM to 250 mM, 70 mM to 250 mM, 80 mM to 250 mMmM, 90 mM to 250 mM, 100 mM to 250 mM, 150 mM to 250 mM, or 200 mM to 250 mM. It is understood that in embodiments where a featured solution contains two or more (e.g., at least two, three, four, five, six, seven, eight, nine, or 10 or more) different buffering agents, each of the two or more buffering agents can independently be present at, e.g., one of the above described concentrations.

In some embodiments, any of the aqueous solutions also appreciate that human blood (such as human blood from 55 described herein can contain a salt, e.g., sodium chloride, potassium chloride, or magnesium chloride. In some embodiments, an aqueous solution described herein contains a salt at a concentration of at least 10 (e.g., at least 15, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, 200, 210, 220, 230, 240, 250, 260, 270, 280, 290, or 300 or more) mM. In some embodiments, an aqueous solution described herein can include a salt at a concentration of less than, or approximately, 200 (e.g., less than, or approximately, 190, 180, 170, 160, 150, 140, 130, 120, 110, 100, 90, 80, 70, 60, 50, 40, 30, 25, 20, 15, or 10) mM. In some embodiments, an aqueous solution described herein can include a salt at a concentration of between about 10 mM to

50 mM, 15 mM to 50 mM, 20 mM to 50 mM, 25 mM to 50 mM, 30 mM to 50 mM, 40 mM to 50 mM, 10 mM to 100 mM, 15 mM to 100 mM, 20 mM to 100 mM, 25 mM to 100 mM, 30 mM to 100 mM, 40 mM to 100 mM, 10 mM to 150 mM, 15 mM to 150 mM, 20 mM to 150 mM, 25 mM to 150 mM, 5 30 mM to 150 mM, 40 mM to 150 mM, 50 mM to 100 mM, 60 mM to 100 mM, 70 mM to 100 mM, 80 mM to 100 mM, 50 mM to 150 mM, 60 mM to 150 mM, 70 mM to 150 mM, 80 mM to 150 mM, 90 mM to 150 mM, 100 mM to 150 mM, 10 mM to 200 mM, 15 mM to 200 mM, 20 mM to 200 mM, 10 25 mM to 200 mM, 30 mM to 200 mM, 40 mM to 200 mM, 50 mM to 200 mM, 60 mM to 200 mM, 70 mM to 200 mM, 80 mM to 200 mM, 90 mM to 200 mM, 100 mM to 200 mM, 150 mM to 200 mM, 10 mM to 250 mM, 15 mM to 250 mM. 20 mM to 250 mM. 25 mM to 250 mM. 30 mM to 250 mM. 15 40 mM to 250 mM, 50 mM to 250 mM, 60 mM to 250 mM, 70 mM to 250 mM, 80 mM to 250 mM, 90 mM to 250 mM, 100 mM to 250 mM, 150 mM to 250 mM, or 200 mM to 250 mM. It is understood that in embodiments where a featured solution contains two or more (e.g., at least two, three, four, 20 five, six, seven, eight, nine, or 10 or more) different salts, each of the two or more salts can independently be present at, e.g., one of the above described concentrations.

In some embodiments, any of the aqueous solutions described herein can contain a carbohydrate excipient. Suitable carbohydrate excipients are described in, e.g., Katakam and Banga (1995) *J Pharm Pharmacol* 47(2):103-107; Andya et al. (2003) *AAPS PharmSci* 52): Article 10; and Shire (2009) "Current Trends in Monoclonal Antibody Development and Manufacturing," Volume 11, Springer, 354 pages.

Carbohydrate excipients suitable for use in the solutions described herein include, without limitation, monosaccharides such as fructose, maltose, galactose, glucose, D-mannose, and sorbose; disaccharides such as lactose, sucrose, trehalose, and cellobiose; polysaccharides such as maltodex- 35 trins, dextrans, and starches; and sugar alcohols such as mannitol, xylitol, maltitol, lactitol, and sorbitol. In some embodiments, a carbohydrate excipient is present in a solution featured herein at a concentration of at least, or approximately, 0.5 (e.g., at least, or approximately, 0.6, 0.7, 0.8, 0.9, 40 1, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 2.1, 2.2, 2.3, 2.4, 2.5, 2.6, 2.7, 2.8, 2.9, 3, 3.25, 3.5, 3.75, 4, 4.25, 4.5, 4.75, 5, 5.5, 6, 6.5, 7, 7.5, 8, 8.5, 9, 9.5, 10, or more) %. In embodiments where a featured solution contains two or more (e.g., at least two, three, four, five, six, seven, eight, nine, or 10 or 45 more) different carbohydrate excipients (e.g., sorbitol and mannitol), each excipient can, independently, be present at any of the above-described concentrations.

In some embodiments, an aqueous solution described herein can contain a surfactant such as an anionic, cationic, or 50 nonionic surfactant. Pharmaceutically-acceptable surfactants include, without limitation: polysorbates: TritonTM (e.g., TritonTM 20 or TritonTM 80), sodium octyl glycoside; lauryl-, myristyl-, linoleyl-, or stearylsulfobetaine; lauryl-, myristyl-, linoleyl- or stearyl-sarcosine; linoleyl-, myristyl-, or cetylbe- 55 taine; lauroamidopropyl-, cocamidopropyl-, linoleamidopropyl-, myristamidopropyl-, palmidopropyl-, or isostearamidopropyl-betaine (e.g. lauroamidopropyl); myristamidopropyl-, palmidopropyl-, or isostearamidopropyl-dimethylamine; sodium methyl cocoyl-, or disodium 60 methyl oleyl-taurate; and polyethyl glycol, polypropyl glycol, and copolymers of ethylene and propylene glycol. In some embodiments, the aqueous solutions described herein contain a surfactant (e.g., any of the pharmaceutically-acceptable surfactants described herein or known in the art) at a 65 concentration of at least, or approximately, 0.001 (e.g., at least, or approximately, 0.002, 0.003, 0.004, 0.005, 0.006,

0.007, 0.008, 0.009, 0.01, 0.02, 0.03, 0.04, 0.05, 0.06, 0.07, 0.08, 0.09, 0.1, 0.11, 0.12, 0.13, 0.14, 0.15, 0.16, 0.17, 0.18, 0.19, 0.2, 0.21, 0.22, 0.23, 0.24, 0.25, 0.26, 0.27, 0.28, 0.29, 0.3, 0.31, 0.32, 0.33, 0.34, 0.35, 0.36, 0.37, 0.38, 0.39, 0.4, 0.41, 0.42, 0.43, 0.44, 0.45, 0.46, 0.47, 0.48, 0.49, or 0.5 or more) %. In some embodiments, an aqueous solution described herein contains no more than 0.2 (e.g., no more than 0.19, 0.18, 0.17, 0.16, 0.15, 0.14, 0.13, 0.12, 0.11, 0.10, 0.09, 0.08, 0.07, 0.06, 0.05, 0.04, 0.03, 0.02, 0.01, 0.009, 0.008, 0.007, 0.006, 0.005, 0.004, 0.003, 0.002, or 0.001) % of a pharmaceutically-acceptable surfactant.

In some embodiments, an aqueous solution described herein can be formulated to comprise the following elements: 20 mM histidine, 50 mM serine, 3% sorbitol, and 1.5% mannitol. In some embodiments, this solution is formulated at pH 7. In some embodiments, the aqueous solution can consist of the foregoing elements along with an anti-C5 antibody (e.g., any one of the anti-C5 antibodies described herein) at any of the high concentrations described herein.

In some embodiments, an aqueous solution described herein can comprise, or consist of: (i) an anti-C5 antibody (e.g., eculizumab) at a concentration of at least, or approximately, 80 (e.g., at least, or approximately, 85, 90, 95, 100, 105, 110, 115, 120, 125, 130, 135, 140, 145, 150, 155, 160, 165, 170, 175, 180, 185, 190, 195, 200, 210, 220, 230, 240, or 250 or more) mg/mL; (ii) at least, or approximately, 10 (e.g., at least, or approximately, 15, 20, 25, 30, 35, 40, 45, or 50 or more) mM histidine; (ii) at least, or approximately, 1 (e.g., at least, or approximately, 1.5, 2, 2.5, or 3 or more) % sorbitol; and (v) at least 0.5 (e.g., at least, or approximately, 1.5, 2, 2.5, or 3 or more) % sorbitol; and (v) at least 0.5 (e.g., at least, or approximately, 0.75, 1, 1.25, or 1.5 or more) % mannitol, wherein the solution is formulated at neutral pH (e.g., a pH of approximately 7).

In some embodiments, an aqueous solution described herein can comprise, or consist of: (i) eculizumab at a concentration of at least, or approximately 100 mg/mL; (ii) at least, or approximately, 20 mM histidine; (ii) at least, or approximately, 50 mM serine; (iv) at least, or approximately, 3% sorbitol; and (v) at least, or approximately, 1.5% mannitol, wherein the solution is formulated at a pH of approximately 7.

In some embodiments, an aqueous solution described herein can be formulated to comprise the following elements: 10 mM histidine HCl, 10% alpha-trehalose dihydrate, and 0.01% polysorbate 20. In some embodiments, this solution is formulated at pH 7. In some embodiments, the aqueous solution can consist of the foregoing elements along with an anti-C5 antibody (e.g., any one of the anti-C5 antibodies described herein) at any of the high concentrations described herein

In some embodiments, the aqueous solutions described herein do not contain the following elements at the recited concentrations and pH: 20 mM histidine; 50 nM glycine; 3% (w/v) sorbitol; 1.5% (w/v) mannitol; 0.001% to 0.02% Tween 80; and a pH of 6 to 8. In some embodiments, an aqueous solution described herein does not contain trehalose (e.g., alpha-trehalose). In some embodiments, an aqueous solution described herein is not a phosphate-based buffer (e.g., phosphate buffered saline). For example, in some embodiments, the solution does not contain sodium phosphate. In some embodiments, the aqueous solutions described herein do not contain the following elements at the recited concentrations: 10 mM sodium phosphate, 150 mM sodium chloride, 0.001% to 0.02% Tween 80, and at a pH of 6 to 8.

In some embodiments, any of the aqueous solutions described herein are isotonic with respect to human blood. In

some embodiments, a solution described herein has an osmotic pressure of between approximately 270 mOsm/kg and 328 mOsm/kg, e.g., approximately, 270 mOsm/kg, 275 mOsm/kg, 280 mOsm/kg, 285 mOsm/kg, 290 mOsm/kg, 295 mOsm/kg, 300 mOsm/kg, 305 mOsm/kg, 310 mOsm/kg, 315 5 mOsm/kg, 320 mOsm/kg, 325 mOsm/kg, or 328 mOsm/kg. In some embodiments, a solution described herein has an osmotic pressure that is at least or greater than 250 (e.g., at least, or greater than, 255, 260, 265, 270, 275, 280, 285, 290, 295, 300, 305, 310, 315, 320, 325, or 327) mOsm/kg, but not more than, or less than, 350 (e.g., not more than, or less than, 345, 340, 335, 327, 325, 320, 315, 310, 305, or 300) mOsm/ kg. In some embodiments, the solutions described herein can contain, or be formulated with, one or more tonicity agents useful for maintaining or modulating the osmotic pressure of a solution. For example, a solution described herein can contain one or more amino acids, certain pharmaceutically-acceptable salts, or sugars.

The aqueous solutions described herein can be sterile, 20 pharmaceutical-grade compositions, e.g., for administration to a subject for the treatment or prevention of a complementassociated disorder. The compositions can be formulated according to standard methods. Pharmaceutical formulation is a well-established art, and is further described in, e.g., 25 Gennaro (2000) "Remington: The Science and Practice of Pharmacy," 20th Edition, Lippincott, Williams & Wilkins (ISBN: 0683306472); Ansel et al. (1999) "Pharmaceutical Dosage Forms and Drug Delivery Systems," 7th Edition, Lippincott Williams & Wilkins Publishers (ISBN: 0683305727); 30 and Kibbe (2000) "Handbook of Pharmaceutical Excipients American Pharmaceutical Association," 3rd Edition (ISBN: 091733096X). Suitable formulation methods for the high concentration antibody solutions described herein are exemplified in the working examples.

In some embodiments, a high concentration antibody solution described herein can be formulated for delivery to the eye. As used herein, the term "eye" refers to any and all anatomical tissues and structures associated with an eye. In some embodiments, an aqueous solution described herein can 40 be administered locally, for example, by way of topical application or intravitreal injection. For example, in some embodiments, solution can be formulated for administration by way of an eye dropper.

In some embodiments, a sterile, aqueous solution contains, 45 e.g., additional ingredients such as, but not limited to, preservatives, buffers, tonicity agents, antioxidants and stabilizers, nonionic wetting or clarifying agents, and viscosity-increasing agents. Suitable preservatives for use in such a solution include benzalkonium chloride, benzethonium chloride, 50 chlorobutanol, thimerosal and the like. Suitable buffers include, e.g., boric acid, sodium and potassium bicarbonate, sodium and potassium borates, sodium and potassium carbonate, sodium acetate, and sodium biphosphate, in amounts sufficient to maintain the pH at between about pH 6 and pH 8 55 (see above for suitable pH ranges). Suitable tonicity agents are dextran 40, dextran 70, dextrose, glycerin, potassium chloride, propylene glycol, and sodium chloride.

Suitable antioxidants and stabilizers include sodium bisulfite, sodium metabisulfite, sodium thiosulfite, and thiourea. Suitable wetting and clarifying agents include polysorbate 80, polysorbate 20, poloxamer 282 and tyloxapol. Suitable viscosity-increasing agents include dextran 40, dextran 70, gelatin, glycerin, hydroxyethylcellulose, hydroxmethylpropylcellulose, lanolin, methylcellulose, petrolatum, polyethylene glycol, polyvinyl alcohol, polyvinylpyrrolidone, and carboxymethylcellulose.

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In some embodiments, a highly-concentrated antibody solution described herein can be formulated for administration to the eye, e.g., topical administration to the eye of the subject in need of treatment (e.g., a subject afflicted with AMD). For example, in some embodiments, a highly-concentrated antibody solution described herein can be formulated as an eye drop. In addition, a highly-concentrated antibody solution described herein can be formulated for use with any of a variety of devices developed for introducing therapeutic compounds into the vitreal cavity of the eye. For example, U.S. patent application publication no. 20020026176 describes a pharmaceutical-containing plug that can be inserted through the sclera such that it projects into the vitreous cavity to deliver the pharmaceutical agent into the vitreous cavity. In another example, U.S. Pat. No. 5,443,505 describes an implantable device for introduction into a suprachoroidal space or an avascular region for sustained release of drug into the interior of the eye. Additional methods and devices (e.g., a transscleral patch and delivery via contact lenses) for delivery of a therapeutic agent to the eye are described in, e.g., Ambati and Adamis (2002) Prog Retin Eye Res 21(2):145-151; Ranta and Urtti (2006) Adv Drug Delivery Rev 58(11): 1164-1181; Barocas and Balachandran (2008) Expert Opin Drug Delivery 5(1): 1-10(10); Gulsen and Chauhan (2004) Invest Opthalmol Vis Sci 45:2342-2347; Kim et al. (2007) Ophthalmic Res 39:244-254; and PCT publication no. WO 04/073551, the disclosures of which are incorporated herein by reference in their entirety.

In some embodiments, an antibody or antigen-binding fragment described herein can be formulated in a composition suitable for intrapulmonary administration (e.g., for administration via nebulization; see below) to a mammal such as a human. Methods for preparing such compositions are well known in the art and described in, e.g., U.S. patent application publication no. 20080202513; U.S. Pat. Nos. 7,112,341 and 6,019,968; and PCT application publication nos. WO 00/061178 and WO 06/122257, the disclosures of each of which are incorporated herein by reference in their entirety. Dry powder inhaler formulations and suitable systems for administration of the formulations are described in, e.g., U.S. patent application publication no. 20070235029, PCT Publication No. WO 00/69887; and U.S. Pat. No. 5,997, 848

Pulmonary drug delivery may be achieved by inhalation, and administration by inhalation herein may be oral and/or nasal. Examples of pharmaceutical devices for pulmonary delivery include metered dose inhalers and nebulizers. For example, an antibody or antigen-binding fragment thereof can be administered to the lungs of a subject by way of a nebulizer. Nebulizers use compressed air to deliver a compound as a liquefied aerosol or mist. A nebulizer can be, e.g., a jet nebulizer (e.g., air or liquid-jet nebulizers) or an ultrasonic nebulizer. Additional devices and intrapulmonary administration methods are set forth in, e.g., U.S. Patent Application Publication 20050271660 Nos. 20090110679, the disclosures of each of which are incorporated herein by reference in their entirety.

In some embodiments, the solutions provided herein are present in unit dosage form, which can be particularly suitable for self-administration. A formulated product of the present disclosure can be included within a container, typically, for example, a vial, cartridge, prefilled syringe or disposable pen. A doser such as the doser device described in U.S. Pat. No. 6,302,855 may also be used, for example, with an injection system of the present disclosure.

An injection system of the present disclosure may employ a delivery pen as described in U.S. Pat. No. 5,308,341. Pen

devices, most commonly used for self-delivery of insulin to patients with diabetes, are well known in the art. Such devices can comprise at least one injection needle (e.g., a 31 gauge needle of about 5 to 8 mm in length), are typically pre-filled with one or more therapeutic unit doses of a therapeutic solution, and are useful for rapidly delivering the solution to a subject with as little pain as possible.

One medication delivery pen includes a vial holder into which a vial of insulin or other medication may be received. The vial holder is an elongate generally tubular structure with 10 proximal and distal ends. The distal end of the vial holder includes mounting means for engaging a double-ended needle cannula. The proximal end also includes mounting means for engaging a pen body which includes a driver and dose setting apparatus. A disposable medication (e.g., a high 15 concentration solution of an anti-C5 antibody) containing vial for use with the prior art vial holder includes a distal end having a pierceable elastomeric septum that can be pierced by one end of a double-ended needle cannula. The proximal end of this vial includes a stopper slidably disposed in fluid tight 20 engagement with the cylindrical wall of the vial. This medication delivery pen is used by inserting the vial of medication into the vial holder. A pen body then is connected to the proximal end of the vial holder. The pen body includes a dose setting apparatus for designating a dose of medication to be 25 delivered by the pen and a driving apparatus for urging the stopper of the vial distally for a distance corresponding to the selected dose. The user of the pen mounts a double-ended needle cannula to the distal end of the vial holder such that the proximal point of the needle cannula pierces the septum on 30 the vial. The patient then selects a dose and operates the pen to urge the stopper distally to deliver the selected dose. The dose selecting apparatus returns to zero upon injection of the selected dose. The patient then removes and discards the needle cannula, and keeps the prior art medication delivery 35 pen in a convenient location for the next required medication administration. The medication in the vial will become exhausted after several such administrations of medication. The patient then separates the vial holder from the pen body. The empty vial may then be removed and discarded. A new 40 vial can be inserted into the vial holder, and the vial holder and pen body can be reassembled and used as explained above. Accordingly, a medication delivery pen generally has a drive mechanism for accurate dosing and ease of use.

A dosage mechanism such as a rotatable knob allows the 45 user to accurately adjust the amount of medication that will be injected by the pen from a prepackaged vial of medication. To inject the dose of medication, the user inserts the needle under the skin and depresses the knob once as far as it will depress. The pen may be an entirely mechanical device or it may be 50 combined with electronic circuitry to accurately set and/or indicate the dosage of medication that is injected into the user. See U.S. Pat. No. 6,192,891.

In some embodiments, the needle of the pen device is disposable and the kits include one or more disposable 55 replacement needles. Pen devices suitable for delivery of the any one of the presently featured antibody solutions are also described in, e.g., U.S. Pat. Nos. 6,277,099; 6,200,296; and 6,146,361, the disclosures of each of which are incorporated herein by reference in their entirety. A microneedle-based pen 60 device is described in, e.g., U.S. Pat. No. 7,556,615, the disclosure of which is incorporated herein by reference in its entirety. See also the Precision Pen Injector (PPI) device, Molly™, manufactured by Scandinavian Health Ltd.

The present disclosure also presents controlled-release or 65 extended-release formulations suitable for chronic and/or self-administration of a medication. The various formulations

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can be administered to a patient in need of treatment with the medication as a bolus or by continuous infusion over a period of time

In some embodiments, a high concentration anti-C5 antibody solution described herein is formulated for sustainedrelease, extended-release, timed-release, controlled-release, or continuous-release administration. In some embodiments, depot formulations are used to administer the antibody to the subject in need thereof. In this method, the antibody is formulated with one or more carriers providing a gradual release of active agent over a period of a number of hours or days. Such formulations are often based upon a degrading matrix which gradually disperses in the body to release the active agent.

In some embodiments, a highly-concentrated antibody solution described herein can be formulated with one or more additional active agents useful for treating or preventing a complement-associated disorder (e.g., an AP-associated disorder or a CP-associated disorder) in a subject. Additional agents for treating a complement-associated disorder in a subject will vary depending on the particular disorder being treated, but can include, without limitation, an antihypertensive (e.g., an angiotensin-converting enzyme inhibitor) [for use in treating, e.g., HELLP syndrome], an anticoagulant, a corticosteroid (e.g., prednisone), or an immunosuppressive agent (e.g., vincristine or cyclosporine A). Examples of anticoagulants include, e.g., warfarin (Coumadin), aspirin, heparin, phenindione, fondaparinux, idraparinux, and thrombin inhibitors (e.g., argatroban, lepirudin, bivalirudin, or dabigatran). An anti-C5 antibody described herein can also be formulated with a fibrinolytic agent (e.g., ancrod, ϵ -aminocaproic acid, antiplasmin-ai, prostacyclin, and defibrotide) for the treatment of a complement-associated disorder. In some embodiments, an anti-C5 antibody can be formulated with a lipid-lowering agent such as an inhibitor of hydroxymethylglutaryl CoA reductase. In some embodiments, an anti-C5 antibody can be formulated with, or for use with, an anti-CD20 agent such as rituximab (RituxanTM; Biogen Idec, Cambridge, Mass.). In some embodiments, e.g., for the treatment of RA, an anti-C5 antibody can be formulated with one or both of infliximab (Remicade®; Centocor, Inc.) and methotrexate (Rheumatrex®, Trexall®). In some embodiments, an anti-C5 antibody described herein can be formulated with a non-steroidal anti-inflammatory drug (NSAID). Many different NSAIDS are available, some over the counter including ibuprofen (Advil®, Motrin®, Nuprin®) and naproxen (Alleve®) and many others are available by prescription including meloxicam (Mobic®), etodolac (Lodine®), nabumetone (Relafen®), sulindac (Clinoril®), tolementin (Tolectin®), choline magnesium salicylate (Trilasate®), diclofenac (Cataflam®, Voltaren®, Arthrotec®), Diflusinal (Dolobid®), indomethicin (Indocin®), ketoprofen (Orudis®, Oruvail®), oxaprozin (Daypro®), and piroxicam (Feldene®). In some embodiments a C5-binding polypeptide can be formulated for use with an anti-hypertensive, an anti-seizure agent (e.g., magnesium sulfate), or an anti-thrombotic agent. Anti-hypertensives include, e.g., labetalol, hydralazine, nifedipine, calcium channel antagonists, nitroglycerin, or sodium nitroprussiate. (See, e.g., Mihu et al. (2007) J Gastrointestin Liver Dis 16(4:419-424.) Anti-thrombotic agents include, e.g., heparin, antithrombin, prostacyclin, or low dose aspirin.

In some embodiments, a highly-concentrated antibody solution described herein can be formulated for administration with one or more additional therapeutic agents for use in treating a complement-associated disorder of the eye. Such additional therapeutic agents can be, e.g., bevacizumab or the Fab fragment of bevacizumab or ranibizumab, both sold by

Roche Pharmaceuticals, Inc., and pegaptanib sodium (Mucogen®; Pfizer, Inc.). Such a kit can also, optionally, include instructions for administering the anti-C5 antibody to a subject.

In some embodiments, a highly-concentrated antibody solution described herein can be formulated with one or more additional therapeutic agents for use in treating a complement-associated pulmonary disorder such as, but not limited to, asthma, chronic obstructive pulmonary disease, acute respiratory distress syndrome, pulmonary fibrosis, α -1 antitrypsin deficiency, emphysema, bronchiectasis, bronchiolitis obliterans, sarcoidosis, a collagen vascular disorder, and bronchitis. Such additional therapeutic agents include, e.g., sympathomimetics (e.g., albuterol), antibiotics, deoxyribonucleases (e.g., Pulmozyme®), anticholinergic drugs, antilgE inhibitors (e.g., anti-IgE antibodies), and corticosteroids.

In some embodiments, a highly-concentrated antibody solution described herein can be formulated for administration to a subject along with intravenous gamma globulin therapy (IVIG), plasmapheresis, plasma replacement, or 20 plasma exchange. In some embodiments, an anti-C5 antibody can be formulated for use before, during, or after a kidney transplant.

When a highly-concentrated antibody solution described herein is to be used in combination with a second active agent, 25 the agents can be formulated separately or together. For example, the respective pharmaceutical compositions can be mixed, e.g., just prior to administration, and administered together or can be administered separately, e.g., at the same or different times (see below).

Methods for Preparing the Highly-Concentrated Antibody Solutions

The disclosure also provides exemplary methods for preparing a highly-concentrated antibody solution containing more than 100 mg/mL of an anti-C5 antibody. For example, 35 as described herein and exemplified in the working examples, the inventors have identified improved methods for concentrating an anti-C5 antibody solution that results not only in a higher recovery of antibody from the process, but also a more concentrated final solution. That is, under this method the 40 anti-C5 antibody eculizumab can be concentrated in solution up to 224 mg/mL with an 85% recovery of the antibody starting material.

The method requires a first aqueous solution comprising an anti-C5 antibody, the first aqueous solution having a first 45 formulation and comprising, preferably, no more than 50 mg/mL of the anti-C5 antibody. In some embodiments, the first aqueous solution having a first formulation comprises no more than approximately 40 mg/mL of the anti-C5 antibody. In some embodiments, the provided solution contains 50 between about 20 to about 50 (e.g., 21, 22, 23, 24, 25, 26, 27, $\,$ 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, or 49) mg/mL of the anti-C5 antibody. In some embodiments, the provided solution contains less than 50 $(e.g., less than \, 49, 48, 47, 46, 45, 44, 43, 42, 41, 40, 39, 38, 37, \ \ 55$ 36, 35, 34, 33, 32, 31, 30, 29, or 28) mg/mL of the anti-C5 antibody. The first formulation buffer can be, e.g., a phosphate-based buffer such as phosphate-buffered saline. Suitable phosphate-based buffers for use in these preparation methods are set forth in the working examples. In some 60 embodiments, the first aqueous solution is initially concentrated to around 30 mg/mL to 50 mg/mL of the anti-C5 antibody. For example, the first aqueous solution can be concentrated to around 30 mg/mL to 40 mg/mL of the anti-C5 antibody using a tangential flow filter (TFF) or a stir cell. In 65 some embodiments, the first aqueous solution is obtained by concentrating a "starting" solution having an anti-C5 anti28

body concentration of less than, or equal to, 15 (e.g., less than or equal to 14, 13, 12, 11, 10, 9, 8, 7, 6, 5, 4, 3, 2 or even 1) mg/mL. The formulation of the starting solution can be the same or can be different from the formulation of the first aqueous solution.

The method includes subjecting the first aqueous solution to diafiltration to thereby produce a second aqueous solution. Diafiltration is a membrane-based separation process useful for reducing, removing, or exchanging salts or other buffer components from a solution of interest. Diafiltration involves a buffering-exchanging process where a protein solution (e.g., the first solution containing the anti-C5 antibody) is placed onto a filter having a specified pore size, wherein pressure applied to the solution upon the column forces components of the solution that are smaller than the pores of the filter to pass through the filter. Higher molecular weight species, such as an anti-C5 antibody, that are unable to pass through the pores of the filter are retained (retentate). By applying a volume of a second buffer solution into the retentate container during the diafiltration process, the lower molecular weight buffer components can be exchanged resulting in a retentate having a different formulation than that of the first solution. Typically, a volume of the second buffer equal to volume of the retentate is applied in each "round" of diafiltration. For example, 5 mL of the first solution can placed on the filter and 5 mL of the second buffer is added to the first solution before or during the application of a pressure suitable to gently force the lower molecular weight components of the buffer (e.g., water, salts, etc.) through the filter. In such an example, pressure would be applied until the initial 10 mL volume was reduced to a 5 mL retentate. A second round of diafiltration could be performed wherein an additional 5 mL of the second buffer is applied to the retentate upon the filter. Pressure is applied to the solution on the filter until the 10 mL volume is reduced again to 5 mL. In some embodiments, diafiltration can involve the use of pressure and/or tangential flow to force low molecular weight molecules across a flat sheet membrane. In some embodiments, the diafiltration buffer can be added continuously during the process to maintain a constant retentate volume while the buffer is exchanged.

In some embodiments, one round of diafiltration will be performed. In some embodiments, two or more (e.g., three, four, five, six, seven, eight, nine, or 10 or more) rounds of diafiltration are performed. Following the one or more rounds of diafiltration, the original formulation of the first aqueous solution has been exchanged to a second formulation, thus resulting in a second aqueous solution. In some embodiments, the second formulation comprises: at least 20 mM histidine; at least 50 mM serine; at least 2.5% (w/v) sorbitol; and at least 1.5% (w/v) mannitol. Exemplary second formulations are described herein and exemplified in the working examples.

The methods can also include, following the diafiltration step, concentrating the second aqueous solution to produce a concentrated antibody solution comprising greater than 100 mg/mL of the anti-C5 antibody. In some embodiments, the concentration step is performed to produce a concentrated antibody solution having greater than 125 (e.g., 130, 135, 140, 145, 150, 155, 160, 165, 170, 175, 180, 185, 190, 195, 200, 205, 210, 215, 220, or greater than 225) mg/mL. The concentration step can include, e.g., tangential flow filtration or a stir cell.

As described herein, the methods allow at least 90 (e.g., at least 91, 92, 93, 94, 95, 96, 97, 98, 99, or even 100) % of the anti-C5 antibody present in the first aqueous solution to be recovered in the high concentration aqueous solution.

Methods for Producing an Antibody

Suitable methods for producing an antibody, or antigenbinding fragments thereof, in accordance with the disclosure are known in the art (see, e.g., U.S. Pat. No. 6,355,245) and described herein. For example, monoclonal anti-C5 antibod- 5 ies may be generated using complement component C5-expressing cells, a C5 polypeptide, or an antigenic fragment of C5 polypeptide, as an immunogen, thus raising an immune response in animals from which antibody-producing cells and in turn monoclonal antibodies may be isolated. The sequence of such antibodies may be determined and the antibodies or variants thereof produced by recombinant techniques. Recombinant techniques may be used to produce chimeric, CDR-grafted, humanized and fully human antibodies based on the sequence of the monoclonal antibodies as well as 15 polypeptides capable of binding to human complement component C5.

Moreover, antibodies derived from recombinant libraries ("phage antibodies") may be selected using C5-expressing cells, or polypeptides derived therefrom, as bait to isolate the 20 antibodies or polypeptides on the basis of target specificity. The production and isolation of non-human and chimeric anti-C5 antibodies are well within the purview of the skilled artisan.

Recombinant DNA technology can be used to modify one 25 or more characteristics of the antibodies produced in non-human cells. Thus, chimeric antibodies can be constructed in order to decrease the immunogenicity thereof in diagnostic or therapeutic applications. Moreover, immunogenicity can be minimized by humanizing the antibodies by CDR grafting 30 and, optionally, framework modification. See, U.S. Pat. Nos. 5,225,539 and 7,393,648, the contents of each of which are incorporated herein by reference.

Antibodies can be obtained from animal serum or, in the case of monoclonal antibodies or fragments thereof, produced in cell culture. Recombinant DNA technology can be used to produce the antibodies according to established procedure, including procedures in bacterial or preferably mammalian cell culture. The selected cell culture system preferably secretes the antibody product.

In another embodiment, a process for the production of an antibody disclosed herein includes culturing a host, e.g., *E. coli* or a mammalian cell, which has been transformed with a hybrid vector. The vector includes one or more expression cassettes containing a promoter operably linked to a first 45 DNA sequence encoding a signal peptide linked in the proper reading frame to a second DNA sequence encoding the antibody protein. The antibody protein is then collected and isolated. Optionally, the expression cassette may include a promoter operably linked to polycistronic (e.g., bicistronic) 50 DNA sequences encoding antibody proteins each individually operably linked to a signal peptide in the proper reading frame

Multiplication of hybridoma cells or mammalian host cells in vitro is carried out in suitable culture media, which include the customary standard culture media (such as, for example Dulbecco's Modified Eagle Medium (DMEM) or RPMI 1640 medium), optionally replenished by a mammalian serum (e.g. fetal calf serum), or trace elements and growth sustaining supplements (e.g. feeder cells such as normal mouse peritoneal exudate cells, spleen cells, bone marrow macrophages, 2-aminoethanol, insulin, transferrin, low density lipoprotein, oleic acid, or the like). Multiplication of host cells which are bacterial cells or yeast cells is likewise carried out in suitable culture media known in the art. For example, for bacteria 65 suitable culture media include medium LE, NZCYM, NZYM, NZM, Terrific Broth, SOB, SOC, 2×YT, or M9 Mini-

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mal Medium. For yeast, suitable culture media include medium YPD, YEPD, Minimal Medium, or Complete Minimal Dropout Medium.

In vitro production provides relatively pure antibody preparations and allows scale-up production to give large amounts of the desired antibodies. Techniques for bacterial cell, yeast, plant, or mammalian cell cultivation are known in the art and include homogeneous suspension culture (e.g. in an airlift reactor or in a continuous stirrer reactor), and immobilized or entrapped cell culture (e.g. in hollow fibers, microcapsules, on agarose microbeads or ceramic cartridges).

Large quantities of the desired antibodies can also be obtained by multiplying mammalian cells in vivo. For this purpose, hybridoma cells producing the desired antibodies are injected into histocompatible mammals to cause growth of antibody-producing tumors. Optionally, the animals are primed with a hydrocarbon, especially mineral oils such as pristane (tetramethyl-pentadecane), prior to the injection. After one to three weeks, the antibodies are isolated from the body fluids of those mammals. For example, hybridoma cells obtained by fusion of suitable myeloma cells with antibody-producing spleen cells from Balb/c mice, or transfected cells derived from hybridoma cell line Sp2/0 that produce the desired antibodies are injected intraperitoneally into Balb/c mice optionally pre-treated with pristane. After one to two weeks, ascitic fluid is taken from the animals.

The foregoing, and other, techniques are discussed in, for example, Kohler and Milstein, (1975) Nature 256:495-497; U.S. Pat. No. 4,376,110; Harlow and Lane, Antibodies: a Laboratory Manual, (1988) Cold Spring Harbor, the disclosures of which are all incorporated herein by reference. Techniques for the preparation of recombinant antibody molecules are described in the above references and also in, e.g.: WO97/ 08320; U.S. Pat. Nos. 5,427,908; 5,508,717; Smith (1985) Science 225:1315-1317; Parmley and Smith (1988) Gene 73:305-318; De La Cruz et al. (1988) J Biol Chem 263:4318-4322; U.S. Pat. Nos. 5,403,484; 5,223,409; WO88/06630; WO92/15679; U.S. Pat. Nos. 5,780,279; 5,571,698; 6,040, 136; Davis et al. (1999) Cancer Metastasis Rev 18(4):421-5; 40 and Taylor et al. (1992) Nucleic Acids Res 20: 6287-6295; Tomizuka et al. (2000) Proc Natl Acad Sci USA 97(2): 722-727, the contents of each of which are incorporated herein by reference in their entirety.

The cell culture supernatants are screened for the desired antibodies, preferentially by immunofluorescent staining of complement component C5-expressing cells, by immunoblotting, by an enzyme immunoassay, e.g. a sandwich assay or a dot-assay, or a radioimmunoassay.

For isolation of the antibodies, the immunoglobulins in the culture supernatants or in the ascitic fluid may be concentrated, e.g., by precipitation with ammonium sulfate, dialysis against hygroscopic material such as polyethylene glycol, filtration through selective membranes, or the like. If necessary and/or desired, the antibodies are purified by the customary chromatography methods, for example gel filtration, ion-exchange chromatography, chromatography over DEAE-cellulose and/or (immuno-) affinity chromatography, e.g. affinity chromatography with one or more surface polypeptides derived from a complement component C5-expressing cell line, or with Protein-A or -G.

Another embodiment provides a process for the preparation of a bacterial cell line secreting antibodies directed against a C5 protein in a suitable mammal. For example a rabbit is immunized with pooled samples from C5-expressing tissue or cells or C5 polypeptide or fragments thereof. A phage display library produced from the immunized rabbit is constructed and panned for the desired antibodies in accor-

dance with methods well known in the art (such as, e.g., the methods disclosed in the various references incorporated herein by reference).

Hybridoma cells secreting the monoclonal antibodies are also disclosed. The preferred hybridoma cells are genetically stable, secrete monoclonal antibodies described herein of the desired specificity, and can be expanded from deep-frozen cultures by thawing and propagation in vitro or as ascites in vivo.

In another embodiment, a process is provided for the 10 preparation of a hybridoma cell line secreting monoclonal antibodies against a complement component C5 protein. In that process, a suitable mammal, for example a Balb/c mouse, is immunized with one or more polypeptides or antigenic fragments of C5 or with one or more polypeptides or antigenic fragments derived from a C5-expressing cell, the C5-expressing cell itself, or an antigenic carrier containing a purified polypeptide as described. Antibody-producing cells of the immunized mammal are grown briefly in culture or fused with cells of a suitable myeloma cell line. The hybrid 20 cells obtained in the fusion are cloned, and cell clones secreting the desired antibodies are selected. For example, spleen cells of Balb/c mice immunized with a C5-expressing Chronic Lymphocytic Leukemia (CLL) cell line are fused with cells of the myeloma cell line PAI or the myeloma cell 25 line Sp2/0-Ag 14. The obtained hybrid cells are then screened for secretion of the desired antibodies and positive hybridoma cells are cloned.

Methods for preparing a hybridoma cell line include immunizing Balb/c mice by injecting subcutaneously and/or 30 intraperitoneally an immunogenic composition containing human C5 protein (or an immunogenic fragment thereof) several times, e.g., four to six times, over several months, e.g., between two and four months. Spleen cells from the immunized mice are taken two to four days after the last injection 35 and fused with cells of the myeloma cell line PAI in the presence of a fusion promoter, preferably polyethylene glycol. Preferably, the myeloma cells are fused with a three- to twenty-fold excess of spleen cells from the immunized mice in a solution containing about 30% to about 50% polyethyl- 40 ene glycol of a molecular weight around 4000. After the fusion, the cells are expanded in suitable culture media as described supra, supplemented with a selection medium, for example HAT medium, at regular intervals in order to prevent normal myeloma cells from overgrowing the desired hybri- 45 doma cells.

The antibodies and fragments thereof can be "chimeric." Chimeric antibodies and antigen-binding fragments thereof comprise portions from two or more different species (e.g., mouse and human). Chimeric antibodies can be produced 50 with mouse variable regions of desired specificity spliced onto human constant domain gene segments (for example, U.S. Pat. No. 4,816,567). In this manner, non-human antibodies can be modified to make them more suitable for human clinical application (e.g., methods for treating or preventing a 55 complement associated disorder in a human subject).

The monoclonal antibodies of the present disclosure include "humanized" forms of the non-human (e.g., mouse) antibodies. Humanized or CDR-grafted mAbs are particularly useful as therapeutic agents for humans because they are 60 not cleared from the circulation as rapidly as mouse antibodies and do not typically provoke an adverse immune reaction. Methods of preparing humanized antibodies are generally well known in the art. For example, humanization can be essentially performed following the method of Winter and 65 co-workers (see, e.g., Jones et al. (1986) *Nature* 321:522-525; Riechmann et al. (1988) *Nature* 332:323-327; and Verhoeyen

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et al. (1988) Science 239:1534-1536), by substituting rodent CDRs or CDR sequences for the corresponding sequences of a human antibody. Also see, e.g., Staelens et al. (2006) Mol Immunol 43:1243-1257. In some embodiments, humanized forms of non-human (e.g., mouse) antibodies are human antibodies (recipient antibody) in which hypervariable (CDR) region residues of the recipient antibody are replaced by hypervariable region residues from a non-human species (donor antibody) such as a mouse, rat, rabbit, or non-human primate having the desired specificity, affinity, and binding capacity. In some instances, framework region residues of the human immunoglobulin are also replaced by corresponding non-human residues (so called "back mutations"). In addition, phage display libraries can be used to vary amino acids at chosen positions within the antibody sequence. The properties of a humanized antibody are also affected by the choice of the human framework. Furthermore, humanized and chimerized antibodies can be modified to comprise residues that are not found in the recipient antibody or in the donor antibody in order to further improve antibody properties, such as, for example, affinity or effector function.

Fully human antibodies are also provided in the disclosure. The term "human antibody" includes antibodies having variable and constant regions (if present) derived from human germline immunoglobulin sequences. Human antibodies can include amino acid residues not encoded by human germline immunoglobulin sequences (e.g., mutations introduced by random or site-specific mutagenesis in vitro or by somatic mutation in vivo). However, the term "human antibody" does not include antibodies in which CDR sequences derived from the germline of another mammalian species, such as a mouse, have been grafted onto human framework sequences (i.e., humanized antibodies). Fully human or human antibodies may be derived from transgenic mice carrying human antibody genes (carrying the variable (V), diversity (D), joining (J), and constant (C) exons) or from human cells. For example, it is now possible to produce transgenic animals (e.g., mice) that are capable, upon immunization, of producing a full repertoire of human antibodies in the absence of endogenous immunoglobulin production. (See, e.g., Jakobovits et al. (1993) Proc Natl Acad Sci USA 90:2551; Jakobovits et al. (1993) Nature 362:255-258; Bruggemann et al. (1993) Year in Immunol. 7:33; and Duchosal et al. (1992) Nature 355:258.) Transgenic mice strains can be engineered to contain gene sequences from unrearranged human immunoglobulin genes. The human sequences may code for both the heavy and light chains of human antibodies and would function correctly in the mice, undergoing rearrangement to provide a wide antibody repertoire similar to that in humans. The transgenic mice can be immunized with the target protein (e.g., a complement component C5 protein, fragments thereof, or cells expressing C5 protein) to create a diverse array of specific antibodies and their encoding RNA. Nucleic acids encoding the antibody chain components of such antibodies may then be cloned from the animal into a display vector. Typically, separate populations of nucleic acids encoding heavy and light chain sequences are cloned, and the separate populations then recombined on insertion into the vector, such that any given copy of the vector receives a random combination of a heavy and a light chain. The vector is designed to express antibody chains so that they can be assembled and displayed on the outer surface of a display package containing the vector. For example, antibody chains can be expressed as fusion proteins with a phage coat protein from the outer surface of the phage. Thereafter, display packages can be screened for display of antibodies binding to a target.

In addition, human antibodies can be derived from phage-display libraries (Hoogenboom et al. (1991) *J Mol Biol* 227: 381; Marks et al. (1991) *J Mol Biol* 222:581-597; and Vaughan et al. (1996) *Nature Biotech* 14:309 (1996)). Synthetic phage libraries can be created which use randomized 5 combinations of synthetic human antibody V-regions. By selection on antigen fully human antibodies can be made in which the V-regions are very human-like in nature. See, e.g., U.S. Pat. Nos. 6,794,132; 6,680,209; and 4,634,666, and Ostberg et al. (1983) *Hybridoma* 2:361-367, the contents of each 10 of which are incorporated herein by reference in their entirety.

For the generation of human antibodies, also see Mendez et al. (1998) *Nature Genetics* 15:146-156 and Green and Jakobovits (1998) *J Exp Med* 188:483-495, the disclosures of which are hereby incorporated by reference in their entirety. 15 Human antibodies are further discussed and delineated in U.S. Pat. Nos. 5,939,598; 6,673,986; 6,114,598; 6,075,181; 6,162,963; 6,150,584; 6,713,610; and 6,657,103 as well as U.S. Patent Application Publication Nos. 20030229905 A1, 20040010810 A1, 20040093622 A1, 20060040363 A1, 20 20050054055 A1, 20050076395 A1, and 20050287630 A1. See also International Patent Application Publication Nos. WO 94/02602, WO 96/34096, and WO 98/24893, and European Patent No. EP 0 463 151 B1. The disclosures of each of the above-cited patents, applications, and references are 25 hereby incorporated by reference in their entirety.

In an alternative approach, others, including GenPharm International, Inc., have utilized a "minilocus" approach. In the minilocus approach, an exogenous Ig locus is mimicked through the inclusion of pieces (individual genes) from the Ig 30 locus. Thus, one or more V_H genes, one or more D_H genes, one or more J_H genes, a mu constant region, and a second constant region (preferably a gamma constant region) are formed into a construct for insertion into an animal. This approach is described in, e.g., U.S. Pat. Nos. 5,545,807; 5,545,806; 5,625, 35 825; 5,625,126; 5,633,425; 5,661,016; 5,770,429; 5,789,650; 5,814,318; 5,591,669; 5,612,205; 5,721,367; 5,789,215; 5,643,763; 5,569,825; 5,877,397; 6,300,129; 5,874,299; 6,255,458; and 7,041,871, the disclosures of which are hereby incorporated by reference. See also European Patent 40 No. 0 546 073 B1, International Patent Application Publication Nos. WO 92/03918, WO 92/22645, WO 92/22647, WO 92/22670, WO 93/12227, WO 94/00569, WO 94/25585, WO 96/14436, WO 97/13852, and WO 98/24884, the disclosures of each of which are hereby incorporated by reference in their 45 entirety. See further Taylor et al. (1992) Nucleic Acids Res 20: 6287; Chen et al. (1993) Int Immunol 5:647; Tuaillon et al. (1993) Proc Natl Acad Sci USA 90: 3720-4; Choi et al. (1993) Nature Genetics 4: 117; Lonberg et al. (1994) Nature 368: 856-859; Taylor et al. (1994) Int Immunol 6: 579-591; Tuail- 50 lon et al. (1995) J Immunol 154: 6453-65; Fishwild et al. (1996) *Nature Biotechnol* 14: 845; and Tuaillon et al. (2000) Eur J Immunol 10: 2998-3005, the disclosures of each of which are hereby incorporated by reference in their entirety.

In certain embodiments, de-immunized anti-C5 antibodies or antigen-binding fragments thereof are provided. De-immunized antibodies or antigen-binding fragments thereof are antibodies that have been modified so as to render the antibody or antigen-binding fragment thereof non-immunogenic, or less immunogenic, to a given species (e.g., to a human). 60 De-immunization can be achieved by modifying the antibody or antigen-binding fragment thereof utilizing any of a variety of techniques known to those skilled in the art (see, e.g., PCT Publication Nos. WO 04/108158 and WO 00/34317). For example, an antibody or antigen-binding fragment thereof 65 may be de-immunized by identifying potential T cell epitopes and/or B cell epitopes within the amino acid sequence of the

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antibody or antigen-binding fragment thereof and removing one or more of the potential T cell epitopes and/or B cell epitopes from the antibody or antigen-binding fragment thereof, for example, using recombinant techniques. The modified antibody or antigen-binding fragment thereof may then optionally be produced and tested to identify antibodies or antigen-binding fragments thereof that have retained one or more desired biological activities, such as, for example, binding affinity, but have reduced immunogenicity. Methods for identifying potential T cell epitopes and/or B cell epitopes may be carried out using techniques known in the art, such as, for example, computational methods (see e.g., PCT Publication No. WO 02/069232), in vitro or in silico techniques, and biological assays or physical methods (such as, for example, determination of the binding of peptides to MHC molecules, determination of the binding of peptide:MHC complexes to the T cell receptors from the species to receive the antibody or antigen-binding fragment thereof, testing of the protein or peptide parts thereof using transgenic animals with the MHC molecules of the species to receive the antibody or antigenbinding fragment thereof, or testing with transgenic animals reconstituted with immune system cells from the species to receive the antibody or antigen-binding fragment thereof, etc.). In various embodiments, the de-immunized anti-C5 antibodies described herein include de-immunized antigenbinding fragments, Fab, Fv, scFv, Fab' and F(ab')₂, monoclonal antibodies, murine antibodies, engineered antibodies (such as, for example, chimeric, single chain, CDR-grafted, humanized, and artificially selected antibodies), synthetic antibodies and semi-synthetic antibodies.

In some embodiments, a recombinant DNA comprising an insert coding for a heavy chain variable domain and/or for a light chain variable domain of an anti-C5 antibody or a C5 protein-expressing cell line is produced. The term DNA includes coding single stranded DNAs, double stranded DNAs consisting of said coding DNAs and of complementary DNAs thereto, or these complementary (single stranded) DNAs themselves.

Furthermore, a DNA encoding a heavy chain variable domain and/or a light chain variable domain of anti-C5 antibodies can be enzymatically or chemically synthesized to contain the authentic DNA sequence coding for a heavy chain variable domain and/or for the light chain variable domain, or a mutant thereof. A mutant of the authentic DNA is a DNA encoding a heavy chain variable domain and/or a light chain variable domain of the above-mentioned antibodies in which one or more amino acids are deleted, inserted, or exchanged with one or more other amino acids. Preferably said modification(s) are outside the CDRs of the heavy chain variable domain and/or the CDRs of the light chain variable domain of the antibody in humanization and expression optimization applications. The term mutant DNA also embraces silent mutants wherein one or more nucleotides are replaced by other nucleotides with the new codons coding for the same amino acid(s). The term mutant sequence also includes a degenerate sequence. Degenerate sequences are degenerate within the meaning of the genetic code in that an unlimited number of nucleotides are replaced by other nucleotides without resulting in a change of the amino acid sequence originally encoded. Such degenerate sequences may be useful due to their different restriction sites and/or frequency of particular codons which are preferred by the specific host, particularly E. coli, to obtain an optimal expression of the heavy chain murine variable domain and/or a light chain murine variable domain.

The term mutant is intended to include a DNA mutant obtained by in vitro mutagenesis of the authentic DNA according to methods known in the art.

For the assembly of complete tetrameric immunoglobulin molecules and the expression of chimeric antibodies, the recombinant DNA inserts coding for heavy and light chain variable domains are fused with the corresponding DNAs coding for heavy and light chain constant domains, then transferred into appropriate host cells, for example after incorporation into hybrid vectors.

Recombinant DNAs including an insert coding for a heavy chain murine variable domain of an anti-C5 antibody-expressing cell line fused to a human constant domain IgG, for example $\gamma 1$, $\gamma 2$, $\gamma 3$ or $\gamma 4$, in particular embodiments $\gamma 1$ or $\gamma 4$, may be used. Recombinant DNAs including an insert coding 15 for a light chain murine variable domain of an antibody fused to a human constant domain κ or λ , preferably κ , are also provided.

Another embodiment pertains to recombinant DNAs coding for a recombinant polypeptide wherein the heavy chain 20 variable domain and the light chain variable domain are linked by way of a spacer group, optionally comprising a signal sequence facilitating the processing of the antibody in the host cell and/or a DNA sequence encoding a peptide facilitating the purification of the antibody and/or a cleavage 25 site and/or a peptide spacer and/or an agent.

Accordingly, the monoclonal antibodies or antigen-binding fragments of the disclosure can be naked antibodies or antigen-binding fragments that are not conjugated to other agents, for example, a therapeutic agent or detectable label. 30 Alternatively, the monoclonal antibody or antigen-binding fragment can be conjugated to an agent such as, for example, a cytotoxic agent, a small molecule, a hormone, an enzyme, a growth factor, a cytokine, a ribozyme, a peptidomimetic, a chemical, a prodrug, a nucleic acid molecule including cod- 35 ing sequences (such as antisense, RNAi, gene-targeting constructs, etc.), or a detectable label (e.g., an NMR or X-ray contrasting agent, fluorescent molecule, etc.). In certain embodiments, an anti-C5 antibody or antigen-binding fragment (e.g., Fab, Fv, single-chain (scFv), Fab', and F(ab')₂) is 40 linked to a molecule that increases the half-life of the antibody or antigen-binding fragment (see above).

Several possible vector systems are available for the expression of cloned heavy chain and light chain genes in mammalian cells. One class of vectors relies upon the inte- 45 gration of the desired gene sequences into the host cell genome. Cells which have stably integrated DNA can be selected by simultaneously introducing drug resistance genes such as E. coli gpt (Mulligan and Berg (1981) Proc Natl Acad Sci USA, 78:2072-2076) or Tn5 neo (Southern and Berg 50 (1982) J Mol Appl Genet 1:327-341). The selectable marker gene can be either linked to the DNA gene sequences to be expressed, or introduced into the same cell by co-transfection (Wigler et al. (1979) Cell 16:777-785). A second class of vectors utilizes DNA elements which confer autonomously 55 replicating capabilities to an extrachromosomal plasmid. These vectors can be derived from animal viruses, such as bovine papillomavirus (Sarver et al. (1982) Proc Natl Acad Sci USA, 79:7147-7151), polyoma virus (Deans et al. (1984) Proc Natl Acad Sci USA 81:1292-1296), or SV40 virus 60 (Lusky and Botchan (1981) Nature 293:79-81).

Since an immunoglobulin cDNA is comprised only of sequences representing the mature mRNA encoding an antibody protein, additional gene expression elements regulating transcription of the gene and processing of the RNA are 65 required for the synthesis of immunoglobulin mRNA. These elements may include splice signals, transcription promoters,

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including inducible promoters, enhancers, and termination signals. cDNA expression vectors incorporating such elements include those described by Okayama and Berg (1983) *Mol Cell Biol* 3:280-289; Cepko et al. (1984) *Cell* 37:1053-1062; and Kaufman (1985) *Proc Natl Acad Sci USA* 82:689-693

As is evident from the disclosure, the anti-C5 antibodies can be used in therapies (e.g., therapies for a complement associated disorder), including combination therapies.

In the therapeutic embodiments of the present disclosure, bispecific antibodies are contemplated. Bispecific antibodies are monoclonal, preferably human or humanized, antibodies that have binding specificities for at least two different antigens. In the present case, one of the binding specificities is for the human complement component C5 antigen and the other one is for any other antigen.

Methods for making bispecific antibodies are within the purview of those skilled in the art. Traditionally, the recombinant production of bispecific antibodies is based on the co-expression of two immunoglobulin heavy-chain/lightchain pairs, where the two heavy chains have different specificities (Milstein and Cuello (1983) Nature 305:537-539). Antibody variable domains with the desired binding specificities (antibody-antigen combining sites) can be fused to immunoglobulin constant domain sequences. The fusion preferably is with an immunoglobulin heavy-chain constant domain, including at least part of the hinge, $C_H 2$, and $C_H 3$ regions. DNAs encoding the immunoglobulin heavy-chain fusions and, if desired, the immunoglobulin light chain, are inserted into separate expression vectors, and are co-transfected into a suitable host organism. For further details of illustrative currently known methods for generating bispecific antibodies see, e.g., Suresh et al. (1986) Methods Enzymol 121:210-228; PCT Publication No. WO 96/27011; Brennan et al. (1985) Science 229:81-83; Shalaby et al. JExp Med (1992) 175:217-225; Kostelny et al. (1992) *J Immunol* 148(5) 1547-1553; Hollinger et al. (1993) Proc Natl Acad Sci USA 90:6444-6448; Gruber et al. (1994) J Immunol 152:5368-5474; and Tutt et al. (1991) J Immunol 147:60-69. Bispecific antibodies also include cross-linked or heteroconjugate antibodies. Heteroconjugate antibodies may be made using any convenient cross-linking methods. Suitable cross-linking agents are well known in the art, and are disclosed in U.S. Pat. No. 4,676,980, along with a number of cross-linking techniques.

Various techniques for making and isolating bispecific antibody fragments directly from recombinant cell culture have also been described. For example, bispecific antibodies have been produced using leucine zippers. See, e.g., Kostelny et al. (1992) J Immunol 148(5):1547-1553. The leucine zipper peptides from the Fos and Jun proteins may be linked to the Fab' portions of two different antibodies by gene fusion. The antibody homodimers may be reduced at the hinge region to form monomers and then re-oxidized to form the antibody heterodimers. This method can also be utilized for the production of antibody homodimers. The "diabody" technology described by Hollinger et al. (1993) Proc Natl Acad Sci USA 90:6444-6448 has provided an alternative mechanism for making bispecific antibody fragments. The fragments comprise a heavy-chain variable domain (VH) connected to a light-chain variable domain (VL) by a linker which is too short to allow pairing between the two domains on the same chain. Accordingly, the VH and VL domains of one fragment are forced to pair with the complementary VL and VH domains of another fragment, thereby forming two antigenbinding sites. Another strategy for making bispecific antibody fragments by the use of single-chain Fv (scFv) dimers has

also been reported. See, e.g., Gruber et al. (1994) *J Immunol* 152:5368-5374. Alternatively, the antibodies can be "linear antibodies" as described in, e.g., Zapata et al. (1995) *Protein Eng* 8(10): 1057-1062. Briefly, these antibodies comprise a pair of tandem Fd segments (V_H — C_H 1- V_H - C_H 1) which form a pair of antigen binding regions. Linear antibodies can be bispecific or monospecific.

The disclosure also embraces variant forms of bispecific antibodies such as the tetravalent dual variable domain immunoglobulin (DVD-Ig) molecules described in Wu et al. (2007) 10 Nat Biotechnol 25(11: 1290-1297. The DVD-Ig molecules are designed such that two different light chain variable domains (VL) from two different parent antibodies are linked in tandem directly or via a short linker by recombinant DNA techniques, followed by the light chain constant domain. 15 Methods for generating DVD-Ig molecules from two parent antibodies are further described in, e.g., PCT Publication Nos. WO 08/024188 and WO 07/024715, the disclosures of each of which are incorporated herein by reference in their entirety.

Methods for Treatment

The above-described compositions (e.g., any of the high concentration antibody solutions) are useful in, inter alia, methods for treating or preventing a variety of complement-associated disorders (e.g., AP-associated disorders or CP-25 associated disorders) in a subject. The compositions can be administered to a subject, e.g., a human subject, using a variety of methods that depend, in part, on the route of administration. The route can be, e.g., intravenous injection or infusion (IV), subcutaneous injection (SC), intraperitoneal (IP) 30 injection, intraocular injection, intraarticular injection, or intramuscular injection (IM).

In some embodiments, a high concentration antibody solution described herein is therapeutically delivered to a subject by way of local administration. As used herein, "local administration" or "local delivery," refers to delivery that does not rely upon transport of the composition or active agent (e.g., an anti-C5 antibody) to its intended target tissue or site via the vascular system. For example, the composition may be delivered by injection or implantation of the composition or agent or by injection or implantation of a device containing the composition or agent. Following local administration in the vicinity of a target tissue or site, the composition or agent, or one or more components thereof, may diffuse to the intended target tissue or site.

In some embodiments, a high concentration antibody solution can be locally administered to a joint (e.g., an articulated joint). For example, in embodiments where the complement-associated disorder is arthritis, the solution can be administered directly to a joint (e.g., into a joint space) or in the 50 vicinity of a joint. Examples of intraarticular joints to which a high concentration antibody solution described herein can be locally administered include, e.g., the hip, knee, elbow, wrist, sternoclavicular, temperomandibular, carpal, tarsal, ankle, and any other joint subject to arthritic conditions. A 55 high concentration solution described herein can also be administered to bursa such as, e.g., acromial, bicipitoradial, cubitoradial, deltoid, infrapatellar, ischial, and any other bursa known in the art of medicine.

In some embodiments, a high concentration antibody solution described herein can be locally administered to the eye, e.g., to treat patients afflicted with a complement-associated disorder of the eye such as wet or dry AMD. As used herein, the term "eye" refers to any and all anatomical tissues and structures associated with an eye. The eye has a wall composed of three distinct layers: the outer sclera, the middle choroid layer, and the inner retina. The chamber behind the

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lens is filled with a gelatinous fluid referred to as the vitreous humor. At the back of the eye is the retina, which detects light. The cornea is an optically transparent tissue, which conveys images to the back of the eye. The cornea includes one pathway for the permeation of drugs into the eye. Other anatomical tissue structures associated with the eye include the lacrimal drainage system, which includes a secretory system, a distributive system and an excretory system. The secretory system comprises secretors that are stimulated by blinking and temperature change due to tear evaporation and reflex secretors that have an efferent parasympathetic nerve supply and secrete tears in response to physical or emotional stimulation. The distributive system includes the eyelids and the tear meniscus around the lid edges of an open eye, which spread tears over the ocular surface by blinking, thus reducing dry areas from developing.

In some embodiments, a high concentration antibody solution described herein is administered to the posterior chamber of the eye. In some embodiments, a high concentration antibody solution is administered intravitreally. In some embodiments, a high concentration antibody solution described herein is administered transsclerally.

It is understood that in some embodiments a high concentration antibody solution described herein can be administered systemically for use in treating, e.g., RA, wet or dry AMD, or any other complement-associated disorder described herein.

A suitable dose of a high concentration antibody solution described herein, which dose is capable of treating or preventing a complement-associated disorder in a subject, can depend on a variety of factors including, e.g., the age, sex, and weight of a subject to be treated and the particular inhibitor compound used. For example, a different dose of an anti-C5 antibody (and thus a different concentration of the antibody in solution or a different volume of a high concentration antibody solution) may be required to treat an elderly subject with RA as compared to the dose of an anti-C5 antibody that is required to treat a younger subject. Other factors affecting the dose administered to the subject include, e.g., the type or severity of the complement-associated disorder. For example, a subject having RA may require administration of a different dosage of a high concentration antibody solution described herein than a subject with AMD. Other factors can include, e.g., other medical disorders concurrently or previously 45 affecting the subject, the general health of the subject, the genetic disposition of the subject, diet, time of administration, rate of excretion, drug combination, and any other additional therapeutics that are administered to the subject. It should also be understood that a specific dosage and treatment regimen for any particular subject is generally governed by the judgment of the treating medical practitioner (e.g., doctor or nurse).

An anti-C5 antibody as part of a high concentration antibody solution described herein can be administered as a fixed dose, or in a milligram per kilogram (mg/kg) dose. In some embodiments where local administration is preferred, a dose can be selected that results in local inhibition of C5 cleavage (through the action of the anti-C5 antibody), but with no substantial effect on systemic complement activity. In some embodiments, the dose can also be chosen to reduce or avoid production of antibodies or other host immune responses against the therapeutic antibodies in the composition. While in no way intended to be limiting, exemplary dosages of an anti-C5 antibody locally administered to the eye of a patient afflicted with AMD include 0.5 mg to 5 mg (e.g., at least 0.5, 0.6, 0.7, 0.8, 0.9, 1, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 2.1, 2.2, 2.3, 2.4, 2.5, 2.6, 2.7, 2.8, 2.9, 3, 3.1, 3.2, 3.3, 3.4, 3.5,

3.6, 3.7, 3.8, 3.9, 4, 4.1, 4.2, 4.3, 4.4, 4.5, 4.6, 4.7, 4.8, 4.9. or 5 mg) per dose. The dose, or pharmaceutical unit dosage form, can be provided to the eye of the patient in a volume of, e.g., up to 50 (e.g., one, two, three, four, five, six, seven, eight, nine, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 5 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, or 50) microliters. Accordingly, the disclosure embraces and features pharmaceutical unit dosage forms of an anti-C5 antibody for use in treating AMD (e.g., wet or dry AMD) in a patient by intravitreal injection, 10 which dosage form includes between 0.5 mg to 5 mg, inclusive in a volume of not more than 50 microliters.

While in no way intended to be limiting, exemplary dosages of an anti-C5 antibody locally administered to a joint (e.g., an articulated joint) of a patient afflicted with rheuma- 15 toid arthritis (RA) include 0.5 to 10 mg (e.g., at least 0.5, 0.6, 0.7, 0.8, 0.9, 1, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 2.1,2.2, 2.3, 2.4, 2.5, 2.6, 2.7, 2.8, 2.9, 3, 3.1, 3.2, 3.3, 3.4, 3.5, 3.6, 3.7, 3.8, 3.9, 4, 4.1, 4.2, 4.3, 4.4, 4.5, 4.6, 4.7, 4.8, 4.9, 5, 5.1, 5.2, 5.3, 5.4, 5.5, 5.6, 5.7, 5.8, 5.9, 6, 6.1, 6.2, 6.3, 6.4, 6.5, 6.6, 20 6.7, 6.8, 6.9, 7, 7.1, 7.2, 7.3, 7.4, 7.5, 7.6, 7.7, 7.8, 7.9, 8, 8.1, 8.2, 8.3, 8.4, 8.5, 8.6, 8.7, 8.8, 8.9, 9, 9.1, 9.2, 9.3, 9.4, 9.5, 9.6, 9.7, 9.8, 9.9, or 10 mg) per dose. The dose, or pharmaceutical unit dosage form, can be provided to a joint of the patient in a volume of, e.g., up to 500 (e.g., one, two, three, four, five, 25 six, seven, eight, nine, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, 200, 210, 220, 230, 240, 250, 260, 270, 280, 290, 300, 30 310, 320, 330, 340, 350, 360, 370, 380, 390, 400, 410, 420, 430, 440, 450, 460, 470, 480, 490, or 500) microliters. Accordingly, the disclosure embraces and features pharmaceutical unit dosage forms of an anti-C5 antibody for use in treating RA in a patient by intraarticular injection, which 35 dosage form includes between 0.5 mg to 10 mg, inclusive in a volume of not more than 500 microliters.

Methods for detecting systemic hemolytic activity, as well as inhibition of said activity, are well known in the art and are described herein.

In some embodiments, the concentrated solution of an anti-C5 antibody can be diluted into a pharmaceutically-acceptable diluent for, e.g., systemic delivery of the antibody to the subject. While in no way intended to be limiting, exemplary dosages of an anti-C5 antibody to be administered systemically to treat complement-associated disorder include, e.g., 1-100 μg/kg, 0.5-50 μg/kg, 0.1-100 μg/kg, 0.5-25 μg/kg, 1-20 μg/kg, and 1-10 μg/kg, 1-20 μg/kg, and 1-10 mg/kg. Exemplary dosages of an antibody described herein include, without limitation, 0.1 μg/kg, 0.5 μg/kg, 1.0 μg/kg, 2.0 μg/kg, 4 μg/kg, and 8 μg/kg, 0.1 mg/kg, 0.5 mg/kg, 1.0 mg/kg, 2.0 mg/kg, 4 mg/kg, and 8 mg/kg.

A therapeutically-effective amount of an anti-C5 antibody described herein can be readily determined by one of ordinary 55 skill in the art based, in part, on the effect of the administered antibody, or the combinatorial effect of the antibody and one or more additional active agents, if more than one agent is used. A therapeutically effective amount of an antibody described herein can also vary according to factors such as the 60 disease state, age, sex, and weight of the individual, and the ability of the antibody (and one or more additional active agents) to elicit a desired response in the individual, e.g., amelioration of at least one condition parameter, e.g., amelioration of at least one symptom of the complement-associated disorder. For example, a therapeutically effective amount of a C5-binding polypeptide can inhibit (lessen the

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severity of or eliminate the occurrence of) and/or prevent a particular disorder, and/or any one of the symptoms of the particular disorder known in the art or described herein. A therapeutically effective amount is also one in which any toxic or detrimental effects of the composition are outweighed by the therapeutically beneficial effects.

Suitable human doses of any of the anti-C5 antibodies described herein can further be evaluated in, e.g., Phase I dose escalation studies. See, e.g., van Gurp et al. (2008) *Am J Transplantation* 8(8): 1711-1718; Hanouska et al. (2007) *Clin Cancer Res* 13(2, part 1):523-531; and Hetherington et al. (2006) *Antimicrobial Agents and Chemotherapy* 50(10): 3499-3500.

The terms "therapeutically effective amount" or "therapeutically effective dose," or similar terms used herein are intended to mean an amount of an agent (e.g., an anti-C5 antibody) that will elicit the desired biological or medical response (e.g., an improvement in one or more symptoms of a complement-associated disorder). In some embodiments, a high concentration antibody solution described herein contains a therapeutically effective amount of the anti-C5 antibody. In some embodiments, the high concentration antibody solution described herein contains an anti-C5 antibody and one or more (e.g., one, two, three, four, five, six, seven, eight, nine, 10, or 11 or more) additional therapeutic agents such that the composition as a whole is therapeutically effective. For example, a high concentration antibody solution can contain anti-C5 antibody and a VEGF inhibitor (e.g., an anti-VEGF antibody such as bevacizumab), wherein the anti-C5 antibody and VEGF inhibitor are each at a concentration that when combined are therapeutically effective for treating or preventing a complement-associated disorder in a subject.

Toxicity and therapeutic efficacy of such compositions can be determined by known pharmaceutical procedures in cell cultures or experimental animals (e.g., animal models of any of the complement-associated disorders described herein). These procedures can be used, e.g., for determining the $\rm LD_{50}$ (the dose lethal to 50% of the population) and the $\rm ED_{50}$ (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index and it can be expressed as the ratio $\rm LD_{50}/ED_{50}$. An anti-C5 antibody that exhibits a high therapeutic index is preferred. While compositions that exhibit toxic side effects may be used, care should be taken to design a delivery system that targets such compounds to the site of affected tissue and to minimize potential damage to normal cells and, thereby, reduce side effects.

The data obtained from the cell culture assays and animal studies can be used in formulating a range of dosage for use in humans. The dosage of such antibodies lies generally within a range of circulating concentrations of the anti-C5 antibody that include the ED₅₀ with little or no toxicity. The dosage may vary within this range depending upon the dosage form employed and the route of administration utilized. For an anti-C5 antibody used as described herein (e.g., for treating or preventing a complement-associated disorder), the therapeutically effective dose can be estimated initially from cell culture assays. A dose can be formulated in animal models to achieve a circulating plasma concentration range that includes the IC₅₀ (i.e., the concentration of the test compound which achieves a half-maximal inhibition of symptoms) as determined in cell culture. Such information can be used to more accurately determine useful doses in humans. Levels in plasma may be measured, for example, by high performance liquid chromatography or by ELISA.

In some embodiments, the methods can be performed in conjunction with other therapies for complement-associated

disorders. For example, the composition can be administered to a subject at the same time, prior to, or after, plasmapheresis, IVIG therapy, plasma replacement, or plasma exchange. See, e.g., Appel et al. (2005) *J Am Soc Nephrol* 16:1392-1404. In some embodiments, a high concentration antibody solution 5 described herein is not administered in conjunction with IVIG. In some embodiments, e.g., for patients with aHUS, the composition can be administered to a subject at the same time, prior to, or after, a kidney transplant.

A "subject," as used herein, can be any mammal. For 10 example, a subject can be a human, a non-human primate (e.g., monkey, baboon, or chimpanzee), a horse, a cow, a pig, a sheep, a goat, a dog, a cat, a rabbit, a guinea pig, a gerbil, a hamster, a rat, or a mouse. In some embodiments, the subject is an infant (e.g., a human infant).

As used herein, a subject "in need of prevention," "in need of treatment," or "in need thereof," refers to one, who by the judgment of an appropriate medical practitioner (e.g., a doctor, a nurse, or a nurse practitioner in the case of humans; a veterinarian in the case of non-human mammals), would reasonably benefit from a given treatment (such as treatment with a solution comprising a high concentration of an anti-C5 antibody).

As described above, the high concentration antibody solutions described herein can be used to treat a variety of comple- 25 ment-associated disorders such as, e.g., AP-associated disorders and/or CP-associated disorders. Such disorders include, without limitation, rheumatoid arthritis (RA); antiphospholipid antibody syndrome; lupus nephritis; ischemia-reperfusion injury; atypical hemolytic uremic syndrome (aHUS); 30 typical or infectious hemolytic uremic syndrome (tHUS); dense deposit disease (DDD); paroxysmal nocturnal hemoglobinuria (PNH); neuromyelitis optica (NMO); multifocal motor neuropathy (MMN); multiple sclerosis (MS); macular degeneration (e.g., age-related macular degeneration 35 (AMD)); hemolysis, elevated liver enzymes, and low platelets (HELLP) syndrome; thrombotic thrombocytopenic purpura (TTP); spontaneous fetal loss; Pauci-immune vasculitis; epidermolysis bullosa; recurrent fetal loss; and traumatic brain injury. (See, e.g., Holers (2008) Immunological 40 Reviews 223:300-316 and Holers and Thurman (2004) Molecular Immunology 41:147-152.) In some embodiments, the complement-associated disorder is a complement-associated vascular disorder such as, but not limited to, a diabetesassociated vascular disorder (e.g., of the eye), central retinal 45 vein occlusion, a cardiovascular disorder, myocarditis, a cerebrovascular disorder, a peripheral (e.g., musculoskeletal) vascular disorder, a renovascular disorder, a mesenteric/enteric vascular disorder, revascularization to transplants and/or replants, vasculitis, Henoch-Schonlein purpura nephritis, 50 systemic lupus erythematosus-associated vasculitis, vasculitis associated with rheumatoid arthritis, immune complex vasculitis, Takayasu's disease, dilated cardiomyopathy, diabetic angiopathy, Kawasaki's disease (arteritis), venous gas embolus (VGE), and restenosis following stent placement, 55 rotational atherectomy, and percutaneous transluminal coronary angioplasty (PTCA). (See, e.g., U.S. patent application publication no. 20070172483.) Additional complement-associated disorders include, without limitation, myasthenia gravis, cold agglutinin disease, dermatomyositis, Graves' disease, atherosclerosis, Alzheimer's disease, Guillain-Barré Syndrome, Degos' disease, graft rejection (e.g., transplant rejection), sepsis, burn (e.g., severe burn), systemic inflammatory response sepsis, septic shock, spinal cord injury, glomerulonephritis, Hashimoto's thyroiditis, type I diabetes, 65 psoriasis, pemphigus, autoimmune hemolytic anemia (AIHA), idiopathic thrombocytopenic purpura (ITP), Good42

pasture syndrome, antiphospholipid syndrome (APS), and catastrophic APS (CAPS). In some embodiments, the high concentration antibody solutions described herein can be used in methods for treating thrombotic microangiopathy (TMA), e.g., TMA associated with a complement-associated disorder such as any of the complement-associated disorders described herein.

Complement-associated disorders also include complement-associated pulmonary disorders such as, but not limited to, asthma, bronchitis, a chronic obstructive pulmonary disease (COPD), an interstitial lung disease, α -1 anti-trypsin deficiency, emphysema, bronchiectasis, bronchiolitis obliterans, alveolitis, sarcoidosis, pulmonary fibrosis, and collagen vascular disorders.

As used herein, a subject "at risk for developing a complement-associated disorder" (e.g., an AP-associated disorder or a CP-associated disorder) is a subject having one or more (e.g., two, three, four, five, six, seven, or eight or more) risk factors for developing the disorder. Risk factors will vary depending on the particular complement-associated disorder, but are well known in the art of medicine. For example, risk factors for developing DDD include, e.g., a predisposition to develop the condition, i.e., a family history of the condition or a genetic predisposition to develop the condition such as, e.g., one or more mutations in the gene encoding complement factor H (CFH), complement factor H-related 5 (CFHR5), and/or complement component C3 (C3). Such DDD-associated mutations as well methods for determining whether a subject carries one or more of the mutations are known in the art and described in, e.g., Licht et al. (2006) Kidnev Int 70:42-50; Zipfel et al. (2006) "The role of complement in membranoproliferative glomerulonephritis," In: Complement and Kidney Disease, Springer, Berlin, pages 199-221; Ault et al. (1997) J Biol Chem 272:25168-75; Abrera-Abeleda et al. (2006) J Med Genet 43:582-589; Poznansky et al. (1989) J Immunol 143:1254-1258; Jansen et al. (1998) Kidnev Int 53:331-349; and Hegasy et al. (2002) Am J Pathol 161:2027-2034. Thus, a human at risk for developing DDD can be, e.g., one who has one or more DDD-associated mutations in the gene encoding CFH or one with a family history of developing the disease.

Risk factors for TTP are well known in the art of medicine and include, e.g., a predisposition to develop the condition, i.e., a family history of the condition or a genetic predisposition to develop the condition such as, e.g., one or more mutations in the ADAMTS13 gene. ADAMTS13 mutations associated with TTP are reviewed in detail in, e.g., Levy et al. (2001) Nature 413:488-494; Kokame et al. (2004) Semin Hematol 41:34-40; Licht et al. (2004) Kidney Int 66:955-958; and Noris et al. (2005) J Am Soc Nephrol 16:1177-1183. Risk factors for TTP also include those conditions or agents that are known to precipitate TTP, or TTP recurrence, such as, but not limited to, cancer, bacterial infections (e.g., Bartonella sp. infections), viral infections (e.g., HIV and Kaposi's sarcoma virus), pregnancy, or surgery. See, e.g., Avery et al. (1998) Am J Hematol 58:148-149 and Tsai, supra. TTP, or recurrence of TTP, has also been associated with the use of certain therapeutic agents (drugs) including, e.g., ticlopidine, FK 506, corticosteroids, tamoxifen, or cyclosporin A (see, e.g., Gordon et al. (1997) Sem in Hematol 34(2):140-147). Hereinafter, such manifestations of TTP may be, where appropriate, referred to as, e.g., "infection-associated TTP," "pregnancy-associated TTP," or "drug-associated TTP." Thus, a human at risk for developing TTP can be, e.g., one who has one or more TTPassociated mutations in the ADAMTS13 gene. A human at

risk for developing a recurrent form of TTP can be one, e.g., who has had TTP and has an infection, is pregnant, or is undergoing surgery.

Risk factors for aHUS are well known in the art of medicine and include, e.g., a predisposition to develop the condition, i.e., a family history of the condition or a genetic predisposition to develop the condition such as, e.g., one or more mutations in complement Factor H (CFH), membrane cofactor protein (MCP; CD46), C4b-binding protein, complement factor B (CFB), or complement factor I (CFI). (See, e.g., Warwicker et al. (1998) Kidney Int 53:836-844; Richards et al. (2001) Am J Hum Genet 68:485-490; Caprioli et al. (2001) Am Soc Nephrol 12:297-307; Neuman et al. (2003) J Med Genet 40:676-681; Richards et al. (2006) Proc Natl Acad Sci USA 100:12966-12971; Fremeaux-Bacchi et al. (2005) J Am Soc Nephrol 17:2017-2025; Esparza-Gordillo et al. (2005) Hum Mol Genet 14:703-712; Goicoechea de Jorge et al. (2007) Proc Natl Acad Sci USA 104(1):240-245; Blom et al. (2008) J Immunol 180(9):6385-91; and Fremeaux-Bacchi et 20 al. (2004) J Medical Genet 41:e84). (See also Kavanagh et al. (2006) supra.) Risk factors also include, e.g., infection with Streptococcus pneumoniae, pregnancy, cancer, exposure to anti-cancer agents (e.g., quinine, mitomycin C, cisplatin, or bleomycin), exposure to immunotherapeutic agents (e.g., 25 cyclosporine, OKT3, or interferon), exposure to anti-platelet agents (e.g., ticlopidine or clopidogrel), HIV infection, transplantation, autoimmune disease, and combined methylmalonic aciduria and homocystinuria (cblC). See, e.g., Constantinescu et al. (2004) Am J Kidney Dis 43:976-982; George 30 (2003) Curr Opin Hematol 10:339-344; Gottschall et al. (1994) Am J Hematol 47:283-289; Valavaara et al. (1985) Cancer 55:47-50; Miralbell et al. (1996) J Clin Oncol 14:579-585; Dragon-Durey et al. (2005) J Am Soc Nephrol 16:555-63; and Becker et al. (2004) Clin Infect Dis 39:S267-S275.

Risk factors for HELLP are well known in the art of medicine and include, e.g., multiparous pregnancy, maternal age over 25 years, Caucasian race, the occurrence of preeclampsia or HELLP in a previous pregnancy, and a history of poor pregnancy outcome. (See, e.g., Sahin et al. (2001) Nagoya 40 Med J 44(3):145-152; Sullivan et al. (1994) Am J Obstet Gynecol 171:940-943; and Padden et al. (1999) Am Fam Physician 60(3):829-836.) For example, a pregnant, Caucasian woman who developed preeclampsia during a first pregnancy can be one at risk for developing HELLP syndrome 45 during, or following, a second pregnancy.

Risk factors for CAD are well known in the art of medicine and include, e.g., conditions or agents that are known to precipitate CAD, or CAD recurrence, such as, but not limited to, neoplasms or infections (e.g., bacterial and viral infec- 50 tions). Conditions known to be associated with the development of CAD include, e.g., HIV infection (and AIDS), hepatitis C infection, Mycoplasma pneumonia infection, Epstein-Barr virus (EBV) infection, cytomegalovirus (CMV) infection, rubella, or infectious mononucleosis. Neoplasms 55 associated with CAD include, without limitation, non-Hodgkin's lymphoma. Hereinafter, such manifestations of CAD may be, where appropriate, referred to as, e.g., "infection-associated CAD" or "neoplasm-associated CAD." Thus, a human at risk for developing CAD can be, e.g., one who has 60 an HIV infection, rubella, or a lymphoma. See also, e.g., Gertz (2006) *Hematology* 1:19-23; Horwitz et al. (1977) Blood 50:195-202; Finland and Barnes (1958) AMA Arch Intern Med 191:462-466; Wang et al. (2004) Acta Paediatr Taiwan 45:293-295; Michaux et al. (1998) Ann Hematol 65 76:201-204; and Chang et al. (2004) Cancer Genet Cytogenet 152:66-69.

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Risk factors for myasthenia gravis (MG) are well known in the art of medicine and include, e.g., a predisposition to develop the condition, i.e., a family history of the condition or a genetic predisposition to develop the condition such as familial MG. For example, some HLA types are associated with an increased risk for developing MG. Risk factors for MG include the ingestion or exposure to certain MG-inducing drugs such as, but not limited to, D-penicillamine. See, e.g., Drosos et al. (1993) Clin Exp Rheumatol 11(4):387-91 and Kaeser et al. (1984) Acta Neurol Scand Suppl 100:39-47. As MG can be episodic, a subject who has previously experienced one or more symptoms of having MG can be at risk for relapse. Thus, a human at risk for developing MG can be, e.g., one who has a family history of MG and/or one who has ingested or been administered an MG-inducing drug such as D-penicillamine.

As used herein, a subject "at risk for developing CAPS" is a subject having one or more (e.g., two, three, four, five, six, seven, or eight or more) risk factors for developing the disorder. Approximately 60% of the incidences of CAPS are preceded by a precipitating event such as an infection. Thus, risk factors for CAPS include those conditions known to precipitate CAPS such as, but not limited to, certain cancers (e.g., gastric cancer, ovarian cancer, lymphoma, leukemia, endometrial cancer, adenocarcinoma, and lung cancer), pregnancy, puerperium, transplantation, primary APS, rheumatoid arthritis (RA), systemic lupus erythematosus (SLE), surgery (e.g., eye surgery), and certain infections. Infections include, e.g., parvovirus B19 infection and hepatitis C infection. Hereinafter, such manifestations of CAPS may be referred to as, e.g., "cancer-associated CAPS," "transplantation-associated CAPS," "RA-associated CAPS," "infectionassociated CAPS," or "SLE-associated CAPS." See, e.g., Soltész et al. (2000) Haematologia (Budep) 30(4):303-311; Ideguchi et al. (2007) Lupus 16(1):59-64; Manner et al. (2008) Am J Med Sci 335(5):394-7; Miesbach et al. (2006) Autoimmune Rev 6(2):94-7; Gómez-Puerta et al. (2006) Autoimmune Rev 6(2):85-8; Gómez-Puerta et al. (2006) Semin Arthritis Rheum 35(5):322-32; Kasamon et al. (2005) Haematologia 90(3):50-53; Atherson et al. (1998) Medicine 77(3):195-207; and Canpolat et al. (2008) Clin Pediatr 47(6): 593-7. Thus, a human at risk for developing CAPS can be, e.g., one who has primary CAPS and/or a cancer that is known to be associated with CAPS.

From the above it will be clear that subjects "at risk for developing a complement-associated disorder" (e.g., an AP-associated disorder or a CP-associated disorder) are not all the subjects within a species of interest.

A subject "suspected of having a complement-associated disorder" (e.g., an alternative complement pathway-associated disorder) is one having one or more (e.g., two, three, four, five, six, seven, eight, nine, or 10 or more) symptoms of the disease. Symptoms of these disorders will vary depending on the particular disorder, but are known to those of skill in the art of medicine. For example, symptoms of DDD include, e.g.: one or both of hematuria and proteinuria; acute nephritic syndrome; drusen development and/or visual impairment; acquired partial lipodystrophy and complications thereof; and the presence of serum C3 nephritic factor (C3NeF), an autoantibody directed against C3bBb, the C3 convertase of the alternative complement pathway. (See, e.g., Appel et al. (2005), supra). Symptoms of aHUS include, e.g., severe hypertension, proteinuria, uremia, lethargy/fatigue, irritability, thrombocytopenia, microangiopathic hemolytic anemia, and renal function impairment (e.g., acute renal failure). Symptoms of TTP include, e.g., microthrombi, thrombocytopenia, fever, low ADAMTS13 metalloproteinase expres-

sion or activity, fluctuating central nervous system abnormalities, renal failure, microangiopathic hemolytic anemia, bruising, purpura, nausea and vomiting (e.g., resulting from ischemia in the GI tract or from central nervous system involvement), chest pain due to cardiac ischemia, seizures, 5 and muscle and joint pain. Symptoms of RA can include, e.g., stiffness, swelling, fatigue, anemia, weight loss, fever, and often, crippling pain. Some common symptoms of rheumatoid arthritis include joint stiffness upon awakening that lasts an hour or longer; swelling in a specific finger or wrist joints; swelling in the soft tissue around the joints; and swelling on both sides of the joint. Swelling can occur with or without pain, and can worsen progressively or remain the same for years before progressing. Symptoms of HELLP are known in the art of medicine and include, e.g., malaise, epigastric pain, 15 nausea, vomiting, headache, right upper quadrant pain, hypertension, proteinuria, blurred vision, gastrointestinal bleeding, hypoglycemia, paresthesia, elevated liver enzymes/ liver damage, anemia (hemolytic anemia), and low platelet count, any of which in combination with pregnancy or recent 20 pregnancy. (See, e.g., Tomsen (1995) Am J Obstet Gynecol 172:1876-1890; Sibai (1986) Am J Obstet Gynecol 162:311-316; and Padden (1999), supra.) Symptoms of PNH include, e.g., hemolytic anemia (a decreased number of red blood cells), hemoglobinuria (the presence of hemoglobin in the 25 urine particularly evident after sleeping), and hemoglobin-

Symptoms of CAPS are well known in the art of medicine and include, e.g., histopathological evidence of multiple small vessel occlusions; the presence of antiphospholipid antibodies (usually at high titer), vascular thromboses, severe 35 multi-organ dysfunction, malignant hypertension, acute respiratory distress syndrome, disseminated intravascular coagulation, microangiopathic hemolytic anemia, schistocytes, and thrombocytopenia. CAPS can be distinguished from APS in that patients with CAPS generally present with 40 severe multiple organ dysfunction or failure, which is characterized by rapid, diffuse small vessel ischemia and thromboses predominantly affecting the parenchymal organs. In contrast, APS is associated with single venous or arterial medium-to-large blood vessel occlusions. Symptoms of MG 45 include, e.g., fatigability and a range of muscle weaknessrelated conditions including: ptosis (of one or both eyes), diplopia, unstable gait, depressed or distorted facial expressions, and difficulty chewing, talking, or swallowing. In some instances, a subject can present with partial or complete 50 paralysis of the respiratory muscles. Symptoms of CAD include, e.g., pain, fever, pallor, anemia, reduced blood flow to the extremities (e.g., with gangrene), and renal disease or acute renal failure. In some embodiments, the symptoms can occur following exposure to cold temperatures.

emia (the presence of hemoglobin in the bloodstream). PNH-

afflicted subjects are known to have paroxysms, which are

defined here as incidences of dark-colored urine, dysphagia,

abdominal pain.

fatigue, erectile dysfunction, thrombosis, and recurrent 30

From the above it will be clear that subjects "suspected of having a complement-associated disorder" are not all the subjects within a species of interest.

In some embodiments, the methods can include identifying the subject as one having, suspected of having, or at risk for 60 developing, a complement-associated disorder in a subject. Suitable methods for identifying the subject are known in the art. For example, suitable methods (e.g., sequencing techniques or use of microarrays) for determining whether a human subject has a DDD-associated mutation in a CFH, 65 CFHR5, or C3 gene are described in, e.g., Licht et al. (2006) *Kidney Int* 70:42-50; Zipfel et al. (2006), supra; Ault et al.

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(1997) J Biol Chem 272:25168-75; Abrera-Abeleda et al. (2006) J Med Genet 43:582-589; Poznansky et al. (1989) J Immunol 143:1254-1258; Jansen et al. (1998) Kidney Int 53:331-349; and Hegasy et al. (2002) Am J Pathol 161:2027-2034. Methods for detecting the presence of characteristic DDD-associated electron-dense deposits are also well known in the art. For example, a medical practitioner can obtain a tissue biopsy from the kidney of a patient and subject the tissue to electron microscopy. The medical practitioner may also examine the tissue by immunofluorescence to detect the presence of C3 using an anti-C3 antibody and/or light microscopy to determine if there is membranoproliferative glomerulonephritis. See, e.g., Walker et al. (2007) Mod Pathol 20:605-616 and Habib et al. (1975) Kidney Int 7:204-215. In some embodiments, the identification of a subject as one having DDD can include assaying a blood sample for the presence of C3NeF. Methods for detecting the presence of C3NeF in blood are described in, e.g., Schwertz et al. (2001) Pediatr Allergy Immunol 12:166-172.

In some embodiments, the medical practitioner can determine whether there is increased complement activation in a subject's serum. Indicia of increased complement activation include, e.g., a reduction in CH50, a decrease in C3, and an increase in C3dg/C3d. See, e.g., Appel et al. (2005), supra. In some embodiments, a medical practitioner can examine a subject's eye for evidence of the development of drusen and/ or other visual pathologies such as AMD. For example, a medical practitioner can use tests of retinal function such as, but not limited to, dark adaptation, electroretinography, and electrooculography (see, e.g., Colville et al. (2003) *Am J Kidney Dis* 42:E2-5).

Methods for identifying a subject as one having, suspected of having, or at risk for developing, TTP are also known in the art. For example, Miyata et al. describe a variety of assays for measuring ADAMTS13 activity in a biological sample obtained from a subject (Curr Opin Hematol (2007) 14(3): 277-283). Suitable ADAMTS13 activity assays, as well as phenotypically normal ranges of ADAMTS13 activity in a human subject, are described in, e.g., Tsai (2003) J Am Soc Nephrol 14:1072-1081; Furlan et al. (1998) New Engl J Med 339:1578-1584; Matsumoto et al. (2004) Blood 103:1305-1310; and Mori et al. (2002) Transfusion 42:572-580. Methods for detecting the presence of inhibitors of ADAMTS13 (e.g., autoantibodies that bind to ADAMTS13) in a biological sample obtained from a subject are known in the art. For example, a serum sample from a patient can be mixed with a serum sample from a subject without TTP to detect the presence of anti-ADAMTS13 antibodies. In another example, immunoglobulin protein can be isolated from patient serum and used in in vitro ADAMTS13 activity assays to determine if an anti-ADAMTS13 antibody is present. See, e.g., Dong et al. (2008) Am J Hematol 83(10):815-817. In some embodiments, risk of developing TTP can be determined by assessing whether a patient carries one or more mutations in the 55 ADAMTS13 gene. Suitable methods (e.g., nucleic acid arrays or DNA sequencing) for detecting a mutation in the ADAMTS13 gene are known in the art and described in, e.g., Levy et al., supra; Kokame et al., supra; Licht et al., supra; and Noris et al., supra.

In addition, methods for identifying a subject as one having, suspected of having, or at risk for developing aHUS are known in the art. For example, laboratory tests can be performed to determine whether a human subject has thrombocytopenia, microangiopathic hemolytic anemia, or acute renal insufficiency. Thrombocytopenia can be diagnosed by a medical professional as one or more of: (i) a platelet count that is less than 150,000/mm³ (e.g., less than 60,000/mm³);

(ii) a reduction in platelet survival time, reflecting enhanced platelet disruption in the circulation; and (iii) giant platelets observed in a peripheral smear, which is consistent with secondary activation of thrombocytopoiesis. Microangiopathic hemolytic anemia can be diagnosed by a medical professional as one or more of: (i) hemoglobin concentrations that are less than 10 mg/dL (e.g., less than 6.5 mg/dL); (ii) increased serum lactate dehydrogenase (LDH) concentrations (>460 U/L); (iii) hyperbilirubinemia, reticulocytosis, circulating free hemoglobin, and low or undetectable haptoglobin concentrations; and (iv) the detection of fragmented red blood cells (schistocytes) with the typical aspect of burr or helmet cells in the peripheral smear together with a negative Coombs test. (See, e.g., Kaplan et al. (1992) "Hemolytic Uremic Syndrome and Thrombotic Thrombocytopenic Purpura," Informa Health Care (ISBN 0824786637) and Zipfel (2005) "Complement and Kidney Disease," Springer (ISBN 3764371668).)

A subject can also be identified as having aHUS by evaluating blood concentrations of C3 and C4 as a measure of complement activation or dysregulation. In addition, as is clear from the foregoing disclosure, a subject can be identified as having genetic aHUS by identifying the subject as harboring one or more mutations in a gene associated with 25 aHUS such as CFI, CFB, CFH, or MCP (supra). Suitable methods for detecting a mutation in a gene include, e.g., DNA sequencing and nucleic acid array techniques. (See, e.g., Breslin et al. (2006) Clin Am Soc Nephrol 1:88-99 and Goicoechea de Jorge et al. (2007) Proc Natl Acad Sci USA 104:240-30 245.)

Methods for diagnosing a subject as one having, suspected of having, or at risk for developing, RA are also known in the art of medicine. For example, a medical practitioner can examine the small joints of the hands, wrists, feet, and knees 35 to identify inflammation in a symmetrical distribution. The practitioner may also perform a number of tests to exclude other types of joint inflammation including arthritis due to infection or gout. In addition, rheumatoid arthritis is associated with abnormal antibodies in the blood circulation of 40 afflicted patients. For example, an antibody referred to as "rheumatoid factor" is found in approximately 80% of patients. In another example, anti-citrulline antibody is present in many patients with rheumatoid arthritis and thus it is useful in the diagnosis of rheumatoid arthritis when evalu- 45 ating patients with unexplained joint inflammation. See, e.g., van Venrooij et al. (2008) *Ann NY Acad Sci* 1143:268-285 and Habib et al. (2007) Immunol Invest 37(8):849-857. Another antibody called "the antinuclear antibody" (ANA) is also frequently found in patients with rheumatoid arthritis. See, 50 e.g., Benucci et al. (2008) Clin Rheumatol 27(1):91-95; Julkunen et al. (2005) Scan J Rheumatol 34(2): 122-124; and Miyawaki et al. (2005) J Rheumatol 32(8):1488-1494.

A medical practitioner can also examine red blood cell sedimentation rate to help in diagnosing RA in a subject. The 55 sedimentation rate can be used as a crude measure of the inflammation of the joints and is usually faster during disease flares and slower during remissions. Another blood test that can be used to measure the degree of inflammation present in the body is the C-reactive protein.

Furthermore, joint x-rays can also be used to diagnose a subject as having rheumatoid arthritis. As RA progresses, the x-rays can show bony erosions typical of rheumatoid arthritis in the joints. Joint x-rays can also be helpful in monitoring the progression of disease and joint damage over time. Bone scanning, a radioactive test procedure, can demonstrate the inflamed joints.

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Methods for identifying a subject as one having, suspected of having, or at risk for developing, HELLP are known in the art of medicine. Hallmark symptoms of HELLP syndrome include hemolysis, elevated liver enzymes, and low platelet count. Thus, a variety of tests can be performed on blood from a subject to determine the level of hemolysis, the concentration of any of a variety of liver enzymes, and the platelet level in the blood. For example, the presence of schistocytes and/or elevated free hemoglobin, bilirubin, or serum LDH levels is an indication of intravascular hemolysis. Routine laboratory testing can be used to determine the platelet count as well as the blood level of liver enzymes such as, but not limited to, aspartate aminotransferase (AST) and alanine transaminase (ALT). Suitable methods for identifying a subject as having HELLP syndrome are also described in, e.g., Sibai et al. (1993), supra; Martin et al. (1990), supra; Padden (1999), supra; and Gleicher and Buttino (1998) "Principles & Practice of Medical Therapy in Pregnancy," 3rd Edition, Appleton & Lange (ISBN 083857677X).

Methods for identifying a subject as having, suspected of having, or at risk for developing PNH are known in the art of medicine. The laboratory evaluation of hemolysis normally includes hematologic, serologic, and urine tests. Hematologic tests include an examination of the blood smear for morphologic abnormalities of red blood cells (RBC), and the measurement of the reticulocyte count in whole blood (to determine bone marrow compensation for RBC loss). Serologic tests include lactate dehydrogenase (LDH; widely performed), and free hemoglobin (not widely performed) as a direct measure of hemolysis. LDH levels, in the absence of tissue damage in other organs, can be useful in the diagnosis and monitoring of patients with hemolysis. Other serologic tests include bilirubin or haptoglobin, as measures of breakdown products or scavenging reserve, respectively. Urine tests include bilirubin, hemosiderin, and free hemoglobin, and are generally used to measure gross severity of hemolysis and for differentiation of intravascular vs. extravascular etiologies of hemolysis rather than routine monitoring of hemolysis. Further, RBC numbers, RBC hemoglobin, and hematocrit are generally performed to determine the extent of any accompanying anemia.

Suitable methods for identifying the subject as having MG can be qualitative or quantitative. For example, a medical practitioner can examine the status of a subject's motor functions using a physical examination. Other qualitative tests include, e.g., an ice-pack test, wherein an ice pack is applied to a subject's eye (in a case of ocular MG) to determine if one or more symptoms (e.g., ptosis) are improved by cold (see, e.g., Sethi et al. (1987) Neurology 37(8):1383-1385). Other tests include, e.g., the "sleep test," which is based on the tendency for MG symptoms to improve following rest. In some embodiments, quantitative or semi-quantitative tests can be employed by a medical practitioner to determine if a subject has, is suspected of having, or is at risk for developing, MG. For example, a medical practitioner can perform a test to detect the presence or amount of MG-associated autoantibodies in a serum sample obtained from a subject. MG-associated autoantibodies include, e.g., antibodies that bind to, and 60 modulate the activity of, acetylcholine receptor (AChR), muscle-specific receptor tyrosine kinase (MuSK), and/or striational protein. (See, e.g., Conti-Fine et al. (2006), supra). Suitable assays useful for detecting the presence or amount of an MG-associated antibody in a biological sample are known in the art and described in, e.g., Hoch et al. (2001) Nat Med 7:365-368; Vincent et al. (2004) Semin Neurol 24:125-133; McConville et al. (2004) Ann Neurol 55:580-584; Boneva et

al. (2006) *J Neuroimmunol* 177:119-131; and Romi et al. (2005) *Arch Neurol* 62:442-446.

Additional methods for diagnosing MG include, e.g., electrodiagnostic tests (e.g., single-fiber electromyography) and the Tensilon (or edrophonium) test, which involves injecting a subject with the acetylcholinesterase inhibitor edrophonium and monitoring the subject for an improvement in one or more symptoms. See, e.g., Pascuzzi (2003) *Semin Neurol* 23(1):83-88; Katirji et al. (2002) *Neurol Clin* 20:557-586; and "Guidelines in Electrodiagnostic Medicine. American Association of Electrodiagnostic Medicine," Muscle Nerve 15:229-253.

A subject can be identified as having CAD using an assay to detect the presence or amount (titer) of agglutinating autoantibodies that bind to the I antigen on red blood cells. 15 The antibodies can be monoclonal (e.g., monoclonal IgM or IgA) or polyclonal. Suitable methods for detecting these antibodies are described in, e.g., Christenson and Dacie (1957) Br J Haematol 3:153-164 and Christenson et al. (1957) Br J Haematol 3:262-275. A subject can also be diagnosed as 20 having CAD using one or more of a complete blood cell count (CBC), urinalysis, biochemical studies, and a Coombs test to test for hemolysis in blood. For example, biochemical studies can be used to detect elevated lactase dehydrogenase levels, elevated unconjugated bilirubin levels, low haptoglobin lev- 25 els, and/or the presence of free plasma hemoglobin, all of which can be indicative of acute hemolysis. Other tests that can be used to detect CAD include detecting complement levels in the serum. For example, due to consumption during the acute phase of hemolysis, measured plasma complement 30 levels (e.g., C2, C3, and C4) are decreased in CAD.

Typical (or infectious) HUS, unlike aHUS, is often identifiable by a prodrome of diarrhea, often bloody in nature, which results from infection with a shiga-toxin producing microorganism. A subject can be identified as having typical 35 HUS when shiga toxins and/or serum antibodies against shiga toxin or LPS are detected in the stool of an individual. Suitable methods for testing for anti-shiga toxin antibodies or LPS are known in the art. For example, methods for detecting antibodies that bind to shiga toxins Stx1 and Stx2 or LPS in 40 humans are described in, e.g., Ludwig et al. (2001) *J Clin Microbiol* 39(6):2272-2279.

In some embodiments, a high concentration antibody solution described herein can be administered to a subject as a monotherapy. Alternatively, as described above, the solution 45 can be administered to a subject as a combination therapy with another treatment, e.g., another treatment for DDD, TTP, wet or dry AMD, aHUS, PNH, RA, HELLP, MG, CAD, CAPS, tHUS, or any other complement-associated disorder known in the art or described herein. For example, the combination therapy can include administering to the subject (e.g., a human patient) one or more additional agents (e.g., anti-coagulants, anti-hypertensives, or corticosteroids) that provide a therapeutic benefit to the subject who has, or is at risk of developing, DDD. In some embodiments, the combi- 55 nation therapy can include administering to the subject (e.g., a human patient) by way of a high concentration antibody solution an anti-C5 antibody and an immunosuppressive agent such as Remicade® for use in treating RA. In some embodiments, a high concentration antibody solution and the 60 one or more additional active agents are administered at the same time. In other embodiments, a high concentration antibody solution is administered first in time and the one or more additional active agents are administered second in time. In some embodiments, the one or more additional active agents 65 are administered first in time and high concentration antibody solution is administered second in time.

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An anti-C5 antibody described herein can replace or augment a previously or currently administered therapy. For example, upon treating with an anti-C5 antibody, administration of the one or more additional active agents can cease or diminish, e.g., be administered at lower levels. In some embodiments, administration of the previous therapy can be maintained. In some embodiments, a previous therapy will be maintained until the level of the anti-C5 antibody reaches a level sufficient to provide a therapeutic effect. The two therapies can be administered in combination.

Monitoring a subject (e.g., a human patient) for an improvement in a complement-associated disorder, as defined herein, means evaluating the subject for a change in a disease parameter, e.g., an improvement in one or more symptoms of the disease (e.g., an improvement in one or more symptoms of a pulmonary disorder). Such symptoms include any of the symptoms of complement-associated disorders known in the art and/or described herein. In some embodiments, the evaluation is performed at least 1 hour, e.g., at least 2, 4, 6, 8, 12, 24, or 48 hours, or at least 1 day, 2 days, 4 days, 10 days, 13 days, 20 days or more, or at least 1 week, 2 weeks, 4 weeks, 10 weeks, 13 weeks, 20 weeks or more, after an administration. The subject can be evaluated in one or more of the following periods: prior to beginning of treatment; during the treatment; or after one or more elements of the treatment have been administered. Evaluating can include evaluating the need for further treatment, e.g., evaluating whether a dosage, frequency of administration, or duration of treatment should be altered. It can also include evaluating the need to add or drop a selected therapeutic modality, e.g., adding or dropping any of the treatments for any of the complementassociated disorders described herein.

Therapeutic Kits

The disclosure also features therapeutic kits containing, among other things, one or more of the high concentration solutions described herein. The therapeutic kits can contain, e.g., a suitable means for delivery of one or more solutions to a patient in need thereof, e.g., a patient afflicted with, suspected of having, or at risk for developing, a complementassociated disorder such as AMD (e.g., wet or dry AMD), a diabetes-associated ocular disorder, central retinal vein occlusion, RA, asthma, or any of the additional complementassociated disorders described herein. In some embodiments, the means is suitable for invasive (e.g., intravascular (e.g., intravenous), subcutaneous, intraarticular, intraocular, intravitreal, or intramuscular) delivery of the solution to a patient. In some embodiments, the means is suitable for subcutaneous delivery of the antibody or antigen-binding fragment thereof to the subject. For example, the means can be a syringe or an osmotic pump. In some embodiments, the solution contained in the kit can be formulated as an eye drop, the means being an eye dropper. In some embodiments, the kit contains a means that is pre-loaded with the solution to be administered. For example, a therapeutic kit can contain a syringe pre-filled with an aqueous solution (e.g., a pen device containing the solution) described herein or the kit can contain a pump (e.g., an osmotic pump) and one or more disposable cassettes configured for use with the pump, the cassettes pre-filled with an aqueous solution described herein. In another example, the kit can contain a transscleral or implantable delivery device (e.g., a plug) that is pre-filled with (or otherwise contains) a high concentration solution described herein.

In some embodiments, the means for delivering the high concentration solution is a pen device for drug delivery.

In some embodiments, the means can be suitable for administration of a high concentration antibody solution described herein to the eye of a patient afflicted with a

complement-associated disorder of the eye such as AMD. The means can be, e.g., a syringe, a transscleral patch, or even a contact lens containing or soaked in the solution. The means can, in some embodiments, be an eye dropper, wherein the solution is formulated for such administration. The means can 5 also be, e.g., a contact lens case in embodiments in which, e.g., the solution is formulated as part of a contact lens hydrating, cleaning, or soaking solution. Such therapeutic kits can also include, e.g., one or more additional therapeutic agents for use in treating complement-associated disorder of the eye. The therapeutic agents can be, e.g., bevacizumab or the Fab fragment of bevacizumab, ranibizumab, both sold by Roche Pharmaceuticals, Inc., pegaptanib sodium (Mucogen®; Pfizer, Inc.), and verteporfin (Visudyne®; Novartis). Such a 15 kit can also, optionally, include instructions for administering a solution described herein to a patient.

In some embodiments, the means can be suitable for intraarticular administration of a solution described herein to a patient in need thereof, e.g., a patient afflicted with complement-associated disorder affecting the joints such as RA. The means can be, e.g., a syringe or a double-barreled syringe. See, e.g., U.S. Pat. Nos. 6,065,645 and 6,698,622. A doublebarreled syringe is useful for administering to a joint two different compositions with only one injection. Two separate 25 syringes may be incorporated for use in administering the therapeutic while drawing off knee fluid for analysis (tapping) in a push-pull fashion. Additional therapeutic agents that can be administered with the high concentration antibody solutions described herein in conjunction with the doublebarreled syringe, or which can otherwise be generally included in the therapeutic kits described herein, include, e.g., NSAIDs, corticosteroids, methotrexate, hydroxychloroquine, anti-TNF agents such as etanercept and infliximab, a B cell depleting agent such as rituximab, an interleukin-1 antagonist, or a T cell costimulatory blocking agent such as abatacept. Such a kit can also, optionally, include instructions for administering a solution described herein to a patient.

In some embodiments, the means is suitable for intrapulmonary delivery of the solutions to a subject, e.g., for use in treatment or prevention of a complement-associated pulmonary disorder such as, but not limited to, COPD or asthma. Accordingly, the means can be, e.g., an oral or nasal inhaler (see above). The inhaler can be, e.g., a metered dose inhaler 45 (MDI) or a nebulizer. Such a kit can also, optionally, include instructions for administering (e.g., self-administration of) the anti-C5a antibody or antigen-binding fragment thereof to a subject. The therapeutic kits are designed for use in treating or preventing a complement-associated pulmonary disorder 50 and can include one or more additional active agents including, but not limited to, another antibody therapeutic (e.g., an anti-IgE antibody, an anti-IL-4 antibody, or an anti-IL-5 antibody), a small molecule anti-IgE inhibitor (e.g., montelukast sodium), a sympathomimetic (e.g., albuterol), an antibiotic 55 (e.g., tobramycin), a deoxyribonuclease (e.g., Pulmozyme®), an anticholinergic drug (e.g., ipratropium bromide), a corticosteroid (e.g., dexamethasone), a β-adrenoreceptor agonist, a leukotriene inhibitor (e.g., zileuton), a 5-lipoxygenase inhibitor, a phosphodiesterase (PDE) inhibitor, a CD23 60 antagonist, an IL-13 antagonist, a cytokine release inhibitor, a histamine H1 receptor antagonist, an anti-histamine, an antiinflammatory agent (e.g., cromolyn sodium or any other antiinflammatory agent known in the art or described herein), or a histamine release inhibitor.

The following examples are intended to illustrate, not limit, the invention.

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EXAMPLES

Example 1

Process for Formulating and Concentrating Solutions of Eculizumab

Several high concentration formulations of the anti-C5 antibody, eculizumab, were prepared as follows.

Materials and Methods

Instrumentation

The formulation process utilized a Millipore Pellicon® XL tangential flow filter (TFF), Biomax™ 50K Polyethersulfone membrane having a 50 cm² surface area. Also used was a Millipore Sterivex 0.22 micron filter unit (catalogue number SVGV010RS).

Reagents

The formulation process also utilized a number of buffers as follows: (a) Formulation Buffer: 20 mM histidine, 50 mM serine, 2.5% sorbitol, 1.5% mannitol, pH 7.4; (b) Phosphate Buffer: 20 mM sodium phosphate, 80 mM NaCl, pH 6.4; (c) Regeneration Buffer: 0.5 M sodium hydroxide; and (d) Storage Buffer: 0.1 M sodium hydroxide. Also used was a buffer containing eculizumab in the above-described Phosphate Buffer.

Formulation

The tangential flow filter (TFF) was prepared by washing from it the storage buffer using 500 mL of deionized water at a feed flow rate (FR) of 50 mL/minute. The permeate outlet was left open during this process. The TFF was equilibrated using 100 mL of the Phosphate Buffer at a feed flow rate of 50 mL/minute with the permeate outlet open. All of these steps were performed at room temperature.

For all subsequent steps, the pressure was maintained at 40 psi by adjusting the permeate outlet flow rate with a clamp. Eculizumab, initially present at approximately 10 mg/mL in Phosphate Buffer (as described above), was concentrated to 50 mg/mL (for the first run and 40 mg/ml for the second run) at a feed flow rate of 50 mL/minute. The concentrated solution was then diafiltered with six equivalent volumes, for the first run, or four equivalent volumes, for the second run, of the Formulation Buffer. Concentration was continued by gradually reducing the feed flow rate to maintain column pressure at 40 psi, until the feed flow rate reached 2 mL/minute.

Recovery Method

To recover the concentrated eculizumab solution from the column, the permeate outlet was closed and the antibody allowed to circulate for five minutes. The TFF was then flushed out with air, while the permeate outlet was closed. The volume of the recovered solution was measured and recorded. A sample of the high concentration antibody solution was filtered through a 0.22 micron Sterivex filter, diluted 1:100 in formulation buffer. The concentration of the antibody in the diluted sample was determined by measuring $\rm A_{280}$ and using an extinction coefficient of 1.46.

Results

The first run process, described above, required 5.3 hours to complete. A detailed description of the physical and chemical parameters of the TFF flow-through (permeate) and retained (retentate) fractions by time is shown in Table 1.

TABLE 1

Time (min)	Time (hrs)	Pressure (psi)	Feed FR (ml/min)	Permeate FR (ml/min)	Retentate Volume (ml)	Permeate Volume (ml)	Permeate Conc. (mg/ml)	Est. Retentate Conc. (mg/ml)	Details
0	0.0	40	50	0.0	550	0	0.05	10.3	x ^a
24	0.4	40	50	7.5	370	180	0.05	15	
52	0.9	40	50	6.1	200	350	0.05	29	
68	1.1	40	50	4.7	120	425	0.05	46	Diafiltr.
104	1.7	40	50	3.9	120	140	0.05	47	Diafiltr.
118	2.0	40	50	3.6	120	190	0.05	47	Diafiltr.
126	2.1	40	50	3.8	120	220	0.05	47	Diafiltr.
146	2.4	40	50	4.0	120	300	0.05	47	Diafiltr.
226	3.8	40	50	3.8	120	600	0.05	47	Diafiltr.
241	4.0	40	47	2.5	70	38	0.05	81	
247	4.1	40	41	1.7	60	48	0.05	95	
254	4.2	40	35	1.0	53	55	0.05	107	
261	4.4	40	27	0.7	48	60	0.05	116	\mathbf{x}^{b}
275	4.6	40	15	0.7	38	70	0.05	150	
294	4.9	40	6	0.3	33	75	0.05	172	
318	5.3	35	2	0.2	28	80	0.05	186	\mathbf{x}^c

x^a: 5.7 g at 100% purity; actual measured retentate concentration.

The "conc." refers to the concentration of eculizumab in each respective fraction (permeate or retentate).

The initial concentration of eculizumab in the Phosphate Buffer was approximately 10 mg/mL (10.3 mg/mL). Following buffer exchange and diafiltration, the concentration of the solution was initially increased to 116 mg/mL with 98% recovery (100% purity) of the antibody starting material.

Further concentration to 186 mg/mL resulted in a 75% recovery (at 100% purity) of the antibody starting material.

The second run process, described above, required 8.8 hours to complete. A detailed description of the physical and chemical parameters of the TFF flow-through (permeate) and retained (retentate) fractions by time is shown in Table 2.

TABLE 2

Time (min)	Time (hrs)	Pressure (psi)	Feed FR (ml/min)	Permeate FR (ml/min)	Retentate Volume (ml)	Permeate Volume (ml)	Permeate Conc. (mg/ml)	Est. Retentate Conc. (mg/ml)	Details
0	0.0	40	50	0.0	620	0	0.06	8	\mathbf{x}^{a}
10	0.2	40	50	7.5	545	75	0.06	10	
33	0.6	40	50	6.5	395	225	0.06	14	
50	0.8	40	50	5.6	300	320	0.06	18	
70	1.2	40	50	4.8	205	415	0.06	26	
85	1.4	40	50	3.7	150	470	0.06	35	
137	2.3	40	50	3.8	150	200	0.06	37	Diafiltr.
237	4.0	40	50	3.0	150	500	0.06	37	Diafiltr.
303	5.1	40	50	3.0	150	700	0.06	37	Diafiltr.
367	6.1	40	50	3.1	150	900	0.06	37	Diafiltr.
377	6.3	40	50	2.6	119	26	0.06	45	
382	6.4	40	50	2.8	105	40	0.06	51	
388	6.5	40	50	2.0	93	52	0.06	58	
397	6.6	40	50	1.8	77	68	0.06	70	
410	6.8	40	50	1.1	63	82	0.06	86	
422	7.0	40	40	0.8	53	92	0.06	101	
431	7.2	40	30	0.7	47	98	0.06	114	
444	7.4	40	21	0.5	41	104	0.06	131	
451	7.5	40	16	0.3	39	106	0.06	138	
456	7.6	40	15	0.4	37	108	0.06	145	
468	7.8	40	9	0.2	35	110	0.06	151	\mathbf{x}^{b}
489	8.2	40	5	0.1	33	112	0.06	163	
527	8.8	40	2	0.2	25	120	0.06	208	\mathbf{x}^c

 x^a : 5.37 g at 100% purity; actual measured retentate concentration

The "cone." refers to the concentration of eculizumab in each respective fraction (permeate or retentate).

x^b: Recovered 98% at 100% purity; actual measured retentate concentration.

 x^c : Recovered 75% at 100% purity; actual measured retentate concentration.

[&]quot;Diafiltr." refers to diafiltration.

[&]quot;FR" refers to flow rate.

x^b: Recovered 98% at 100% purity; actual measured retentate concentration.

 $[\]mathbf{x}^c$: Recovered 85% at 100% purity; actual measured retentate concentration.

[&]quot;Diafiltr." refers to diafiltration.

[&]quot;FR" refers to flow rate.

The initial concentration of eculizumab in the Phosphate Buffer was approximately 10 mg/mL (8.4 mg/mL). Following buffer exchange and diafiltration, the concentration of the solution was initially increased to 151 mg/mL with 98% recovery (100% purity as determined by SEC-HPLC) of the antibody starting material. Further concentration to 208 mg/mL resulted in an 85% recovery (at 100% purity) of the antibody starting material.

From the results of the first and second run, diafiltering at 40 mg/mL not only improved the final recovery by 10%, but 10 also allowed for the production of high concentration solution having an even higher final concentration of eculizumab (208 mg/mL).

Example 2

Production of Additional Exemplary Eculizumab Formulations

Two additional formulations were developed and evaluated 20 as to their ability to support high concentration solutions of eculizumab. One of the formulations was a histidine/serine/sorbitol/mannitol (HSSM) formulation and the other, a histidine/trehalose/Tween®-20 (HTT) formulation (see below). Two concentrations of eculizumab, approximately 30 mg/mL 25 and approximately 100 mg/mL, were evaluated in each formulation buffer. A third, phosphate-based buffer was also evaluated. A detailed description of the five different antibody solutions (solutions I to V) evaluated is set forth below.

I. 105 mg/mL eculizumab;

20 mM histidine HCl;

50 mM serine;

3% sorbitol; and

1.5% mannitol; at pH 7.0.

II. 30 mg/mL eculizumab;

20 mM histidine HCl;

50 mM serine;

3% sorbitol; and

1.5% mannitol; at pH 7.0.

III. 105 mg/mL eculizumab;

10 mM histidine HCl;

10% alpha-trehalose dihydrate; and

0.01% polysorbate 20; at pH 7.0.

IV. 30.2 mg/mL eculizumab;

10 mM histidine HCl;

10% alpha-trehalose dehydrate; and

0.01% polysorbate 20; at pH 7.0.

V. 10 mg/mL eculizumab;

10 mM sodium phosphate;

150 mM sodium chloride; and

0.02% polysorbate 80; at pH 7.0.

Solutions I-V were prepared by way of concentration and formulation as described above in Example 1, or with only routine and minor modifications to the procedures. Briefly, to prepare solutions I and II, a 10 mg/mL solution of eculizumab 55 in a phosphate-based buffer was concentrated to 30 mg/mL using a TFF. Next, the 30 mg/mL concentrate was subjected to six rounds of diafiltration (as described above in Example 1) into the HSSM formulation (20 mM histidine, 50 mM serine, 3% sorbitol, and 1.5% mannitol, at pH 7.0) to thereby 60 produce solution II. A portion of solution II was further concentrated as described in Example 1 to 100 mg/mL, to produce solution I (the retentate).

Solution IV was prepared by diafiltration of the 30 mg/mL, phosphate-based eculizumab solution in the HTT buffer (10 $\,$ 65 mM histidine HCl; 10% alpha-trehalose dihydrate and 0.01% polysorbate 20, at pH 7.0), followed by the addition of Tween

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20 to 0.01%. To prepare solution III, the 30 mg/mL, phosphate-based eculizumab solution was diafiltered in HTT buffer and then further concentrated using the TFF as described in Example 1. Tween 20 was added to the retentate to a concentration of 0.01%. A flow chart depicting the steps for formulation of the five solutions is shown in FIG. 1. Each of the solutions was passed through a 0.22 μM filter.

Example 3

Stability of an Anti-C5 Antibody Formulated at High Concentration

A series of experiments was performed to evaluate the structural and functional stability of eculizumab formulated at high concentrations in aqueous solution (as prepared in Example 2). Sample aliquots of 2 mL were stored at -20° C., 2-8° C., and 37° C., and then evaluated at specified time intervals (e.g., one month, two months, three months, six months, nine months, 12 months, 18 months, and 24 months). The solutions were subjected to a number of chemical evaluations: appearance (visual inspection), osmolality, concentration (using a UV spectrophotometer), purity (by size exclusion chromatography-HPLC), isoelectric focusing (IEF), and sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE). The functional stability of the antibody in each solution was tested using a human C5-binding assay. The results of each of the evaluations are provided below in the 30 following tables (Roman numerals I to V in the tables correspond to the five named solutions above).

Methods and Results

A. Appearance (Visual Color, Clarity, Particles)

The appearance of each solution (under the varied storage conditions) was evaluated visually by observation of the vial against both a white and a black background. The test was performed at the intervals recited in the following tables for samples maintained at 2-8° C., -20° C., and 37° C. All solutions stored at 2-8° C. were found to be clear, colorless, and particulate free even after 24 months of storage.

All solutions stored at -20° C. were found to be clear, colorless, and particulate free at one month of storage. Solution V was not tested beyond 1 month, but solutions I to IV were clear, colorless, and particulate free for up to 6 months of storage. At 12 months, solution II contained small white particles. At 24 months solution I also contained small white particles. Solutions III and IV remained clear, colorless, and particulate free for at least 24 months.

All solutions stored at 37° C. were found to be clear, colorless, and particulate free for at least 1 month. After two months of storage at 37° C., solutions I and III appeared pale yellow in color. After three months of storage at 37° C., solutions I, II, III, and IV all appeared pale yellow in color and remained unchanged through six months.

B. Osmolality

The osmolality of each solution was measured using freezing point depression.

The test was performed at the time intervals recited in the tables below for samples stored at $2-8^{\circ}$ C., -20° C., and 37° C. Samples were tested in triplicate and the value reported herein is the mean of the three results.

All solutions stored at 2-8° C. had initial (T_0) osmolalities ranging from 299 to 365 mOsm/kg. After 24 months of storage at 2-8° C., the osmolality for each solution showed slight fluctuations, which were within the error of the method. The measured osmolalities for solutions I, II, and IV stored at 2-8° C. remained within +15% of the initial measured osmolality,

which is typically the osmolality specification for solutions in the earliest stage of development.

All solutions stored at -20° C. had initially-measured osmolalities ranging from 299 to 365 mOsm/kg. The osmolality for each solution stored up to 24 months at -20° C. showed slight fluctuations, which are within the error of the method. The osmolalities for all solutions remained within +15% of the initially-measured osmolalities.

All solutions stored at 37° C. had initial (T_0) osmolalities ranging from 299 to 365 mOsm/kg. The osmolality of solutions I, II, and IV showed slight fluctuations during the first six months measurement, all of which were within the error of the method. The osmolalities for all solutions remained within +15% of the initial osmolality. Solution III, however, had a measured osmolality of 863 mOsm/kg at 6 months, well above±15% of the initial osmolality. While the disclosure is not bound by any particular theory or mechanism of action, it is believed that the solution by six months storage at 37° C. had undergone significant degradation, which resulted in an aberrant measurement at this time point.

C. Protein Concentration

Absorbance at 280 nm was used to determine the protein concentration in each test sample using an extinction coefficient of 1.46. The test sample was diluted to give an absorbance reading in the linear range of the assay (0.2 to 1.0 absorbance units). The absorbance of triplicate samples was measured by one operator, and then repeated independently by a second operator. The value reported is determined from the absorbance mean of the six measurements and the applied extinction coefficient.

D. Purity by HPLC Gel Permeation

The relative percents of monomeric IgG, aggregate, and fragments of the anti-C5 antibody were determined using SEC-HPLC (also referred to as gel permeation (GP)-HPLC). Test samples were injected onto a TSKgel G3000 SWXL column (Sigma-Aldrich) equilibrated with phosphate buffered saline (PBS), pH 7.0. The isocratic elution of the proteins is accomplished with a 20 minute run using the PBS, pH 7.0, at a flow rate of 1.0 mL/minute. Protein peaks were monitored by spectrophotometry at a wavelength of 214 nm and the percent purity of the monomeric IgG is expressed as a percentage of the total integrated peak area. Detection of the larger mass multimers was by observation of peaks eluting prior to the monomer peak. A measurement was made at each of the intervals recited in the following tables for samples stored at 2-8° C., -20° C., and 37° C.

All solutions stored at 2-8° C. had an initially-measured (T_0) purity of 99.1% monomer or greater. The purity for most 50 of the solutions at up to six months of storage at 2-8° C. showed slight fluctuations equal to the variability of the method throughout the study. The purity for all solutions remained equal to or greater than 98.0% for up to 24 months of storage. The stability profile of solution II (in the so-called 55 "HSSM" formulation) most closely resembled the profile of the control (Solution IV—10 mg/mL in 10 mM sodium phosphate, 150 mM sodium chloride, 0.02% polysorbate 80, pH 7.0).

All solutions stored at -20° C. had a purity of 99.1% 60 monomer or greater at T=0. From T=0 through T=24 months, the purity for most of the solutions showed slight fluctuations equal to the variability of the method throughout the study. The purity of Solutions 1 and 2 remained virtually unchanged through T=24 months. The purity of Solution 3 remained 65 above 98.7% through 12 months, and then dropped to 97.3% at 24 months. Solution 4 had a purity of ≥98.5% through 24

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months. Solutions 1, 2, and 4 all remained at above 98.0% through 24 months. Solution 3 remained above 95.0% through 24 months.

All solutions stored at 37° C. had a purity of 99.1% monomer or greater at T=0. Beginning at the 1 month time point, significant increases in aggregate and fragments were seen in all solutions. The purity of solutions I, II, and III remained above 90.5% through six months. Solution IV had a purity of \geq 97.4% through six months.

Solutions III and IV contained detectable amounts of fragment at T_0 (0.3 and 0.7%, respectively) which then increased in all storage conditions during the study.

E. SDS-PAGE (Non-Reduced Coomassie)

Samples of the stored solutions were denatured by heating
15 in the presence of sodium dodecyl sulfate (SDS). The
polypeptides present in the samples were separated according
to molecular size by electrophoresis through a gradient 4% to
20% w/v pre-cast Tris-glycine SDS-polyacrylamide gel. The
proteins within the gel were visualized by staining the gel
with Coomassie Blue.

Polypeptide bands within the gels were quantified using laser densitometry. The limit of quantitation of the staining procedure was approximately $0.08\,\mu\text{g/polypeptide}$ band. That is, when an $8\,\mu\text{g}$ test sample was applied, the limit of quantitation of a single discrete impurity is equivalent to approximately 1.0% of the total protein. The reproducibility of the method expressed as a coefficient of variation is approximately 1.8%. The test was performed at the intervals in the following tables (below) for samples stored at $2-8^{\circ}$ C., -20° C., and 37° C.

All solutions stored at 2-8° C. had an initially-measured (T_0) relative percentage of at least 90% IgG. The percent IgG for all solutions stored at 2-8° C. was \geq 90% at 24 months. All solutions stored at -20° C. also had an initially-measured relative percentage of 90% IgG. The percent IgG for solutions I and IV was \geq 90% at 24 months. Solutions II and III had percentage of IgG of 89% and 88%, respectively, at 24 months. As the percentage of IgG at T_0 was 90%, 88% and 89% were within the error of the method and did not necessarily represent a significant change in quality.

All solutions stored at 37° C. had a relative percent IgG of 90% at T_0 . The percentage of IgG for solution IV remained \geq 90% through 3 months of storage. Solution I contained 88% IgG at 3 months, which was within the error of the method, and did not necessarily represent a significant change in quality. At 6 months of storage at 37° C., all solutions were \leq 73% IgG, indicating significant degradation of the antibody.

F. SDS-PAGE (Reduced Coomassie)

Samples were prepared and analyzed as described under Section E "SDS-PAGE (Non-reduced Coomassie)"; however, the samples were further denatured in the presence of 50 mM dithiothreitol (DTT) to disrupt disulfide bonds within the antibody structure. The test was performed at the intervals in the following tables (below) for samples stored at 2-8° C., -20° C., and 37° C.

All solutions stored at 2-8° C. had a relative percent of IgG as heavy and light chain of 100% at $T_{\rm o}$. The percent of IgG as heavy and light chain for all of the solutions stored at 2-8° C. remained at 100% throughout the study. All solutions stored at -20° C. also had a relative percent of IgG as heavy and light chain of 100% throughout the study.

All solutions stored at 37° C. had a relative percent of IgG as heavy and light chain of 100% at T_{0} . At 1 month, the relative percentage of IgG as heavy and light chain remained at 100% for solutions III and IV. The percentage of IgG as heavy and light chains for solution I fell to 99% and for Solution II fell to 98% after one month of storage at 37° C.

After two months of storage at 37° C., all solutions were ≤98.0% IgG as heavy and light chains. The percentage of IgG as heavy and light chains continued to fall through six months of storage at 37° C. for each solution, indicating significant degradation of the antibody.

G. Isoelectric Focusing

The isoelectric focusing studies used a flat bed electrophoresis system. Pre-cast agarose gels covering a pH range of 3.0 to 10.0 were employed. Samples of the stored samples were loaded onto the gel at a predetermined optimized load position along with commercially available pI marker standards. Following focusing for a set number of volt-hours, separated charge variants were visualized by staining with a Coomassie Blue staining solution. The banding pattern of samples on the stained gel was analyzed using laser densitometry. The pI of the separated isoforms and the relative mass of each isoform (as a percentage of total mass) was also determined. pIs are calculated by interpolation from a standard curve established by the pI marker standards. The relative mass of each isoform as a percent of total mass is calculated from its response relative to the total response of the sample load. The measurements were performed at the intervals in the following tables (below) for samples stored at 2-8° C., -20° C., and 37° C.

All solutions stored at 2-8° C. had banding patterns comparable to a reference antibody standard and pI ranges of approximately 5.66 to 6.35 at $\rm T_{\rm O}$. Minor variations in pI were seen throughout the study, which are within the variability of the method. After up to 12 months of storage, banding patterns of the antibody present in the samples were comparable to the reference antibody standard and the pI of all bands remained between 5.48 and 6.60 for all of the solutions. Solutions I and II had banding patterns which remained comparable to the reference antibody material and the pI of all bands between 5.48 and 6.60 through 24 months of storage at 2-8° C. Solutions III and IV did exhibit a change in the banding pattern beginning at the 18 month time point. That is, the most basic band was not detected.

All solutions stored at -20° C. had banding patterns comparable to the reference antibody material and pI ranges of -5.66 to 6.35 at T_0 . Minor variations in pI were seen throughout the study, which are within the variability of the method. Throughout 24 months of storage at -20° C., banding patterns were comparable to the reference antibody material and the pI of all bands was between 5.61 and 6.44 for all of the solutions.

All solutions stored at $37^{\rm o}$ C. had banding patterns comparable to the reference antibody and pI ranges of -5.66 to 6.35

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at T=0. After up to 1 month of storage, banding patterns were comparable to reference material and the pI of all bands was between 5.45 and 6.43 for all of the solutions. After two months of storage, smearing between the bands appeared, an additional acidic band appeared, and the intensities of the bands diminished. No significant difference was seen between the four solutions tested at this temperature condition. The appearance of smearing, development of more acidic bands, and changes in intensity of the main band continued for the duration of the 24 month study.

H. Potency by C5 Binding Assay

The C5 binding assay used to test the functional characteristics of the stored solutions was a quantitative immunoassay. A standard curve was prepared from a reference anti-C5 antibody standard to include concentrations at 500, 250, 125, 62.5 and 31.3 binding units (BU)/mL. A four-parameter fit was applied to the standard curve and test sample results were interpolated from the curve. Each sample was diluted and tested in triplicate at each of three dilutions predetermined to fall within the linear range of the assay. Results were averaged and observed test results in units of BU/mL were divided by the product concentration in mg/mL to obtain results in BU/mg. The measurements were performed at the intervals in the following tables (below) for samples stored at 2-8° C., -20° C., and 37° C.

All solutions stored at 2-8° C. exhibited binding activity ranging from 946,875 to 1,063,353 BU/mg at T_0 . Throughout the 24 month study, the purity for all of the tested solutions remained between 855,801 and 1,194,123 BU/mg. At 12 months, solution I contained 1,306,497 BU/mg, but at 18 and 24 months solution I contained 1,013,876 and 920,747 BU/mg, respectively. All solutions stored at -20° C. contained 946,875 to 1,063,353 BU/mg at T_0 . Throughout the 24 month study, the activity present in all solutions stored at -20° C. remained between 778,672 and 1,148,100 BU/mg.

All solutions stored at 37° C. exhibited initially-measured binding activity ranging from 946,875 to 1,063,353 BU/mg. After six months of storage at 37° C., solutions I and IV exhibiting activity of between 827,206 and 1,202,435 BU/mg. Solutions II and III exhibited binding activity between 1,019,401 and 1,243,601 BU/mg after 3 months, whereas, at six months, the activity of the antibody maintained at 37° C. in solution II and III was 1,679,080 and 1,976,400 BU/mg respectively. Again, while the disclosure is not bound by any particular theory or mechanism of action, it is believed that the solution by six months storage at 37° C. had undergone significant degradation, which resulted in an aberrant measurement at this time point.

TABLE 3

Analytical	Time Point	2-8° C.						
method	(months)	I	II	III	IV	V		
Appearance	0	Clear and colorless, Particulate free	Clear and colorless, Particulate free					
	1	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free		
	2	Clear and colorless, Particulate free						

TABLE 3-continued

Analytical method	Time Point	2-8° C.						
	(months)	I	II	III	IV	V		
	3	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free		
	6	Clear and colorless, Particulate	Clear and colorless, Particulate	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free		
	9	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free		
	12	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free		
	18	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free		
	24	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free		
Osmolality	0 1 2 3	365 mOsm/kg	333 mOsm/kg	356 mOsm/kg	313 mOsm/kg	299 mOsm/kg		
	6 9	367 mOsm/kg	336 mOsm/kg	364 mOsm/kg	326 mOsm/kg	300 mOsm/kg		
	12 18	371 mOsm/kg 366 mOsm/kg	335 mOsm/kg 337 mOsm/kg	366 mOsm/kg 358 mOsm/kg	310 mOsm/kg 324 mOsm/kg	305 mOsm/kg 303 mOsm/kg		
	24	372 mOsm/kg	338 mOsm/kg	382 mOsm/kg	321 mOsm/kg	306 mOsm/kg		

TABLE 4

Analytical	Time Point	2-8° C.					
method	(months)	I	II	III	IV	V	
Protein	0	105.9 mg/mL	29.3 mg/mL	107.6 mg/mL	31.1 mg/mL	10.0 mg/mL	
Concentration	1	108.4 mg/mL	29.9 mg/mL	108.6 mg/mL	31.2 mg/mL	10.2 mg/mL	
	2	104.0 mg/mL	29.1 mg/mL	101.6 mg/mL	30.0 mg/mL	10.0 mg/mL	
	3	102.8 mg/mL	29.0 mg/mL	104.3 mg/mL	30.2 mg/mL	9.8 mg/mL	
	6	104.0 mg/mL	29.1 mg/mL	103.1 mg/mL	30.4 mg/mL	9.9 mg/mL	
	9	110.0 mg/mL	30.8 mg/mL	112.1 mg/mL	31.6 mg/mL	10.0 mg/mL	
	12	106.2 mg/mL	30.2 mg/mL	109.8 mg/mL	31.3 mg/mL	10.1 mg/mL	
	18	108.7 mg/mL	30.6 mg/mL	110.9 mg/mL	31.2 mg/mL	10.1 mg/mL	
	24	106.4 mg/mL	30.7 mg/mL	111.8 mg/mL	32.5 mg/mL	10.3 mg/mL	
SEC-HPLC	0	0.3%	0.2%	0.3%	0.2%	0.2%	
		aggregates 99.7%	aggregates 99.8%	aggregates 99.5%	aggregates 99.1%	aggregates 99.8%	
		monomer 0%	monomer 0%	monomer 0.3%	monomer 0.7%	monomer 0%	
	1	fragments 0.4%	fragments 0.3%	fragments 0.4%	fragments 0.4%	fragments 0.3%	
		aggregates 99.6%	aggregates 99.7%	aggregates 98.7%	aggregates 98.7%	aggregates 99.7%	
		monomer 0%	monomer 0%	monomer 0.9%	monomer 0.9%	monomer 0%	
	2	fragments 0.5%	fragments 0.4%	fragments 0.4%	fragments 0.3%	fragments 0.3%	
		aggregates 99.5%	aggregates 99.6%	aggregates 98.6%	aggregates 98.6%	aggregates 99.7%	

TABLE 4-continued

A malestical	Time Point			2-8° C.		
Analytical method			II		IV	V
memod	(months)	1	11	111	1 V	V
		monomer	monomer	monomer	monomer	monomer
		0%	0%	1.0%	1.2%	0%
	3	fragments 0.5%	fragments 0.4%	fragments 0.3%	fragments 0.2%	fragments 0.3%
	,	aggregates 99.5%	aggregates 99.6%	aggregates 98.7%	aggregates 98.5%	aggregates
		monomer	monomer	monomer	monomer	monomer
		0%	0%	1.0%	1.3%	0%
	6	fragments 0.6%	fragments 0.4%	fragments 0.4%	fragments 0.3%	fragments 0.3%
		aggregates 99.4%	aggregates 99.6%	aggregates 98.8%	aggregates 98.8%	aggregates 99.6%
		monomer 0%	monomer 0%	monomer 0.7%	monomer 1.0%	monomer 0.1%
	9	fragments 0.9%	fragments 0.3%	fragments 0.5%	fragments 0.3%	fragments 0.4%
		aggregates 99.2%	aggregates 99.7%	aggregates 99.3%	aggregates 99.4%	aggregates 99.7%
		monomer	monomer	monomer	monomer	monomer
		0%	0%	0.2%	0.3%	0%
	12	fragments 1.3%	fragments 0.7%	fragments 0.5%	fragments 0.4%	fragments 0.6%
		aggregates 98.7%	aggregates 99.3%	aggregates 98.9%	aggregates 98.8%	aggregates 99.5%
		monomer	monomer	monomer	monomer	monomer
		0%	0%	0.6%	0.9%	0%
	18	fragments 1.6%	fragments 0.8%	fragments 0.6%	fragments 0.3%	fragments 0.5%
		aggregates 98.4%	aggregates 99.2%	aggregates 99.2%	aggregates 99.3%	aggregates 99.5%
		monomer	monomer	monomer	monomer	monomer
		0%	0%	0.2%	0.3%	0%
	24	fragments 2.0%	fragments 0.9%	fragments 0.7%	fragments 0.3%	fragments 0.6%
		aggregates 98.0%	aggregates 99.1%	aggregates 99.1%	aggregates 99.4%	aggregates 99.4%
		monomer	monomer	monomer	monomer	monomer
		0% fragments	0% fragments	0.2% fragments	0.3% fragments	0% fragments
		Tragillenis	nagments	nagments	Tragilients	nagments

TABLE 5

Analytical	Time Point	llysis of Eculizumab Solutions Stored at 2 to 8° C. for 24 Months 2-8° C.					
method	(months)	I	II	III	IV	v	
SDS-PAGE	0	90% IgG	90% IgG	90% IgG	90% IgG	90% IgG	
Non-reduced	1	92% IgG	92% IgG	92% IgG	92% IgG	92% IgG	
	2	91% IgG	91% IgG	92% IgG	90% IgG	93% IgG	
	3	92% IgG	92% IgG	92% IgG	92% IgG	91% IgG	
	6	89% IgG	89% IgG	90% IgG	90% IgG	90% IgG	
	9	91% IgG	91% IgG	91% IgG	91% IgG	90% IgG	
	12	90% IgG	90% IgG	90% IgG	90% IgG	91% IgG	
	18	90% IgG	90% IgG	91% IgG	90% IgG	91% IgG	
	24	91% IgG	90% IgG	91% IgG	90% IgG	91% IgG	
SDS-PAGE	0	100% IgG as	100% IgG as	100% IgG as	100% IgG as	100% IgG as	
Reduced		heavy and	heavy and	heavy and	heavy and	heavy and	
		light chains	light chains	light chains	light chains	light chains	
	1	100% IgG as	100% IgG as	100% IgG as	100% IgG as	100% IgG as	
		heavy and	heavy and	heavy and	heavy and	heavy and	
		light chains	light chains	light chains	light chains	light chains	
	2	100% IgG as	100% IgG as	99% IgG as	99% IgG as	100% IgG as	
		heavy and	heavy and	heavy and	heavy and	heavy and	
		light chains	light chains	light chains	light chains	light chains	

TABLE 5-continued

SD	S-PAGE Ans	alysis of Eculiz	umab Solution	s Stored at 2 to	8° C. for 24 M	onths
Analytical	Time Point			2-8° C.		
method	(months)	I	II	III	IV	v
	3	100% IgG as heavy and				
	6	light chains 100% IgG as heavy and				
	9	light chains 100% IgG as heavy and	light chains 100% IgG as heavy and	light chains 100% IgG as heavy and	heavy and light chains 100% IgG as heavy and	light chains 100% IgG as heavy and
	12	light chains 100% IgG as heavy and	light chains 100% IgG as heavy and	light chains 100% IgG as heavy and		light chains 100% IgG as heavy and
	18	light chains 100% IgG as heavy and				
	24	light chains 100% IgG as heavy and light chains				

TABLE 6

	IEF and C5-b	inding Analysis	of Eculizumab So	iutions Stored at 2	2 to 8° C. for 24 N	/IOIIThs
Analytical	Time Point			2-8° C.		
method	(months)	I	II	III	IV	V
IEF	0	3 major bands All major and minor bands resolved				
	1	between pI 5.68 and 6.35 3 major bands All major and minor bands	between pI 5.67 and 6.35 3 major bands All major and minor bands	5.67 and 6.35 3 major bands All major and minor bands	between pI 5.66 and 6.35 3 major bands All major and minor bands	5.72 and 6.35 3 major bands All major and minor bands
	2	resolved between pI 5.68 and 6.44 3 major bands	resolved between pI 5.69 and 6.57 3 major bands	resolved between pI 5.68 and 6.54 3 major bands	resolved between pI 5.69 and 6.54 3 major bands	resolved between pI 5.67 and 6.40 3 major bands
		All major and minor bands resolved between pI 5.50 and 6.42	All major and minor bands resolved between pI 5.48 and 6.40	All major and minor bands resolved between pI 5.51 and 6.43	All major and minor bands resolved between pI 5.53 and 6.46	All major and minor bands resolved between pI 5.54 and 6.48
	3	3 major bands All major and minor bands resolved between pI				
	6	5.69 and 6.42 3 major bands All major and minor bands resolved between pI	5.68 and 6.41 3 major bands All major and minor bands resolved between pI	5.68 and 6.41 3 major bands All major and minor bands resolved between pI	5.69 and 6.41 3 major bands All major and minor bands resolved between pI	5.69 and 6.41 3 major bands All major and minor bands resolved between pI
	9	5.79 and 6.40 3 major bands All major and minor bands resolved between pI	5.79 and 6.39 3 major bands All major and minor bands resolved between pI	5.78 and 6.40 3 major bands All major and minor bands resolved between pI	5.81 and 6.42 3 major bands All major and minor bands resolved between pI	5.82 and 6.44 3 major bands All major and minor bands resolved between pI
	12	5.83 and 6.40 3 major bands All major and minor bands resolved between pI 5.83 and 6.47	5.83 and 6.41 3 major bands All major and minor bands resolved between pI 5.84 and 6.51	5.85 and 6.42 3 major bands All major and minor bands resolved between pI 5.87 and 6.60	5.77 and 6.35 3 major bands All major and minor bands resolved between pI 5.87 and 6.54	5.80 and 6.36 3 major bands All major and minor bands resolved between pI 5.85 and 6.52

TABLE 6-continued

Analytical	Time Point		2-8° C.								
method	(months)	I	II	III	IV	V					
	18	3 major bands All major and minor bands resolved between pI 5.60 and 6.31	3 major bands All major and minor bands resolved between pI 5.58 and 6.30	2 major bands All major and minor bands resolved between pI 5.58 and 6.30 (Does not compare to Reference).	2 major bands All major and minor bands resolved between pI 5.60 and 6.33 (Does not compare to Reference).	3 major bands All major and minor bands resolved between pI 5.61 and 6.34					
	24	3 major bands All major and minor bands resolved between pI 5.64 to 6.29	3 major bands All major and minor bands resolved between pI 5.63 to 6.26	2 major bands All Major and minor bands between pI 5.60 to 6.24 Does not compare to Reference.	2 major bands All Major and minor bands between pI 5.75 to 6.25 Does not compare to Reference.	3 major bands All Major and minor bands between pI 5.62 to 6.26					
C5 Binding	0	1,026,912 BU/mg	1,063,353 BU/mg	1,019,401 BU/mg	967,645 BU/mg	946,875 BU/mg					
	1	1,067,612 BU/mg	1,025,293 BU/mg	981,238 BU/mg	1,078,726 BU/mg	960,989 BU/mg					
	2	1,038,662 BU/mg	1,172,680 BU/mg	1,103,182 BU/mg	1,052,083 BU/mg	1,097,917 BU/mg					
	3	1,031,534 BU/mg	1,127,155 BU/mg	1,074,624 BU/mg	968,543 BU/mg	1,140,519 BU/mg					
	6	879,407 BU/mg	856,959 BU/mg	1,121,484 BU/mg	860,403 BU/mg	894,360 BU/mg					
	9	1,009,470 BU/mg	1,015,625 BU/mg	965,470 BU/mg	973,674 BU/mg	1,026,042 BU/mg					
	12	1,306,497 BU/mg	1,194,123 BU/mg	1,107,127 BU/mg	1,097,244 BU/mg	1,127,269 BU/mg					
	18	1,013,876	855,801	958,070	948,518	898,309					
	24	BU/mg 920,747 BU/mg	BU/mg 958,880 BU/mg	BU/mg 1,036,747 BU/mg	BU/mg 848,043 BU/mg	BU/mg 914,522 BU/mg					

[&]quot;BU" refers to binding units.

TABLE 7

	Appeara	, , , , , , , , , , , , , , , , , , , ,	and Protein Conce d at 37° C. for Up		zumab Solutions						
Analytical	Time Point		37° C.								
method	(months)	I	II	III	IV	V					
Appearance	0	Clear and colorless,	Clear and colorless,	Clear and colorless,	Clear and colorless,	Clear and colorless,					
	1	Clear and colorless,	Clear and colorless,	Clear and colorless,	Clear and colorless,	n/a					
	2	Particulate free Clear, pale yellow, Particulate free	Particulate free Clear and colorless, Particulate free	Particulate free Clear, pale yellow, Particulate free	Particulate free Clear and colorless, Particulate free	n/a					
	3	Clear, pale yellow, Particulate free	Clear, pale yellow, Particulate free	Clear, pale yellow, Particulate free	Clear, pale yellow, Particulate free	n/a					
	6	Clear, pale yellow, Particulate free	Clear, pale yellow, Particulate free	Clear, pale yellow, Particulate free	Clear, pale yellow, Particulate free	n/a					
Osmolality	0 1 2 3	365 mOsm/kg	333 mOsm/kg	356 mOsm/kg	313 mOsm/kg	299 mOsm/kg					
	6	394 mOsm/kg	349 mOsm/kg	863 mOsm/kg	332 mOsm/kg	n/a					

TABLE 7-continued

	Appearance, Osmolality, and Protein Concentration of Eculizumab Solutions Stored at 37° C. for Up to 24 months											
Analytical	Time Point		37° C.									
method	(months)	I	II	III	IV	V						
Protein	0	105.9 mg/mL	29.3 mg/mL	107.6 mg/mL	31.1 mg/mL	10.0 mg/mL						
Concentration	1	99.0 mg/mL	30.1 mg/mL	111.7 mg/mL	31.8 mg/mL	n/a						
	2	115.1 mg/mL	29.8 mg/mL	114.3 mg/mL	31.7 mg/mL	n/a						
	3	106.2 mg/mL	30.1 mg/mL	113.7 mg/mL	31.5 mg/mL	n/a						
	6	109.5 mg/mL	38.6 mg/mL	139.3 mg/mL	35.7 mg/mL	n/a						

TABLE 8

	SEC-F	IPLC and SDS Stored at	-PAGE Analysi : 37° C. for Up		b Solutions	
Analytical	Time Point			37° C.		
method	(months)	I	II	III	IV	V
SEC-HPLC	0	0.3%	0.2%	0.3%	0.2%	0.2%
		aggregates 99.7%	aggregates 99.8%	aggregates 99.5%	aggregates 99.1%	aggregates 99.8%
		monomer 0%	monomer 0%	monomer 0.3%	monomer 0.7%	monomer 0%
	1	fragments 2.3%	fragments 1.4%	fragments	fragments 0.7%	fragments n/a
		aggregates 97.6%	aggregates 98.5%	aggregates 97.9%	aggregates 98.3%	II a
		monomer 0.1%	monomer 0.1%	monomer 0.7%	monomer	
	2	fragments 4.1%	fragments 2.5%	fragments 2.2%	fragments 1.0%	n/a
		aggregates 95.7%	aggregates 97.3%	aggregates 97.1%	aggregates 98.0%	
		monomer 0.2%	monomer 0.2%	monomer 0.7%	monomer 1.0%	
	3	fragments 5.4%	fragments 3.2%	fragments 5.6%	fragments 1.1%	n/a
		aggregates 94.5%	aggregates 96.5%	aggregates 96.1%	aggregates 97.9%	
		monomer 0.2%	monomer 0.3%	monomer 1.4%	monomer 1.0%	
	6	fragments 9.1%	fragments 6.0%	fragments 3.8%	fragments 1.8%	n/a
		aggregates 90.5%	aggregates 93.4%	aggregates 94.9%	aggregates 97.4%	
		monomer 0.4%	monomer 0.5%	monomer 1.3%	monomer 0.8%	
SDS-PAGE	0	fragments 90% IgG	fragments 90% IgG	fragments 90% IgG	fragments 90% IgG	n/a
Non-reduced	1	92% IgG	91% IgG	92% IgG	92% IgG	n/a
	2	91% IgG	92% IgG	91% IgG	92% IgG	n/a
	3	88% IgG	79% IgG	84% IgG	91% IgG	n/a
	6	62% IgG	60% IgG	70% IgG	73% IgG	n/a
SDS-PAGE	0	100% IgG as	100% IgG as	100% IgG as	100% IgG as	100% IgG as
Reduced		heavy and	heavy and	heavy and	heavy and	heavy and
	_	light chains	light chains	light chains	light chains	light chains
	1	99% IgG as heavy and	98% IgG as heavy and	100% IgG as heavy and	100% IgG as heavy and	n/a
	2	light chains	light chains	light chains	light chains	n/a
	2	96% IgG as heavy and	96% IgG as heavy and	98% IgG as heavy and	97% IgG as heavy and	II/a
	3	light chains 95% IgG as heavy and	light chains 94% IgG as heavy and	light chains 97% IgG as heavy and	light chains 97% IgG as heavy and	n/a
	6	light chains 88% IgG as	light chains 85% IgG as	light chains 93% IgG as	light chains 91% IgG as	n/a
		heavy and light chains	heavy and light chains	heavy and light chains	heavy and light chains	

TABLE 9

Analytical	Time Point			37° C.		
method	(months)	I	II	III	IV	V
IEF	0	3 major bands All major and minor bands resolved between pI 5,68 and 6,35	3 major bands All major and minor bands resolved between pI 5.67 and 6.35	3 major bands All major and minor bands resolved between pI 5.67 and 6.35	3 major bands All major and minor bands resolved between pI 5.66 and 6.35	3 major bands All major and minor bands resolved between pI 5.72 and 6.35
	1	3 major bands All major and minor bands resolved between pI 5.58 and 6.42	3 major bands All major and minor bands resolved between pI 5.56 and 6.43	3 major bands All major and minor bands resolved between pI 5.55 and 6.43	3 major bands All major and minor bands resolved between pI 5.56 and 6.41	n/a
	2	3 major bands All major and minor bands resolved between pI 5.21 and 6.33	3 major bands All major and minor bands resolved between pI 5.17 and 6.32	3 major bands All major and minor bands resolved between pI 5.18 and 6.31	3 major bands All major and minor bands resolved between pI 5.19 and 6.31	n/a
	3	3 major bands All major and minor bands resolved between pI 5.49 and 6.39	3 major bands All major and minor bands resolved between pI 5.49 and 6.39	3 major bands All major and minor bands resolved between pI 5.50 and 6.43	3 major bands All major and minor bands resolved between pI 5.49 and 6.42	n/a
	6	6 major bands 2 minor bands All major and minor bands resolved between pI 5.62 and 6.44	6 major bands 1 minor bands All major and minor bands resolved between pI 5.61 and 6.28	5 major bands 2 minor bands All major and minor bands resolved between pI 5.70 and 6.40	5 major bands 3 minor bands All major and minor bands resolved between pI 5.62 and 6.41	n/a
C5 Binding	0	1,026,912 BU/mg	1,063,353 BU/mg	1,019,401 BU/mg	967,645 BU/mg	946,875 BU/mg
	1	1,153,199 BU/mg	1,122,301 BU/mg	1,076,171 BU/mg	950,275 BU/mg	n/a
	2	1,157,508 BU/mg	1,243,601 BU/mg	1,056,248 BU/mg	1,042,981 BU/mg	n/a
	3	1,126,020 BU/mg	1,167,982 BU/mg	1085,642 BU/mg	1,013,889 BU/mg	n/a
	6	1,202,435 BU/mg	1,679,080 BU/mg	1,976,400 BU/mg	827,206 BU/mg	n/a

TABLE 10

Analytical	Time Point	−20° C.								
method	(months)	I	II	III	IV	V				
Appearance	0	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free				
	1	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free	n/a				
	6	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free	Clear and colorless, Particulate free	n/a				
	12	Clear and colorless, Particulate free	Clear and colorless Small, white particles observed	Clear and colorless, Particulate free	Clear and colorless, Particulate free	n/a				

TABLE 10-continued

App	earance, Osn			ation Determina or Up to 24 Mo		izumab				
Analytical	Time Point		−20° C.							
method	(months)	I	II	III	IV	V				
	24	Clear and colorless, Small, white particles	Clear and colorless, Small, white particles observed	Clear and colorless, Particulate free	Clear and colorless, Particulate free	n/a				
Osmolality	0	365	333	356	313	299				
		mOsm/kg	mOsm/kg	mOsm/kg	mOsm/kg	mOsm/kg				
	1				2					
	6	361	336	356	316	n/a				
		mOsm/kg	mOsm/kg	mOsm/kg	mOsm/kg					
	12	366	337	362	316	n/a				
		mOsm/kg	mOsm/kg	mOsm/kg	mOsm/kg					
	24	369	339	368	316	n/a				
		mOsm/kg	mOsm/kg	mOsm/kg	mOsm/kg					
Protein	0	105.9 mg/mL	29.3 mg/mL	107.6 mg/mL	31.1 mg/mL	10.0 mg/mL				
Conc.	1	107.5 mg/mL	29.3 mg/mL	109.1 mg/mL	_	n/a				
	6	104.3 mg/mL	_	104.9 mg/mL	30.3 mg/mL	n/a				
	12	105.7 mg/mL	30.2 mg/mL	105.6 mg/mL	31.2 mg/mL	n/a				
	24	107.3 mg/mL	30.6 mg/mL	110.1 mg/mL	32.2 mg/mL	n/a				

TABLE 11

Analytical	Time			−20° C.		
method	Point	I	II	III	IV	V
SEC-HPLC	0	0.3%	0.2%	0.3%	0.2%	0.2%
		aggregates 99.7%	aggregates 99.8%	aggregates 99.5%	aggregates 99.1%	aggregates 99.8%
		monomer 0%	monomer 0%	monomer 0.3%	monomer 0.7%	monomer 0%
	1	fragments 0.4%	fragments 0.3%	fragments 0.4%	fragments 0.3%	fragments n/a
		aggregates 99.6%	aggregates 99.7%	aggregates 98.7%	aggregates 98.6%	
		monomer 0	monomer 0%	monomer	monomer 1.2%	
	6	fragments 0.3%	fragments	fragments 0.9%	fragments	n/a
	-	aggregates 99.7%	aggregates	aggregates 99.3%	aggregates 98.7%	
		monomer 0	monomer 0%	monomer 0.1%	monomer	
	12	fragments	fragments 0.3%	fragments 0.4%	fragments 0.3%	n/a
	12	aggregates 99.6%	aggregates	aggregates 98.7%	aggregates 98.5%	IV d
		monomer 0	monomer 0%	monomer 1.0%	monomer 1.3%	
	24	fragments	fragments	fragments	fragments	n/a
		aggregates 99.6%	aggregates	aggregates 97.3%	aggregates 98.6%	
		monomer 0 %	monomer 0%	monomer 1.0%	monomer	
CDC DACE	0	fragments	fragments	fragments	fragments	(-
SDS-PAGE Non-reduced	0 1	90% IgG 92% IgG	90% IgG 91% IgG	90% IgG 92% IgG	90% IgG 92% IgG	n/a n/a
readood	6	90% IgG	89% IgG	89% IgG	89% IgG	n/a
	12	90% IgG	90% IgG	90% IgG	90% IgG	n/a
	24	90% IgG	89% IgG	88% IgG	91% IgG	n/a
SDS-PAGE Reduced	0	100% IgG as heavy and	n/a			
reduced		neavy and light chains	neavy and light chains	neavy and light chains	neavy and light chains	

TABLE 11-continued

Analytical	Time			−20° C.		
method	Point	I	II	III	IV	V
	1	100% IgG as	100% IgG as	100% IgG as	100% IgG as	n/a
		heavy and	heavy and	heavy and	heavy and	
	_	light chains	light chains	light chains	light chains	
	6	100% IgG as	100% IgG as	100% IgG as	100% IgG as	n/a
		heavy and	heavy and	heavy and	heavy and	
	1.2	light chains	light chains	light chains	light chains	(
	12	100% IgG as	100% IgG as	100% IgG as	100% IgG as	n/a
		heavy and	heavy and	heavy and	heavy and	
	24	light chains	light chains	light chains	light chains	(-
	24	100% IgG as	100% IgG as	100% IgG as	100% IgG as	n/a
		heavy and	heavy and	heavy and	heavy and	
ŒF	0	light chains 3 major bands	light chains 3 major bands	light chains 3 major bands	light chains 3 major bands	3 major bende
.EF	U					3 major bands
		All major and minor bands	All major and minor bands	All major and minor bands	All major and minor bands	All major and minor bands
		resolved	resolved	resolved	resolved	resolved
		between pI	between pI	between pI	between pI	between pI
		5.68 and 6.35	5.67 and 6.35	5.67 and 6.35	5.66 and 6.35	5.72 and 6.35
	1	3 major bands	3 major bands	3 major bands	3 major bands	n/a
	1	All major and	All major and	All major and	All major and	IV &
		minor bands	minor bands	minor bands	minor bands	
		resolved	resolved	resolved	resolved	
		between pI	between pI	between pI	between pI	
		5.67 and 6.41	5.64 and 6.37	5.61 and 6.36	5.63 and 6.35	
	6	3 major bands	3 major bands	3 major bands	3 major bands	n/a
		All major and	All major and	All major and	All major and	12.0
		minor bands	minor bands	minor bands	minor bands	
		resolved	resolved	resolved	resolved	
		between pI	between pI	between pI	between pI	
		5.76 and 6.44	5.75 and 6.42	5.72 and 6.41	5.73 and 6.42	
	12	3 major bands	3 major bands	3 major bands	3 major bands	n/a
		All major and	All major and	All major and	All major and	
		minor bands	minor bands	minor bands	minor bands	
		resolved	resolved	resolved	resolved	
		between pI	between pI	between pI	between pI	
		5.83 and 6.38	5.82 and 6.37	5.83 and 6.38	5.81 and 6.37	
	24	3 major bands	3 major bands	3 major bands	3 major bands	n/a
		All major and	All major and	All major and	All major and	
		minor bands	minor bands	minor bands	minor bands	
		resolved	resolved	resolved	resolved	
		between pI	between pI	between pI	between pI	
		5.64 to 6.27	5.62 to 6.26	5.63 to 6.27	5.63 to 6.26	
C5 Binding	0	1,026,912	1,063,353	1,019,401	967,645	946,875
		BU/mg	BU/mg	BU/mg	BU/mg	BU/mg
	1	1,032,946	1,071,886	903,223	806,290	n/a
	_	BU/mg	BU/mg	BU/mg	BU/mg	,
	6	1,067,633	904,948	912,575	778,672	n/a
		BU/mg	BU/mg	BU/mg	BU/mg	,
	12	1,148,100	1,081,333	1,144,255	1,068,710	n/a
	2.4	BU/mg	BU/mg	BU/mg	BU/mg	,
	24	894,507	918,210	954,360	790,264	n/a

DISCUSSION

As set forth in Tables 3 to 6, the high concentration antibody formulations described herein were markedly stable over a two year period. Each of solutions I to IV remained clear, colorless, and particulate-free over the course of the study, which indicated that no visible precipitation occurred during a two-year storage period. There was also no significant change in osmolality or protein concentration of these solutions, even at 24 months.

Moreover, the antibody present in solutions I and III (105 mg/mL antibody) remained at least 98% monomeric. As shown in Table 5, the antibody present in solution III 65 remained over 99% monomeric even at the 2 year testing. Each of the solutions maintained the anti-C5 antibody as over

99% monomer when stored at 2° C. to 8° C. for up to 9 months. The highly-concentrated solutions not only maintained a high percentage of monomeric antibody, but contained very few aggregates or degradation or fragmentation products. For example, solutions I and II contained no detectable fragmentation products as determined by SEC-HPLC, even at 24 months of storage at 2° C. to 8° C. Solutions III and IV contained less than 0.5% fragments at 24 months (0.2% antibody fragments in solution III at 24 months and 0.3% antibody fragments in solution IV at 24 months). None of the solutions contains more than 2% aggregates at 24 months, with solutions II, III, and IV containing less than 1% aggregates at 24 months. These results indicate that the formulations described herein are capable of substantially maintaining the structural integrity of the anti-C5 antibody dissolved therein for at least 24 months storage at 2° C. to 8° C.

The solutions were also evaluated for retention of functional activity by measuring C5-binding activity. As set forth in Table 6, each of the solutions tested retained approximately 90% or more of their C5-binding activity after 24 months of storage at 2° C. to 8° C. The antibody formulated in solution III retained 100% of its C5-binding ability at 24 months. Virtually no change in the binding activity of the antibody to C5 was detected in any of the formulations tested at one year. These results indicate that the formulations described herein maintain the functional as well as structural stability of the anti-C5 antibodies dissolved therein for at least 2 years at 2° C. to 8° C.

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While the present disclosure has been described with reference to the specific embodiments thereof, it should be understood by those skilled in the art that various changes may be made and equivalents may be substituted without departing from the true spirit and scope of the disclosure. In addition, many modifications may be made to adapt a particular situation, material, composition of matter, process, process step or steps, to the objective, spirit and scope of the present disclosure. All such modifications are intended to be within the scope of the disclosure.

SEQUENCE LISTING

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Ala Phe Asp Ala Thr Ile Ser Ile Lys Ser Tyr Pro Asp Lys Lys Phe
Ser Tyr Ser Ser Gly His Val His Leu Ser Ser Glu Asn Lys Phe Gln 65 70 75 80
As Ser Ala Ile Leu Thr Ile Gln Pro Lys Gln Leu Pro Gly Gln Gln 85 90 95
Asn Pro Val Ser Tyr Val Tyr Leu Glu Val Val Ser Lys His Phe Ser
Lys Ser Lys Arg Met Pro Ile Thr Tyr Asp Asn Gly Phe Leu Phe Ile 115 $120\ 
His Thr Asp Lys Pro Val Tyr Thr Pro Asp Gln Ser Val Lys Val Arg
Val Tyr Ser Leu Asn Asp Asp Leu Lys Pro Ala Lys Arg Glu Thr Val
Leu Thr Phe Ile Asp Pro Glu Gly Ser Glu Val Asp Met Val Glu Glu
Ile Asp His Ile Gly Ile Ile Ser Phe Pro Asp Phe Lys Ile Pro Ser
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Asn Pro Arg Tyr Gly Met Trp Thr Ile Lys Ala Lys Tyr Lys Glu Asp
                            200
Phe Ser Thr Thr Gly Thr Ala Tyr Phe Glu Val Lys Glu Tyr Val Leu
                        215
Pro His Phe Ser Val Ser Ile Glu Pro Glu Tyr Asn Phe Ile Gly Tyr
Lys Asn Phe Lys Asn Phe Glu Ile Thr Ile Lys Ala Arg Tyr Phe Tyr
Asn Lys Val Val Thr Glu Ala Asp Val Tyr Ile Thr Phe Gly Ile Arg
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Glu Asp Leu Lys Asp Asp Gln Lys Glu Met Met Gln Thr Ala Met Gln
                         280
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Thr 305	Ala	Val	Lys	Glu	Leu 310	Ser	Tyr	Tyr	Ser	Leu 315	Glu	Asp	Leu	Asn	Asn 320
ГÀЗ	Tyr	Leu	Tyr	Ile 325	Ala	Val	Thr	Val	Ile 330	Glu	Ser	Thr	Gly	Gly 335	Phe
Ser	Glu	Glu	Ala 340	Glu	Ile	Pro	Gly	Ile 345	Lys	Tyr	Val	Leu	Ser 350	Pro	Tyr
ГÀа	Leu	Asn 355	Leu	Val	Ala	Thr	Pro 360	Leu	Phe	Leu	ГЛа	Pro 365	Gly	Ile	Pro
Tyr	Pro 370	Ile	Lys	Val	Gln	Val 375	Lys	Asp	Ser	Leu	Asp 380	Gln	Leu	Val	Gly
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Phe	Asn	Val 435	Lys	Thr	Asp	Ala	Pro 440	Asp	Leu	Pro	Glu	Glu 445	Asn	Gln	Ala
Arg	Glu 450	Gly	Tyr	Arg	Ala	Ile 455	Ala	Tyr	Ser	Ser	Leu 460	Ser	Gln	Ser	Tyr
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Thr	His	Tyr	Asn 500	Tyr	Leu	Ile	Leu	Ser 505	Lys	Gly	ГÀз	Ile	Ile 510	His	Phe
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Leu	Asn	Ile	Glu	Glu 565	ràa	CÀa	Gly	Asn	Gln 570	Leu	Gln	Val	His	Leu 575	Ser
Pro	Asp	Ala	Asp 580		Tyr	Ser	Pro		Gln			Ser	Leu 590	Asn	Met
Ala	Thr	Gly 595	Met	Asp	Ser	Trp	Val 600	Ala	Leu	Ala	Ala	Val 605	Asp	Ser	Ala
Val	Tyr 610	Gly	Val	Gln	Arg	Gly 615	Ala	Lys	Lys	Pro	Leu 620	Glu	Arg	Val	Phe
Gln 625	Phe	Leu	Glu	Lys	Ser 630	Asp	Leu	Gly	Cys	Gly 635	Ala	Gly	Gly	Gly	Leu 640
Asn	Asn	Ala	Asn	Val 645	Phe	His	Leu	Ala	Gly 650	Leu	Thr	Phe	Leu	Thr 655	Asn
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Lys	Tyr 690	Lys	His	Ser	Val	Val 695	Lys	Lys	Сув	Сув	Tyr 700	Asp	Gly	Ala	Сув

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Glu	Ala 1010		ı Let	ı Met	Sei	7 Va:		al P:	ro V	al Pi		yr ' 020	Val :	Phe :	His
Tyr	Leu 1025		ı Thi	r Gly	/ Asr	103		rp A	sn I	le P		is : 035	Ser 2	Asp	Pro
Leu	Ile 1040		ı Lys	s Glr	ı Lys	Le: 104		As Pi	ys L	γs L		ys (050	Glu (Gly 1	Met
Leu	Ser 1055		e Met	Se:	ту1	100		en A	la A	sp T		er ' 065	Tyr :	Ser	Val
Trp	Lys 1070	_	/ Gly	/ Sei	Ala	Sei 10		nr T	rp L	eu Ti		la 1	Phe 1	Ala :	Leu
Arg	Val		ı Gly	/ Glr	n Val	109		ys T	yr Va	al G		ln 2	Asn (Gln .	Asn
Ser	Ile 1100	-	a Ası	ı Sei	: Leu	ı Lei 110		rp L	eu V	al G		sn '	Tyr (Gln :	Leu
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		-			**- 7	T] - 0	· · · · · · · · · · · · · · · · · · ·	1 - D-				01
1	GIU	GIII		1 y r 5	val	iie s	er A	10 PI		/S I.	le Phe	e Arg	15	GIY
Ala	Ser		Asn 20	Ile	Val :	Ile G	ln V. 2	_	yr G	Ly Ty	r Thi	Glu 30	. Ala	Phe
Asp		Thr 35	Ile	Ser	Ile		er T	yr Pi	ro As	sp Ly	/s Lys 45	Phe	Ser	Tyr
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Ala 65	Ile	Leu	Thr		Gln i 70	Pro L	ys G	ln Le	eu Pr 75		ly Gly	/ Gln	Asn	Pro 80
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Lys	Arg	Met	Pro 100	Ile	Thr	Tyr A	_	sn G] 05	Ly Pl	ne Le	eu Phe	e Ile 110		Thr
Asp	_	Pro 115	Val '	Tyr	Thr :		sp G 20	ln Se	er Va	al Ly	/s Val	_	Val	Tyr
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	Tle	Asp	Dro 1	Glu	Gly :	Ser G	lu V	al As	ap Me	et Va	al Glu	ı Glu	. Ile	Asp
145		-	110		150				15					160
			Ile		150	Phe P	ro A	sp Pł	15	55	le Pro	Ser	Asn 175	Pro
His	Ile	Gly	Ile	Ile 165	150 Ser :		ys A	sp Ph 17	15 ne Ly 70	55 /s I]	le Pro		175 Phe	Pro

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Val	Val	Thr	Glu	Ala 245	Asp	Val	Tyr	Ile	Thr 250	Phe	Gly	Ile	Arg	Glu 255	Asp
Leu	Lys	Asp	Asp 260	Gln	Lys	Glu	Met	Met 265	Gln	Thr	Ala	Met	Gln 270	Asn	Thr
Met	Leu	Ile 275	Asn	Gly	Ile	Ala	Gln 280	Val	Thr	Phe	Asp	Ser 285	Glu	Thr	Ala
Val	Lys 290	Glu	Leu	Ser	Tyr	Tyr 295	Ser	Leu	Glu	Asp	Leu 300	Asn	Asn	Lys	Tyr
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Glu	Ala	Glu	Ile	Pro 325	Gly	Ile	Lys	Tyr	Val 330	Leu	Ser	Pro	Tyr	Lys 335	Leu
Asn	Leu	Val	Ala 340	Thr	Pro	Leu	Phe	Leu 345	Lys	Pro	Gly	Ile	Pro 350	Tyr	Pro
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Thr Gl	n Asn 515	Met	Val	Pro	Ser	Ser 520	Arg	Leu	Leu	Val	Tyr 525	Tyr	Ile	Val
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Gly Va	ıl Gln 595	Arg	Gly	Ala	Lys	Lys 600	Pro	Leu	Glu	Arg	Val 605	Phe	Gln	Phe
Leu Gl 61	_	Ser	Asp	Leu	Gly 615	Cys	Gly	Ala	Gly	Gly 620	Gly	Leu	Asn	Asn
Ala As 625	n Val	Phe		Leu 630		Gly	Leu		Phe 635		Thr	Asn		Asn 640
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Val Il 1	e Asp.	Hls	GIN 5	GIŸ	Tnr	гуз	Ser	ser 10	гуз	Cys	vaı	Arg	GIN 15	гув
Val Gl	u Gly	Ser 20	Ser											
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                          40
Tyr Gly Ala Thr Asn Leu Ala Asp Gly Val Pro Ser Arg Phe Ser Gly
Ser Gly Ser Gly Thr Asp Phe Thr Leu Thr Ile Ser Ser Leu Gln Pro
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                                       75
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Pro Ser Val Phe Ile Phe Pro Pro Ser Asp Glu Gln Leu Lys Ser Gly
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Lys Val Gln Trp Lys Val Asp Asn Ala Leu Gln Ser Gly Asn Ser Gln
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\ <u>L</u> <u>L</u> .		ојуре			1101	. Бе.	3011	20101	I OI	ALC.	LL IC.	Lai .	sequi	LIICC	. Synthetic
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Lys 65	Asp	Arg	Val	Thr	Met 70	Thr	Arg	Asp	Thr	Ser 75	Thr	Ser	Thr	Val	Tyr 80
Met	Glu	Leu	Ser	Ser 85	Leu	Arg	Ser	Glu	Asp 90	Thr	Ala	Val	Tyr	Tyr 95	Сув
Ala	Arg	Tyr	Phe 100	Phe	Gly	Ser	Ser	Pro 105	Asn	Trp	Tyr	Phe	Asp 110	Val	Trp
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Ala	Val	Leu	Gln 180	Ser	Ser	Gly	Leu	Tyr 185	Ser	Leu	Ser	Ser	Val 190	Val	Thr
Val	Pro	Ser 195	Ser	Asn	Phe	Gly	Thr 200	Gln	Thr	Tyr	Thr	Сув 205	Asn	Val	Asp
His	Lys 210	Pro	Ser	Asn	Thr	Lys 215	Val	Asp	Lys	Thr	Val 220	Glu	Arg	Lys	Сув
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Arg	Trp	Gln	Glu 420	Gly	Asn	Val	Phe	Ser 425	Cys	Ser	Val	Met	His 430	Glu	Ala
Leu	His	Asn 435	His	Tyr	Thr	Gln	Lys 440	Ser	Leu	Ser	Leu	Ser 445	Leu	Gly	Lys

What is claimed is:

- 1. A method for treating a patient having a complementassociated disorder comprising administering to the patient a stable aqueous solution comprising:
 - (a) an anti-C5 antibody at a concentration of about 100 20 mg/mL to about 200 mg/mL, wherein the anti-C5 antibody comprises CDR1, CDR2, and CDR3 domains of a heavy chain comprising the amino acid sequence set forth in SEQ ID NO: 13 and CDR1, CDR2, and CDR3 domains of a light chain comprising the amino acid ²⁵ sequence set forth in SEQ ID NO:12,
 - (b) about 20 mM histidine,
 - (c) about 50 mM serine,
 - (d) about 3% (w/v) sorbitol, and
 - (e) about 1.5% (w/v) mannitol.
- 2. A method for treating a patient having a complementassociated disorder comprising administering to the patient a stable aqueous solution consisting of:
 - (a) an anti-C5 antibody at a concentration of about 100 mg/mL to about 200 mg/mL, wherein the anti-C5 antibody comprises CDR1, CDR2, and CDR3 domains of a heavy chain comprising the amino acid sequence set forth in SEQ ID NO: 13 and CDR1, CDR2, and CDR3 domains of a light chain comprising the amino acid sequence set forth in SEQ ID NO:12,
 - (b) about 20 mM histidine,
 - (c) about 50 mM serine,
 - (d) about 3% (w/v) sorbitol, and
 - (e) about 1.5% (w/v) mannitol.
- 3. A method for treating a patient having a complementassociated disorder comprising administering to the patient a stable aqueous solution comprising:
 - (a) eculizumab,
 - (b) about 20 mM histidine,
 - (c) about 50 mM serine,
 - (d) about 3% (w/v) sorbitol, and
 - (e) about 1.5% (w/v) mannitol.
- **4.** The method of claim **1** or **2**, wherein the anti-C5 anti-body comprises a heavy chain comprising the amino acid sequence set forth in SEQ ID NO:13 and a light chain comprising the amino acid sequence set forth in SEQ ID NO:12.
- 5. The method of claim 1 or 2, wherein the anti-C5 anti-body is eculizumab.

- **6.** The method of any one of claims **1-3**, wherein the pH of the solution is between 6.5 and 7.5.
- 7. The method of claim 6, wherein the pH of the solution is 7.0.
- **8**. The method of claim **1** or **2**, wherein the anti-C5 antibody is at a concentration of 100 mg/mL.
- 9. The method of any one of claims 1-3, wherein the solution is sterile.
- 10. The method of claim 1 or 2, wherein the anti-C5 anti-body remains at least 97% monomeric during storage at 2° C. to 8° C. for at least six months as determined by SEC-HPLC.
- 11. The method of claim 1 or 2, wherein the anti-C5 anti-body remains at least 97% monomeric during storage at 2° C. to 8° C. for at least one year as determined by SEC-HPLC.
- 12. The method of claim 1 or 2, wherein less than 2% of the anti-C5 antibody in the solution is aggregated as determined by SEC-HPLC.
- 13. The method of claim 1 or 2, wherein less than 1% of the anti-C5 antibody in the solution is fragmented as determined by SEC-HPLC.
- 14. The method of claim 1 or 2, wherein during storage at 2° C. to 8° C. for at least six months the anti-C5 antibody retains at least 90% of its C5-binding activity, as compared to a reference anti-C5 antibody corresponding to the anti-C5 antibody prior to storage.
- 15. The method of claim 1 or 2, wherein during storage at 2° C. to 8° C. for at least six months the anti-C5 antibody retains at least 95% of its ability to inhibit hemolysis, as compared to a reference anti-C5 antibody corresponding to the anti-C5 antibody prior to storage.
- **16**. The method of claim **1** or **2**, wherein the anti-C5 antibody is administered by intravenous infusion.
- 17. The method of any one of claims 1-3, wherein the complement-associated disorder is selected from the group consisting of ischemia-reperfusion injury, atypical hemolytic uremic syndrome (aHUS), typical or infectious hemolytic uremic syndrome (tHUS), dense deposit disease (DDD), paroxysmal nocturnal hemoglobinuria (PNH), neuromyelitis optica (NMO), macular degeneration, thrombotic thrombocytopenic purpura (TTP); myasthenia gravis, cold agglutinin disease, Guillain-Barre Syndrome, Degos' disease, graft rejection, sepsis, glomerulonephritis, and thrombotic microangiopathy (TMA).

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